

# SCIENTIFIC REPORTS

OPEN

## Toxic effects of the neem oil (*Azadirachta indica*) formulation on the stink bug predator, *Podisus nigrispinus* (Heteroptera: Pentatomidae)

Received: 11 September 2015

Accepted: 09 May 2016

Published: 06 September 2016

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This research investigated the effects of neem oil on mortality, survival and malformations of the non-target stink bug predator, *Podisus nigrispinus*. Neurotoxic and growth inhibitor insecticides were used to determine the lethal and sublethal effects from neem oil on this predator. Six concentrations of neem oil were topically applied onto nymphs and adults of this predator. The mortality rates of third, fourth, and fifth instar nymphs increased with increasing neem oil concentrations, suggesting low toxicity to *P. nigrispinus* nymphs. Mortality of adults was low, but with sublethal effects of neem products on this predator. The developmental rate of *P. nigrispinus* decreased with increasing neem oil concentrations. Longevity of fourth instar nymphs varied from 3.74 to 3.05 d, fifth instar from 5.94 to 4.07 d and adult from 16.5 and 15.7 d with 0.5 and 50% neem doses. *Podisus nigrispinus* presented malformations and increase with neem oil concentrations. The main malformations occur in wings, scutellum and legs of this predator. The neem oil at high and sub lethal doses cause mortality, inhibits growth and survival and results in anomalies on wings and legs of the non-target predator *P. nigrispinus* indicating that its use associated with biological control should be carefully evaluated.

Botanical insecticides have eco-toxicological advantages compared to traditional synthetic insecticides, because they can have favorable eco-toxicological properties (low human toxicity, rapid degradation and reduced environmental impact), which make them suitable insecticides for organic agriculture<sup>1–3</sup>. Botanical insecticides have secondary metabolites such as alkaloids, amides, chalcones, flavones, phenols, lignans, neolignans or kawapirones; which are important in plant-insect interactions and may be used in integrated pest management (IPM) programs<sup>2,4–6</sup>. They act as repellents with unpleasant odors or irritants, growth regulators and have deterrence on oviposition and feeding, and biocide activity<sup>1,3,6</sup>.

The neem oil, *Azadirachta indica* A. Juss (Sapindales: Meliaceae) have insecticide effect against pests as reported for Coleoptera<sup>7,8</sup>, Diptera<sup>9,10</sup>, Hemiptera<sup>11–13</sup>, and Lepidoptera<sup>14,15</sup>.

Azadirachtin is the main compound of the neem oil with insecticidal activity and can be found in fruits and leaves<sup>16–18</sup>. Other neem oil compounds (tetranortriterpenoids group) are desacetylnimbin, desacetylsalannin, nimbin and salannin<sup>18</sup>.

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Compounds	Stage	<sup>a</sup> LC	<sup>b</sup> EV	<sup>c</sup> CI	<sup>d</sup> X <sup>2</sup>
Neem oil	Nymph	50	14.98 (0.249*)	12.89 (0.214*)–17.43 (0.290*)	14.29
		90	29.14 (0.485*)	25.44 (0.423*)–34.61 (0.576*)	
	Adult	50	41.92 (0.698*)	35.31 (58.84*)–49.75 (0.829*)	27.96
		90	95.81 (1.513*)	82.84 (1.380*)–114.7 (1.911*)	
Pyriproxifen	Nymph	50	5.699	4.269–7.167	28.55
		90	16.85	14.24–20.95	
	Adult	50	7.345	5.589–9.189	28.51
		90	21.66	18.32–26.90	
Imidacloprid	Nymph	50	7.822	6.408–9.404	23.67
		90	18.91	16.27–22.86	
	Adult	50	17.03	13.88–20.72	25.64
		90	44.53	37.90–54.68	

**Table 1. Lethal concentrations of the neem oil extract compared with pyriproxifen (growth inhibitor) and imidacloprid (neurotoxic insecticide) on *Podisus nigrispinus* (Heteroptera: Pentatomidae) nymphs and adults.** Doses of compounds were topically applied. <sup>a</sup>LC<sub>50</sub> and LC<sub>90</sub> concentrations causing 50 and 90% mortality; <sup>b</sup>EV, Estimated value (mg mL<sup>-1</sup>); <sup>c</sup>CI, Confidence interval (mg L<sup>-1</sup>); <sup>d</sup>X<sup>2</sup>, Chi-square value for lethal concentrations and fiducial limits based on a log scale with significance level at  $P < 0.0001$ . \*Estimated value (mg L<sup>-1</sup>) in ppm.

The neem oil is a feeding inhibitor, delaying development and growth, reducing fecundity and fertility, changing behavior and causing anomalies in eggs, larvae and adults of insects or mites<sup>7,19–21</sup>.

Azadirachtin has higher toxicity by ingestion than by contact, which allows it to be used somewhat selectively against phytophagous pests<sup>22,23</sup>. Although natural enemies may ingest contaminated prey, adverse indirect effects may be negligible because 90% of azadirachtin consumed is eliminated from the body of phytophagous insects seven hours after ingestion<sup>24</sup>.

The stink bug predator, *Podisus nigrispinus* Dallas (Heteroptera: Pentatomidae) is a common zoophytophagous insect that is used in the biological control of agriculture and forest pests in the America<sup>25–28</sup>. The potential of *P. nigrispinus* in biological control has been reported for larvae of the defoliators *Alabama argillacea* Hübner, *Anticarsia gemmatalis* Hübner, *Spodoptera exigua* Hübner, and *Trichoplusia ni* Hübner (Lepidoptera: Noctuidae)<sup>28–32</sup> in cotton, tomato, soybean, and *Eucalyptus* plantations<sup>27,33–35</sup>. The compatibility between *P. nigrispinus* and pesticides such as chlorantraniliprole, deltamethrin, imidacloprid, methamidophos, spinosad and thiamethoxam has been successfully demonstrated<sup>36,37</sup>.

Despite the potential benefits of neem oil, this product is mainly sprayed directly onto plant leaves<sup>12,38</sup>, which exposes non-target species to this compound<sup>36,39</sup>. Although these studies show that neem oil can be used in IPM programs, they are limited for a few number of species, additional studies of the potential of direct and indirect effects of neem oil on other natural enemies are yet necessary before it can be recommended for IPM programs.

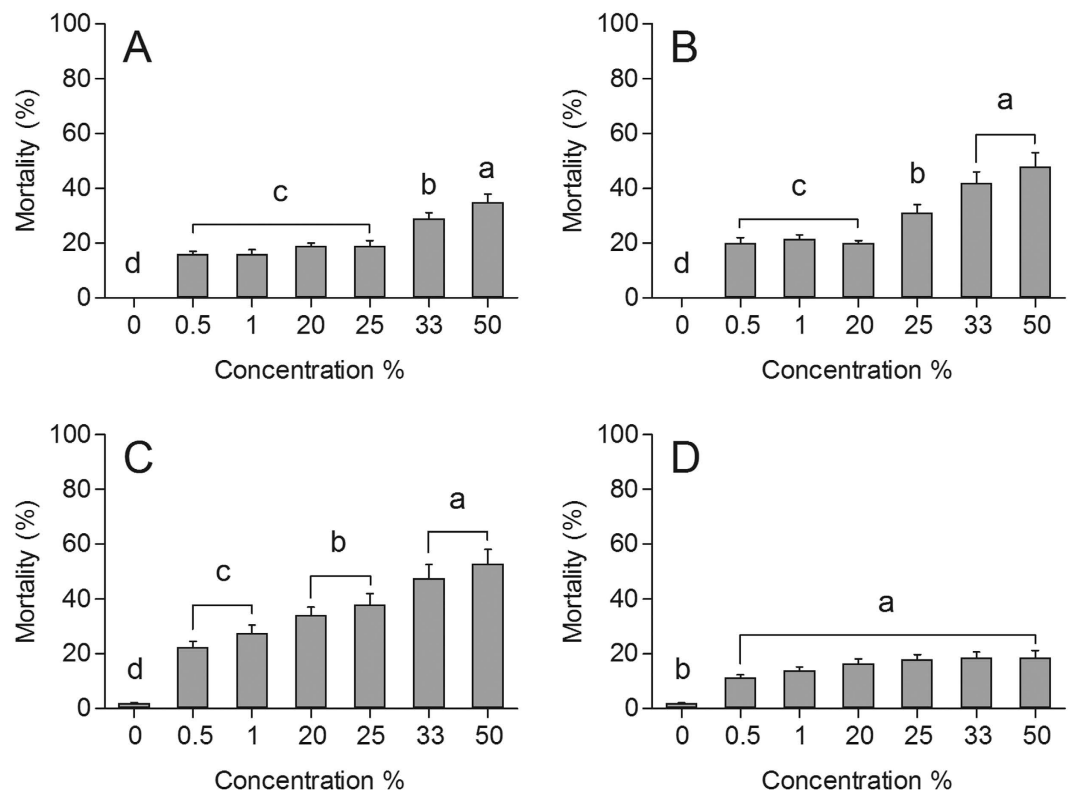
The objective of this study was to evaluate the toxic effects of topical applications of different concentrations of neem oil on the survival and development of the non-target predator *P. nigrispinus*.

## Results

**Comparative toxicity of neem oil and insecticides against *P. nigrispinus*.** The highest mortalities of this predator were obtained with 50 and 100 μL mL<sup>-1</sup> of neem oil, pyriproxifen, and imidacloprid. The two different lethal concentrations levels (LC<sub>50</sub> and LC<sub>90</sub>) (Table 1) of each treatment was estimated by Probit (X<sup>2</sup>;  $P < 0.001$ ). The LC<sub>50</sub> and LC<sub>90</sub> values indicated that pyriproxifen (X<sup>2</sup> = 28.55; df = 5) and imidacloprid (X<sup>2</sup> = 23.67; df = 5) were the most toxic compounds to the *P. nigrispinus* nymphs followed by neem oil (X<sup>2</sup> = 14.29; df = 5). The lethal concentrations (LC<sub>50,90</sub>) value of the pyriproxifen (X<sup>2</sup> = 28.51; df = 5), neem oil (X<sup>2</sup> = 27.96; df = 5), and imidacloprid (X<sup>2</sup> = 25.64; df = 5) showed the toxicity of these compounds to the *P. nigrispinus* adults. Mortality was always <1% in the control.

**Lethal effect of neem oil on *P. nigrispinus*.** The neem oil had lethal effect on the third, fourth and fifth instar nymphs and adults *P. nigrispinus* (Fig. 1). Mortality rates of third instar *P. nigrispinus* nymphs were directly proportional to the neem oil concentrations with values of 15.38, 15.38, 19.23, 19.23, 28.46 and 34.61% (F<sub>1,51</sub> = 3.24;  $P < 0.05$ ) with the concentrations of 0.5, 1, 20, 25, 33 and 50%, respectively (Fig. 1A). The mortality increased in fourth instar to 23.07, 26.92, 34.61, 38.46, 46.15 and 53.84% with increasing neem oil concentrations (F<sub>1,51</sub> = 6.56;  $P < 0.05$ ) (Fig. 1B). The mortality caused on fifth instar differed with the neem oil concentrations (F<sub>1,51</sub> = 11.93;  $P < 0.05$ ) (Fig. 1C). Mortality of *P. nigrispinus* adults was proportional to the neem oil concentration with values of 11.49, 13.91, 16.66, 17.98, 18.51, and 18.66% (F<sub>1,51</sub> = 3.12;  $P < 0.05$ ), in the concentrations of 0.5, 1, 20, 25, 33 and 50%, respectively (Fig. 1A). Mortality never exceeded 2% in the control.

**Development and survival.** The developmental rate of *P. nigrispinus* varied at different concentrations for fourth (F<sub>1,137</sub> = 2.19;  $P < 0.05$ ) and fifth (F<sub>1,137</sub> = 5.10;  $P < 0.05$ ) instars and adults (F<sub>1,137</sub> = 1.45;  $P < 0.05$ ) compared to the control (Table 2). Nymph longevity decreased with increasing concentrations of neem oil (0.5 > 1 > 20 > 25 > 33 > 50%). Fourth instar longevity varied from 3.74 to 3.05 d, the fifth from 5.94 to 4.07 d and that of adult between 16.5 and 15.7 d from 0.5 to 50% of neem oil.



**Figure 1.** Mortality of *Podisus nigrispinus* (Heteroptera: Pentatomidae) nymphs after topical application of the neem oil: third (A) fourth (B) and fifth (C) instar and adults (D). Concentrations means (percent mortality  $\pm$  SEM) differ significantly at  $P < 0.05$  (Tukey's mean separation test).

Neem oil concentrations had a strong effect on the survival of nymph to adult *P. nigrispinus* from the fourth instar (Table 3). This survival varied with concentrations of this oil for fourth ( $F_{1,137} = 5.24$ ;  $P < 0.05$ ) and fifth ( $F_{1,137} = 7.53$ ;  $P < 0.05$ ) instar nymphs and adults ( $F_{1,137} = 3.47$ ;  $P < 0.05$ ). In general, survival rates declined at 20, 25, 33 and 50% neem oil concentrations.

**Malformations caused for the neem oil.** Nymphs and adults *P. nigrispinus* presented malformations ( $X^2$ ,  $P < 0.0001$ ) with its number increasing proportionally with the neem concentrations (0.5 > 1 > 20 > 25 > 33 > 50%) (Table 3). Malformations in the fifth instar nymphs varied from 1.9 to 21.6% with 0.5 to 50% concentrations of neem oil, whereas it ranged from 2.5 to 30.8% in adults.

Irreversible malformations occurred in *P. nigrispinus* adults with higher severity as the neem concentrations increased. The main malformations were the hemelytra size reduction (Fig. 2A), low number of veins and reduced membranous area of wings (Fig. 2B), asymmetric scutellum (Fig. 2C), and extension and folding of the legs (Fig. 2D).

## Discussion

Chemical and biological pest control procedures need high selectivity for use in IPM programs, for instance, broad-spectrum insecticides are not suitable. Selectivity of insecticides can be achieved with environmental measures, by minimizing exposure to predators, or physiologically, with insecticides that are more toxic to pests than to predators<sup>6,11,40</sup>.

The toxicity profiles for the neem oil compared to two insecticides as positive control on the stink bug predator, *P. nigrispinus* were determined from the bioassays. The neem oil, pyriproxifen, and imidacloprid caused substantial mortality of *P. nigrispinus* nymphs and adults under laboratory conditions. The susceptibility of the Hemiptera may vary with exposure in the different concentrations of neem oil and insecticides<sup>41–43</sup>. The  $LC_{50}$  and  $LC_{90}$  values indicated that lethality of neem oil and imidacloprid were lower on *P. nigrispinus* than pyriproxifen with the concentrations evaluated. However, the lethality of neem oil is confirmed on *N. nigrispinus* depending on the concentration applied and can be compared to neurotoxic insecticides and growth inhibitors, as a potent natural insecticide. Mortality of *P. nigrispinus* was not restricted to the third instar nymphs. Lethal effect on adult stage is also apparent even when exposed during the first hours. This is a common toxic effect of pyriproxifen and imidacloprid, and also was observed with the neem oil. As would be expected for a juvenile hormone (JH) mimic as the neem oil and pyriproxifen, which also leads to adult malformation and reproductive impairment of individuals emerged<sup>41,43</sup>.

Here, the term selectivity refers to the toxicological selectivity of neem oil on non-target organism as *P. nigrispinus*. Our results showed that neem oil at different concentrations (i.e., 0.5, 1, 20, 25, 33 and 50%) affected the nymphs

Neem oil concentrations (%)	Fourth instar	Fifth instar	Adult
	Development (days)*		
Control	3.48 ± 0.1b	5.00 ± 0.2a	16.2 ± 0.5a
0.5	3.74 ± 0.2a	5.94 ± 0.3b	16.5 ± 0.1a
1	3.42 ± 0.2b	5.11 ± 0.2c	16.3 ± 0.1a
20	3.70 ± 0.2a	4.84 ± 0.2d	16.3 ± 0.3a
25	3.23 ± 0.3c	4.62 ± 0.2d	16.1 ± 0.4a
33	3.05 ± 0.2d	4.62 ± 0.2d	15.6 ± 0.3b
50	3.05 ± 0.2d	4.07 ± 0.2e	15.7 ± 0.7b
F <sub>1,137</sub>	2.19	5.10	1.45
P	0.0031	0.0001	0.0167
	Survival (%)*		
Control	94.2 ± 0.5a	98.8 ± 0.1a	99.7 ± 0.3a
0.5	93.4 ± 0.7a	99.4 ± 0.5a	96.7 ± 0.7a
1	94.9 ± 0.1a	95.8 ± 0.4a	96.1 ± 0.5a
20	83.5 ± 0.2b	84.8 ± 0.1b	96.3 ± 0.6a
25	72.3 ± 0.3c	76.4 ± 0.2c	91.1 ± 0.1b
33	60.3 ± 0.2d	64.62 ± 0.5d	85.6 ± 0.3b
50	55.8 ± 0.2e	54.3 ± 0.7e	85.8 ± 0.3b
F <sub>1,137</sub>	5.24	7.53	3.47
P	0.0001	0.0001	0.0019

**Table 2. Development (days) and survival (%) (mean ± SE) of fourth and fifth instar nymphs and adults *Podisus nigrispinus* (Heteroptera: Pentatomidae) submitted to topical application of different concentrations (%) of neem oil (25 ± 4 °C, 70 ± 5% RH and 12 h photophase). \*Means followed by the same letter within each column are not different according to the Tukey's test (P < 0.05).**

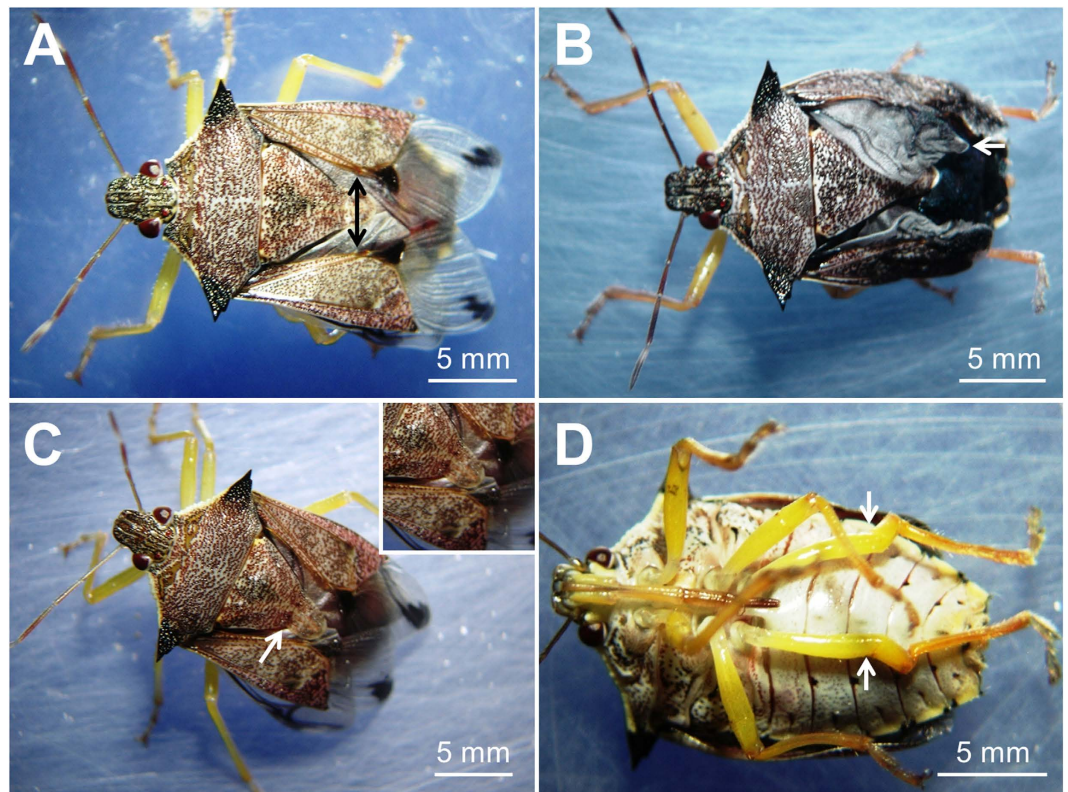
Stage	Concentration (%)	n	Normal	Malformation	X <sup>2</sup>
Nymph	Control	50	100	0	7.407
	0.5	40	98.1	1.9	
	1	37	94.7	5.3	
	20	35	92.3	7.7	
	25	31	88.4	11.6	
	33	27	80.7	19.3	
	50	26	78.4	21.6	
Adult	Control	50	100	0	50.52
	0.5	40	97.5	2.5	
	1	37	97.2	2.8	
	20	35	82.8	17.2	
	25	31	77.4	32.6	
	33	27	74.0	26.0	
	50	26	69.2	30.8	

**Table 3. Malformation (%) in fifth instar nymphs and adults *Podisus nigrispinus* (Heteroptera: Pentatomidae) after topical application of the neem oil extracts in the third instar (25 ± 4 °C, 70 ± 5% RH and 12 h photophase).**

and adults *P. nigrispinus*. Survival was higher in adults (91.4%) than in fourth (72.3%) and fifth (76.4%) instar nymphs, indicating that *P. nigrispinus* was more susceptible in the immature stages. Previous studies on its prey, *Anticarsia gemmatilis* Hübner (Lepidoptera: Noctuidae), show 18.5% larval survival at 25% concentration of neem oil and displayed abnormalities after the last larval moult<sup>44</sup>. Sub-lethal effects on this insect may greatly hinder the survival, and fitness of nymphs with malformations in the adults of this predator and its prey.

The concentrations of neem oil caused mortality on the third, fourth, and fifth instar nymphs. On the other hand, neem as a botanical pesticide has many excellent attributes including its broad-spectrum in insect growth regulatory effects, systemic action in some plants, minimal effects on natural enemies and pollinators, rapid degradation in the environment, and no toxicity to vertebrates<sup>2,45,46</sup>. The increased mortality of third, fourth and fifth instar *P. nigrispinus* nymphs may be a sublethal effects of the neem oil on this predator. *Spodoptera littoralis* Boisduval (Lepidoptera: Noctuidae) larvae died before the pupa stage after application of 0.5 ppm azadirachtin, probably due to feeding inhibition<sup>47</sup>. The stink bug predator, *Macrolophus caliginosus* Wagner (Hemiptera: Miridae) showed similar chronic toxicity in all instars with different neem oils<sup>11</sup>. Oil from neem seeds containing 0.05 and 0.1 g/L of azadirachtin reduces population growth of the pest aphid *Myzus persicae* Sulzer (Hemiptera:





**Figure 2.** Malformations in adults *Podisus nigrispinus* (Heteroptera: Pentatomidae) after topical application of neem oil on its nymphs. (A) Defective hemelytra (arrow), (B) low number of veins and membranous area of wings (arrow), (C) asymmetric scutellum (arrow) and, (D) extension and folding the legs (arrows).

Aphididae) and had sublethal effects on the predatory ladybird beetle *Eriopis connexa* Germar (Coleoptera: Coccinellidae)<sup>8</sup>.

The longevity of fourth and fifth instar *P. nigrispinus* nymphs after exposure to 33 and 50% neem oil suggests susceptibility of this predator to neem compounds. Increase duration of the nymph stage may affect reproductive fitness, because females with shorter lifespan lay higher numbers of eggs<sup>29</sup>. Thus, studies on collateral effects of neem formulations are necessary to detect their potential impact on non-target insects.

The survival <25% of fourth and fifth instar *P. nigrispinus* nymphs after treatment with concentrations of neem oil, suggests a degree of tolerance. Concentrations of neem oil between 0.1 and 10% cause mortality in various hemipteran pests and predators such as *Bemisia argentifolii* Bellows & Perring (Aphididae)<sup>21</sup>, *Clavigralla scutellaris* Westwood (Coreidae)<sup>19</sup>, *Macrolophus caliginosus* Wagner (Miridae)<sup>11</sup>, *Myzus persicae* Sulzer (Aphididae)<sup>8</sup>, *Nezara viridula* Linnaeus (Pentatomidae)<sup>13</sup>, *Nilaparvata lugens* Stal (Delphacidae)<sup>47</sup>, and *Picromerus bidens* Linnaeus (Pentatomidae)<sup>49</sup>. The bioneem is recommended in Brazil to control phytophagous insects at concentrations of 0.5 and 1%. Therefore, this product may be used against pests in the presence of *P. nigrispinus* if used in concentrations lower than 25%.

Several malformations, possibly related to defective molting, were observed from 1 to 50% of neem oil concentrations. Azadirachtin was established as an insect growth regulator with a novel mode of action. The basis for its mode of action was known to involve the neurosecretory–neuroendocrine pathway and perhaps other sites including cell cycle<sup>46</sup>. Studies on the insect growth regulatory mode of azadirachtin action in *Calliphora vicina* Robineau-Desvoidy (Diptera: Calliphoridae), *Manduca sexta* Linnaeus (Lepidoptera: Sphingidae), and *Oncopeltus fasciatus* Dallas (Hemiptera: Lygaeidae), show that JH biosynthesis and catabolism were affected by azadirachtin improving the insect growth regulatory effects<sup>50–52</sup>. Azadirachtin induces supernumerary molts, lack of black pigment and malformations<sup>53,54</sup>. Malformations on the legs, thorax and wings of *P. nigrispinus* adults exposed to neem oil in the third instar were similar to those found in the coccinellids *Chilochorus bipustulatus* and *Phrosychnus anchorago*<sup>55</sup> and the lacewing *M. caliginosus*<sup>11</sup>. Nymphs of the phytophagous stink bug *Nezara viridula* Linnaeus (Hemiptera: Pentatomidae) exposed to commercial neem oils have malformations in the antennae, ocelli, tarsi, odoriferous glands, scutellum, genitalia and mouth parts<sup>11,56–58</sup>.

Ideally, phytochemical insecticides should be toxic to pests with low or no impact on predators. In this study, topical applications of neem oil showed low acute toxicity for *P. nigrispinus* nymphs, especially at concentrations <25%. However, sublethal effects, such as increased mortality rates of fourth and fifth instars, longer instar duration and body malformations indicate that the use of neem oil associated with biological control using the predator *P. nigrispinus* should be previously evaluated for the use in IPM.

## Methods

**Maintenance of Insect Culture.** Nymphs and adults *P. nigrispinus* were obtained from mass rearing of the Laboratório de Controle Biológico do Instituto de Biologia Aplicada à Agropecuária (BIOAGRO, Universidade Federal de Viçosa, Minas Gerais, Brazil). They were maintained at  $25 \pm 2^\circ\text{C}$  at  $75 \pm 5\%$  relative humidity and 12-h photophase. The insects were kept in wooden cages ( $30 \times 30 \times 30$  cm) coated with nylon and glass, and received *ad libitum* *Tenebrio molitor* (L.) pupae (Coleoptera: Tenebrionidae), *Eucalyptus grandis* (W. Hill ex. Maiden) leaves and water<sup>27</sup>.

**Neem oil.** Vegetable Bioneem is an organic Brazilian product composed of oil obtained from cold extraction of neem seeds, without the addition of solvents or pesticides, and certified by Ecocert Brazil as a natural insecticide with repellent properties. This product was developed with azadirachtin ( $25\text{ g L}^{-1}$ ) and others neem oil isomers concentrations for tropical regions. Ecocert Brazil is accredited by the Ministry of Agriculture, Livestock and Supply of Brazil and by international organizations according to ISO Guide 34<sup>59</sup>. This provides Brazilian producers with licenses, and ensures unrestricted access to major world organic product markets.

**Comparative toxicity of neem oil and insecticides.** Neem oil with 1800 to 2200 ppm of azadirachtin (Bioneem, Tecnologia Consultoria Indústria Comércio, Brazil) was compared with two different insecticides used as positive control in this study. The following commercial insecticides were tested at their maximum label rates: pyriproxifen (Tiger EC, Sumitomo Chemical Corporation, Brazil),  $100\text{ g L}^{-1}$  and imidacloprid (Evidence WG, Bayer, Germany),  $700\text{ g L}^{-1}$ . These insecticides were diluted in 1 L water to produce a stock solution by adjusting  $100\text{ g L}^{-1}$  per insecticide and to obtain the required concentrations. Insecticide efficacy was determined by calculating the lethal concentrations ( $\text{LC}_{50}$  and  $\text{LC}_{90}$ ) values under laboratory conditions for each formulation. Six concentrations of vegetable bioneem, pyriproxifen, imidacloprid besides the control (distilled water) were adjusted in 1 mL stock solution (treatments and distilled water): 1.56, 3.12, 6.25, 12.5, 25, and 50%. For each treatment, aliquots were taken from the stock solution and mixed with distilled water in 5 mL glass vials. Different concentrations of the treatments were applied in  $1\ \mu\text{L}$  of topical solution in the body of each individual of *P. nigrispinus*. Fifty third instar nymphs and fifty adults were used per concentration and were placed individually in glass vials ( $2 \times 10$  cm) with a cotton lid and maintained in the dark. The number of dead insects in each vial was counted after neem and insecticides exposure at intervals of 6 h over 6 days.

**Mortality test.** Six concentrations of neem oil besides the control (liquid glycerin), were adjusted in  $10\ \mu\text{L}$  stock solution (neem oil and liquid glycerin): 0.5, 1, 20, 25, 33 and 50% aliquots were taken from the stock solution and mixed with liquid glycerine in 5 mL glass vials. Different neem oil concentrations were topically applied in  $1\ \mu\text{L}$  solution onto insect scutellum with an analytical  $10\ \mu\text{L}$  syringe. The glycerin was chosen due to applied solution adherence in the *P. nigrispinus* scutellum in order to ensure the absorption of the pesticide. Fifty-two individuals of third, fourth, fifth instar nymphs and adults (1:1 males and females proportion) of *P. nigrispinus* were used per concentration and individually placed in plastic containers ( $15 \times 10$  cm) with a perforated lid, fed on *T. molitor* pupae, *E. grandis* leaves, and water under laboratory conditions. First and second instar nymphs were not tested because they are too small to the volume of insecticide applied. The number of dead insects per concentration was daily counted after neem oil exposure until adult emergence.

## Development

The development of *P. nigrispinus* nymphs and adults was daily registered. To monitor nymph and adult development, 1,200 third-instar nymphs were placed individually in Petri dishes ( $90 \times 15$  mm) with moistened cotton ball and fed on *T. molitor* pupae. Nymphs were maintained at  $25 \pm 2^\circ\text{C}$  at  $75 \pm 5\%$  RH and 12-h photophase until, fourth and fifth instars and adult emergence. Survival and duration of fourth and fifth instar nymphs, and adults, after emergence, were recorded. Longevity and survival were determined from third instar nymph with the six neem oil concentrations.

**Malformations.** The number of malformed nymphs and adults was quantified per concentration after topical application of the neem oil on third instar nymphs. Severe morphological abnormalities of the *P. nigrispinus* adults were photographed.

**Statistics.** The  $\text{LC}_{50}$  and  $\text{LC}_{90}$ , and their confidence limits were determined by logistic regression based on the concentration probit-mortality<sup>60</sup>, with the program XLSTAT-PRO v.7.5 for Windows<sup>61</sup>. Mortality, development time and survival data of nymph and adult were analyzed by one-way ANOVA. Mortality variables were summarized in percentages and the data transformed to arcsine square root. Tukey's Honestly Significant Difference test (HSD) was used for comparing the means at the 5% significance level (PROC ANOVA) using SAS v9.0<sup>62</sup>. The malformations of fifth instar nymphs and adults were compared by frequency of responses with chi-square test using SPSS v17.0 for Windows<sup>63</sup>.

## References

- Bourguet, D., Genissel, A. & Raymond M. Insecticide resistance and dominance levels. *J Econ Entomol* **93**, 1588–1595 (2000).
- Isman, M. B. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annu Rev Entomol* **51**, 45–66 (2006).
- Chermenskaya, T. D., Stepanycheva, E. A., Shchenikova, A. V. & Chakaeva, A. S. Insecto acaricidal and deterrent activities of extracts of Kyrgyzstan plants against three agricultural pests. *Ind Crop Prod* **32**, 157–163 (2010).
- Parmar, V. S. *et al.* Phytochemistry of the genus *Piper*. *Phytochemistry* **46**, 597–673 (1997).
- Abou-Fakhr, H., Zournajian, E. M. H. & Talhouk, S. Efficacy of extracts of *Melia azedarach* L. callus, leaves and fruits against adults of the sweet potato whitefly *Bemisia tabaci*. *J Appl Entomol* **125**, 483–488 (2001).

6. Martínez, L. C., Plata-Rueda, A., Zanoncio, J. C. & Serrão, J. E. Bioactivity of six plant extracts on adults of *Demotispá neivai* (Coleoptera: Chrysomelidae). *J Insect Sci* **15**, 34 (2015).
7. Ventura, M. U. & Ito, M. Antifeedant activity of *Melia azedarach* (L.) extracts to *Diabrotica speciosa* (Genn.) (Coleoptera: Chrysomelidae) beetles. *Brazilian Arch Biol Tech* **43**, 215–219 (2000).
8. Venzon, M., Rosado, M. C., Pallini, A., Fialho, A. & Pereira, C. J. Toxicidade letal e subletal do nim sobre o pulgão-verde e seu predador *Eriopsis connexa*. *Pesqui Agropecu Bras* **42**, 627–631 (2007).
9. Souza, A. P. & Vendramim, J. D. Efeito translaminar, sistêmico e de contato de extrato aquoso de sementes de nim sobre *Bemisia tabaci* (Genn.) biótipo B em tomateiro. *Neotrop Entomol* **34**, 83–87 (2005).
10. Yasmin, N., Khan, M. F., Channa, M. S. & Zeeshan, A. Effects of a neem sample on protein patterns of *Bactrocera cucurbitae*. *Turk J Zool* **32**, 1–5 (2008).
11. Tedeschi, R., Alma, A. & Tavella, L. Side-effects of three neem (*Azadirachta indica* A. Juss) products on the predator *Macrolophus caliginosus* Wagner (Het., Miridae). *J Appl Entomol* **125**, 397–402 (2001).
12. Kumar, P., Poehling, H. M. & Borgemeister, C. Effects of different application methods of azadirachtin against sweetpotato whitefly *Bemisia tabaci* Gennadius (Hom., Aleyrodidae) on tomato plants. *J Appl Entomol* **129**, 489–497 (2005).
13. Singha, A., Thareja, V. & Singla, A. K. Application of neem seed kernel extrats result in mouthpart deformites and subsequent mortality in *Nezara viricula* (L.) (hem: Pentatomidae). *J Appl Entomol* **131**, 197–201 (2007).
14. Viana, P. A. & Prates, H. T. Desenvolvimento e mortalidade larval de *Spodoptera frugiperda* em folhas de milho tratadas com extrato aquoso de folhas de *Azadirachta indica*. *Bragantia* **62**, 69–74 (2003).
15. Charbonneau, C., Côté, R. & Charpentier, G. Effects of azadirachtin and of simpler epoxy-alcohols on survival and behaviour of *Galleria mellonella* (Lepidoptera). *J Appl Entomol* **131**, 447–452 (2007).
16. Bruce, Y. A., Gounou, S., Chabi-Olaye, A., Smith, H. & Schulthess, F. The effect of neem (*Azadirachta indica* A. Juss) oil on oviposition, development and reproductive potentials of *Sesamia calamistis* Hampson (Lepidoptera: Noctuidae) and *Eldana saccharina* Walker (Lepidoptera: Pyralidae). *Agric Forest Entomol* **6**, 223–232 (2004).
17. Medina, P., Budia, F., Del Estal, P. & Vinuela, E. Influence of azadirachtin, a botanical insecticide, on *Chrysoperla carnea* (Stephens) reproduction: toxicity and ultrastructural approach. *J Econ Entomol* **97**, 43–50 (2004).
18. Silva, J. C. T., Jham, G. N., Oliveira, R. D. L. & Brown, L. Purification of the seven tetranortriterpenoids in neem (*Azadirachta indica*) seed by counter-current chromatography sequentially followed by isocratic preparative reversed-phase high-performance liquid chromatography. *J Chromatogr* **1151**, 203–210 (2007).
19. Mitcheli, P. L., Gupta, R., Singh, A. K. & Kumar, P. Behavioural and development effects of neem extracts on *Clavigralla scutellaris* (Hemiptera: Heteroptera: Coreidae) and its egg parasitoid, *Gryon fulviventre* (Hymenoptera: Scelionidae). *J Econ Entomol* **97**, 916–923 (2004).
20. Mourão, S. A. *et al.* Seletividade de extratos de Nim (*Azadirachta indica* A. Juss) ao ácaro predador *Iphiseiodes zuluagai* (Denmark & Mina) (Acari: Phytoseiidae). *Neotrop Entomol* **33**, 613–617 (2004).
21. Masood, K. K., ur-Rashid, M., Syed, A., Hussain, S. & Islam, T. Comparative effect of neem (*Azadirachta indica* A. Juss) oil, neem seed water extract and baythroid against whitefly, jassids and thrips on cotton. *Pak Entomol* **28**, 31–37 (2006).
22. Akol, A. M., Sithanatham, S., Njagi, P. G. N., Varela, A. & Mueke, J. M. Relative safety of sprays of two neem insecticides to *Diadegma mollipla* (Holmgren), a parasitoid of the diamondback moth: effects on adult longevity and foraging behavior. *Crop Protect* **21**, 853–859 (2002).
23. Gaspari, M., Lykouressis, D., Perdikis, D. & Polissiou, D. Nettle extract effects on the aphid *Myzus persicae* and its natural enemy, the predator *Macrolophus pygmaeus* (Hem: Miridae). *J Appl Entomol* **131**, 652–657 (2007).
24. Schmutterer, H. *The neem tree* (VHC Weinheim, 1995).
25. Cohen, A. C. Feeding adaptations of some predaceous Heteroptera. *Ann Entomol Soc Am* **83**, 1215–1223 (1990).
26. Medeiros, R. S., Ramalho, F. S., Lemos, W. P. & Zanoncio, J. C. Age-dependent fecundity and life-fertility tables for *Podisus nigrispinus* (Dallas) (Heteroptera: Pentatomidae). *J Appl Entomol* **124**, 319–324 (2000).
27. Lemos, W. P., Medeiros, R. S., Ramalho, F. S. & Zanoncio, J. C. Effects of plant feeding on the development, survival and reproduction of *Podisus nigrispinus* (Dallas) (Heteroptera: Pentatomidae). *Int J Pest Manage* **47**, 89–93 (2001).
28. Mohaghegh, J., De Clercq, P. & Tirry, L. Functional response of the predators *Podisus maculiventris* (Say) and *Podisus nigrispinus* (Dallas) (Heteroptera: Pentatomidae) to the beet armyworm, *Spodoptera exigua* (Hübner) (Lepidoptera: Noctuidae): effect of temperature. *J Appl Entomol* **125**, 131–134 (2001).
29. Medeiros, R. S., Ramalho, F. S., Zanoncio, J. C. & Serrão, J. E. Effect of temperature on life table parameters of *Podisus nigrispinus* (Het., Pentatomidae) fed with *Alabama argillacea* (Lep., Noctuidae) larvae. *J Appl Entomol* **127**, 209–213 (2003).
30. Martínez, L. C. *et al.* Stink bug predator kills prey with salivary non-proteinaceous compounds. *Insect Biochem Mol Biol* **68**, 71–78 (2016).
31. Neves, R. C. S., Torres, J. B. & Zanoncio, J. C. Production and storage of mealworm beetle as prey for predatory stinkbug. *Biocontrol Sci Techn* **20**, 1013–1025 (2010).
32. De Bortoli, S. A., Otuka, A. K., Vacari, A. M., Martins, M. I. E. G. & Volpe, H. L. X. Comparative biology and production costs of *Podisus nigrispinus* (Hemiptera: Pentatomidae) when fed different types of prey. *Biol Control* **58**, 127–132 (2011).
33. Zanoncio, J. C., Alves, J. B., Zanoncio, T. V. & García, J. L. Hemipterous predators of eucalypt defoliator caterpillars. *Forest Ecol Manag* **65**, 65–73 (1994).
34. Matos Neto, F. C., Zanoncio, J. C., Cruz, I. & Torres, J. B. Nymphal development of *Podisus nigrispinus* (Heteroptera: Pentatomidae) preying on larvae of *Anticarsia gemmatalis* (Lepidoptera: Noctuidae) fed with resistant and susceptible soybeans. *Rev Bras Entomol* **46**, 237–241 (2002).
35. Oliveira, J. E. M., Torres, J. B., Moreira, A. & Barros, R. Efeito das plantas do algodoeiro e do tomateiro, como complemento alimentar, no desenvolvimento e na reprodução do predador *Podisus nigrispinus* (Dallas) (Heteroptera: Pentatomidae). *Neotrop Entomol* **31**, 101–108 (2002).
36. Torres, J. B. & Ruberson, J. R. Toxicity of thiamethoxam and imidacloprid to *Podisus nigrispinus* (Dallas) (Heteroptera: Pentatomidae) nymphs associated to aphid and whitefly control in cotton. *Neotrop Entomol* **33**, 99–106 (2004).
37. De Castro, A. A. *et al.* Survival and behavior of the insecticide-exposed predators *Podisus nigrispinus* and *Supputius cincticeps* (Heteroptera: Pentatomidae). *Chemosphere* **93**, 1043–1050 (2013).
38. Calvo, D. & Molina, J. M. Effects of a commercial neem (*Azadirachta indica*) extract on *Streblote panda* larvae. *Phytoparasitica* **31**, 365–370 (2003).
39. Batista Filho, A. *et al.* Manejo integrado de pragas de soja: impacto de inseticidas sobre inimigos naturais. *Arq Inst Biol* **70**, 61–67 (2003).
40. Pedigo, L. P. *Entomology and pest management*. New York: Macmillan. 646p (1988).
41. Schmutterer, H. Properties and potential of natural pesticides from the neem tree, *Azadirachta indica*. *Annu Rev Entomol* **35**, 271–297 (1990).
42. Matsuda, K. *et al.* Neonicotinoids: insecticides acting on insect nicotinic acetylcholine receptors. *Trends Pharmacol Sci* **22**, 573–580 (2001).
43. Ishaaya, I., Barazani, A., Kantsedalov, S. & Horowitz, A. R. Insecticides with novel modes of action: Mechanism, selectivity and cross-resistance. *Entomol Res* **37**, 148–152 (2007).



44. Mourão, S. A. *et al.* Mortality of *Anticarsia gemmatalis* (Lepidoptera: Noctuidae) caterpillars post exposure to a commercial neem (*Azadirachta indica*, Meliaceae) oil formulation. *Fla Entomol* **97**, 555–561 (2014).
45. Mordue, A. J. L. & Nisbet, A. J. Azadirachtin from the neem tree *Azadirachta indica*: its action against insects. *An Soc Entomol Bras* **29**, 615–632 (2000).
46. Gilbert, L. I. & Gill, S. S. *Insect control biological and synthetic agents* (Academic Press-Elsevier, 2010).
47. Martinez, S. S. & van Endem, H. F. Growth disruption, abnormalities and mortality of *Spodoptera littoralis* caused by azadirachtin. *Neotrop Entomol* **30**, 113–125 (2001).
48. Nathan, S. S. *et al.* Effect of azadirachtin on acetylcholinesterase (AChE) activity and histology of the brown planthopper *Nilaparvata lugens* (Stal). *Ecotox Environ Safe* **70**, 244–250 (2008).
49. Mahdian, K., Tirry, L. & De Clercq, P. Functional response of *Picromerus bidens*: Effect of host plant. *J Appl Entomol* **131**, 160–164 (2007).
50. Schlüter, U. Occurrence of weight-gain reduction and inhibition of metamorphosis and storage protein information in last larval instars of the Mexican bean beetle, *Epilachna varivestis*, after injection of azadirachtin. *Entomol Exp Appl* **39**, 191–195 (1985).
51. Bidmon, H. J. Ultrastructural changes of the prothorax glands of untreated and with azadirachtin treated *Manduca sexta* larvae (Lepidoptera, Sphingidae). *Entomol Gen* **12**, 1–17 (1986).
52. Dorn, A., Rademacher, J. M. & Sehn, E. Effects of azadirachtin on the moulting cycle, endocrine system and ovaries in last-instar larvae of the milkweed bug, *Oncopeltus fasciatus*. *J Insect Physiol* **32**, 321–328 (1986).
53. Schlüter, U., Bidmon, H. J. & Grewe, S. Azadirachtin affects growth and endocrine events in larvae of the tobacco hornworm, *Manduca sexta*. *J Insect Physiol* **31**, 773–777 (1985).
54. Beckage, N. E., Metcalf, J. S., Nielsen, B. D. & Nesbit, D. J. Disruptive effects of azadirachtin on development of *Cotesia congregata* in host tobacco hornworm larvae. *Arch Insect Biochem Physiol* **9**, 47–65 (1988).
55. Peveling, R. & Ely, S. O. Side-effects of botanical insecticides derived from Meliaceae on coccinellid predators of the date palm scale. *Crop Prot* **25**, 1253–1258 (2006).
56. Abudulai, M., Shepard, B. M. & Mitchell, P. L. Antifeedant and toxic effects of a neem (*Azadirachta indica* A. Juss) based formulation Neemix against *Nezara viridula* (L.) (Hemiptera: Pentatomidae). *J Entomol Sci* **38**, 398–408 (2003).
57. Durmusoglu, E., Karsavuran, Y., Ozgen, I. & Guncan, A. Effects of two different neem products on different stages of *Nezara viridula* (L.) (Heteroptera, Pentatomidae). *J Pestic Sci* **76**, 151–154 (2003).
58. Riba, M., Marti, J. & Sans, A. Influence of azadirachtin on development and reproduction of *Nezara viridula* L. (Het., Pentatomidae). *J Appl Entomol* **127**, 37–41 (2003).
59. Accustandard. *Analytical Chemical Reference Standards* (Pesticide Reference Guide, 2010).
60. Finney, D. J. *Probit Analysis* (Cambridge University press, 1971).
61. XLSTAT. XLSTAT for Microsoft Excel. XLSTAT Addinsoft, Paris, France. URL: <http://www.xlstat.com/fr> (2004).
62. SAS Institute. *SAS The Statistical Analysis System*. SAS Institute, Cary, NC, USA. URL: <http://www.sas.com> (2002).
63. SPSS. IBM SPSS Statistics. Chicago, IL, USA URL: <http://www.spss.com> (2007).

## Acknowledgements

This research was supported by “Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq)”, and “Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG)”. To Asia Science for correction and English editing this manuscript.

## Author Contributions

S.A.M., L.C.M. and C.F.W. performed experiments, and analyzed the data; F.S.R. and M.A.S. analyzed the data; A.P.-R., L.C.M., J.E.S. and J.C.Z. designed experiments and wrote the manuscript.

## Additional Information

**Competing financial interests:** The authors declare no competing financial interests.

**How to cite this article:** Mourão, S. A. *et al.* Toxic effects of the neem oil (*Azadirachta indica*) formulation on the stink bug predator, *Podisus nigrispinus* (Heteroptera: Pentatomidae). *Sci. Rep.* **6**, 30261; doi: 10.1038/srep30261 (2016).



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