Bioactive compounds of juices from two Brazilian grape cultivars

Juliana Kelly da Silva, a Cinthia Baú Betim Cazarin, a Luiz Claudio Correa, b Ângela Giovana Batista, a Cibele Priscila Busch Furlan, a Aline Camarão Telles Biasoto, b Giuliano Elias Pereira, c Adriano Costa de Camargo d and Mário Roberto Maróstica Junior a*

Abstract

BACKGROUND: Grape juice consumption may prevent several chronic diseases owing to the presence of phenolic compounds, which have an important role in the reduction of oxidative stress. This study investigated the polyphenol content and antioxidant activities of grape juices from two cultivars: BRS-Cora and Isabella. Total polyphenol content (TPC), anthocyanins, antioxidant capacity (oxygen radical absorbance capacity, ferric reducing antioxidant power and 1,1-diphenyl-2-picrylhydrazyl), and phenolic profile (high-performance liquid chromatography with diode array and fluorescence detection – HPLC-DAD-FLD) were determined.

RESULTS: BRS-Cora grape juice showed higher concentrations of total polyphenols and anthocyanins, as well as higher antioxidant potential, than those of Isabella grape juice. A significant positive correlation was found in TPC or anthocyanin contents when correlated with the remaining antioxidant assays. In addition, HPLC-DAD-FLD showed a higher total phenolic content in BRS-Cora grape juice compared to Isabella.

CONCLUSION: The present results show BRS-Cora as a promising cultivar for grape juice production with an improved functional potential.

INTRODUCTION

Grapes, grape juice and wine have shown both in vitro and in vivo potential to prevent several chronic diseases. The benefits stem from their high polyphenol content, which are secondary metabolites produced by plants as a defense mechanism against environmental stress.1–4 The protective effects against health problems such as cancer and cardiovascular diseases may be attributed to their antioxidant activity and antiproliferative properties.5 Furthermore, interest in polyphenols has increased owing to their potential activity as antitoxic and cytoprotective molecules with a modulatory action in inflammatory processes and neurodegenerative diseases.1–3,6,7

The search for natural sources of bioactive compounds is a strong trend nowadays. At the same time, an increase in the consumption of grapes and their products was noted, focusing not only on nutrition but also on their beneficial health effects. Grapes are sources of polyphenols belonging to different subclasses such as monomeric flavan-3-ols, proanthocyanidins, flavonoids, anthocyanins, phenolic acids and the stilbene resveratrol.8–11 Several factors such as geographic origin, cultivar and environmental conditions may influence polyphenol concentration.9,12–15 Different agronomic strategies have been employed to develop new crops, focusing on sensory quality, field production yield and resistance to pests and diseases,16,17 which, in turn, may influence the biosynthesis of phenolic compounds.12

Red wine is the most famous grape product, which has been correlated with the ‘French paradox’, in which low coronary disease death rates have been observed in France, despite the high amount of dietary cholesterol and saturated fatty acids. However, several studies have also reported improvements on health aspects with grape juice intake. Supplementation with Concord grape juice (8 mL kg⁻¹ body weight d⁻¹) for 14 days positively altered the endothelial function in adults with coronary artery...
Grapes were sanitized with hypochlorite solution (100mgL⁻¹) and then rinsed with water. Fruits were then processed by the steam extraction method for 60min at 75–85°C and the mixture was kept in the dark at room temperature for 2h. The absorbance was read at 725nm using a spectrophotometer (Synergy HT, Biotek, Winooski, VT, USA). Gallic acid was used to prepare a standard curve and the results were expressed as gallic acid equivalents (mg GAE 100mL⁻¹ of juice).

**MATERIAL AND METHODS**

**Sample and juice preparation**

Grapes (cv. Isabella and BRS-Cora) were harvested and provided by Embrapa Tropical Semi-arid, Petrolina, Pernambuco State, Brazil (latitude 9° 8’ 8.9'' S, longitude 40° 18’33.6'' W, altitude 373 m). Ripe fruits, which showed mean values of 0.7 g 100 mL⁻¹, 3.8 and 23 °Brix for total acidity, pH and sugar level, respectively, were harvested and stored at 10°C for 12h after processing. First, the grapes were sanitized with hypochlorite solution (100 mg L⁻¹) for 15 min and rinsed with water. Fruits were then processed by the steam extraction method for 60 min at 75–85°C. Sterilized glass bottles were used for juice storage (16°C) until analysis, within 30 days of juice bottling.

**Total polyphenol content (TPC)**

TPC was determined using the Folin–Ciocalteu method, with slight modifications. The extracts, distilled water and Folin–Ciocalteau’s reagent were added to vials and mixed thoroughly. After 3 min, 2 mol L⁻¹ sodium carbonate solution was added and the mixture was kept in the dark at room temperature for 2h. The absorbance was read at 725 nm using a spectrophotometer (Synergy HT, Biotek, Winooski, VT, USA). Gallic acid was used to prepare a standard curve and the results were expressed as gallic acid equivalents (mg GAE 100 mL⁻¹ of juice).

**Total anthocyanins**

Total anthocyanins were measured according to Wrolstad, with some modifications, as described by Abe et al. Appropriate dilutions from the samples were carried out using 0.025 mol L⁻¹ potassium chloride buffer (pH 1.0). The absorbance was read at 510 and 700 nm using a microplate reader (Synergy HT). A second dilution was made for each sample using 0.4 mol L⁻¹ sodium acetate buffer (pH 4.5) and the absorbance was read at the same wavelengths. The calculations were done as shown in the equation below:

\[
A = \left( \frac{(A_{510 \text{ nm}} - A_{700 \text{ nm}})}{pH = 1.0} \right) - \left( \frac{(A_{510 \text{ nm}} - A_{700 \text{ nm}})}{pH = 4.5} \right)
\]

(1)

The results were expressed as mg cyanidin 3-glucoside 100 mL⁻¹. The final results were calculated as follows:

\[
C (\text{mg 100 mL}^{-1}) = A.\text{MW.DF.100} ÷ \xi.1
\]

(2)

where \(\xi\) is molar absorptivity (26900 mol L⁻¹ cm⁻¹), \(l\) is path length (cm), \(\text{MW}\) is molecular weight of cyanidin 3-glucoside and \(\text{DF}\) is dilution factor.

**Analysis by high-performance liquid chromatography (HPLC)**

Phenolic compounds were identified and quantified by HPLC technique using a Waters Alliance e2695 HPLC system equipped with a photodiode array detector (DAD) and fluorescence detectors (FLD). Separations were performed on a C18 column (150 x 4.6 mm, 3 μm Gemini NX, Phenomenex, Torrance, CA, USA); preceding the analytical column a C18 guard column was used (4.0 x 3.0 mm, Gemini NX) at 40°C. The mobile phase consisted of a 25 mmol L⁻¹ solution of potassium dihydrogen phosphate in ultrapure water (PURELAB Option Q, Elga Systems, Woodridge, IL, USA) with pH adjusted to 2.05 using phosphoric acid (Fluka, Switzerland) as solvent A, methanol (HPLC grade, JT Baker, Phillipsburg, NJ, USA) as solvent B and acetonitrile (HPLC grade, JT Baker) as solvent C. The elution gradient used was as follows: 0 min 100% A; 18 min 87.5% A, 2.5% B, 10.0% C; 30 min 83.5% A, 3.2% B, 13.3% C; 36 min 75.0% A, 5.0% B, 20.0% C; 48.5 min 65.0% A, 8.3% B, 26.7% C; 50 min 65.0% A, 8.3% B, 26.7% C and 65 min 100% A. The flow rate and injection volume were 0.6 mL min⁻¹ and 10 μL, respectively. The juices were previously filtered through a 0.45 μm membrane before injection into the HPLC system. Phenolic compounds were identified by comparing their relative retention times (RT) and UV spectral data with those of identical standards at 520 nm (pelargonidin 3-O-glucoside, cyanidin 3-O-glucoside, delphinidin 3-O-glucoside, peonidin 3-O-glucoside, malvidin 3-O-glucoside), 220 nm (gallic acid, procyanidin B1, (--)-epicatechin gallate and (--)-epigallocatechin gallate), 320 nm (chlorogenic acid, caffeic acid, p-coumaric acid and ferrulic acid) and 360 nm (isorhamnetin-3-O-glucoside and rutin). The fluorescence detector was used for the detection of the vanillic acid, procyanidin B2 and A2, (--)-epicatechin, (+)-catechin and syringic acid at 280 nm of the excitation and 360 nm of the emission. The standards caffeic, vanillic, ferrulic and gallic acids were purchased from Chem Service (West Chester, PA, USA). Chlorogenic, syringic and p-coumaric acids were purchased from Sigma (UK). Pelargonidin 3-O-glucoside, cyanidin 3-O-glucoside, delphinidin 3-O-glucoside, peonidin 3-O-glucoside, malvidin 3-O-glucoside, procyanidin B1, procyanidin B2, procyanidin A2, epicatechin, catechin, epicatechin gallate, epigallocatechin gallate, rutin and isorhamnetin-3-O-glucoside were obtained from Extrasynthese (Genay, France). Calibration curves were prepared.
for these 21 standard phenolic compounds using the same chromatographic conditions. Table 1 shows the concentration range of each standard used for the calibration curves, standards purity (SP), equations of the calibration curves, coefficients of determination (R²), detection limits (LD), quantification limits (LQ) and assay repeatability through coefficient of variation (CV). Data collection and analysis were carried out using Empore 2 software (Milford, CT, USA).

Antioxidant activity

**DPPH radical scavenging activity**

The antioxidant activity was determined using the DPPH (1,1-diphenyl-2-picrylhydrazil) assay according to Brand-Williams et al., with some modifications. Briefly, 61 μmol L⁻¹ DPPH ethanol solution were added to the extracts, followed by shaking and incubation for 30 min in the dark. The absorbance was read at 515 nm using a Synergy HT spectrometer. The results were expressed as μmol Trolox equivalents (TE) 100 mL⁻¹ of juice.

**Hydrophilic oxygen radical absorbance capacity (ORAC) assay**

The ORAC assay was determined according to Dávalos et al. The extracts were prepared in phosphate buffer (PB) (75 mmol L⁻¹, pH 7.4). The peroxyl radical was generated using 2,2′-azobis (2-amidinopropane) dihydrochloride solution (AAPH) and fluorescein was used as the substrate. The reaction mixture consisted of extracts, 0.0014 mmol L⁻¹ fluorescein, and 0.4 mol L⁻¹ AAPH. The reaction was carried out in a plate reader (Synergy HT) with filters of emission at 520 nm and excitation at 485 nm. The incubation was conducted at 37 °C. The fluorescence was recorded every minute for 80 min. The Trolox standard was used to prepare a standard curve and the results were expressed as μmol TE 100 mL⁻¹ of juice.

**Ferric reducing antioxidant power (FRAP) assay**

The FRAP assay was carried out according to Rufino et al., with slight modifications. In the dark, FRAP reagent was prepared with 300 mmol L⁻¹ acetate buffer (pH 3.6), 10 mmol L⁻¹ 2,4,6-tris(2-pyridyl)-s-triazine (TPTZ) in a 40 mmol L⁻¹ HCl solution and 20 mmol L⁻¹ FeCl₃. Samples or standard solutions, ultrapure water and FRAP reagent were mixed and allowed to react in a water bath for 30 min at 37 °C. After cooling to room temperature, the absorbance of the samples and standards was read at 595 nm. Results were expressed in μmol TE 100 mL⁻¹ juice.

**Statistical analysis**

All analyses were carried out in triplicate and data were expressed as the mean value ± standard error (SEM). The statistical analyses were carried out using GraphPad Prism 5.0 software (GraphPad Software, Inc., La Jolla, CA, USA). The significance of the data was determined using Student’s t-test at P < 0.05.

**RESULTS AND DISCUSSION**

**Total phenolic and anthocyanin content**

Fruits and plant-derived beverages such as fruit juices are dietary sources of polyphenols. Polyphenols have received increasing interest from consumers and the food industry because of the direct association between their consumption and the prevention of several diseases. Plant polyphenols have antioxidant properties and therefore an important role against radical oxygen species (ROS). Enhancements in the cognitive function of adults after the ingestion of 6–9 mL grape juice kg⁻¹ body weight for 12 weeks have also been reported. Other authors have found positive effects in the lipid profile, antioxidant status, and antimutagenic activity after grape juice consumption, which supports the potential health benefits of this product.

In the present study, BRS-Cora grape juice showed higher bioactive substance concentrations than that found in grape juice prepared with the Isabella cultivar. BRS-Cora grape juice showed three times higher TPC (221.6 ± 1.83 mg GAE 100 mL⁻¹) than Isabella (74.86 ± 4.25 mg GAE 100 mL⁻¹) (Fig. 1a). Furthermore, BRS-Cora grape juice also had an anthocyanin content that was five times higher (73.88 ± 1.30 mg cyanidin-3-glucoside 100 mL⁻¹) than Isabella grape juice (15.40 ± 0.20 mg cyanidin-3-glucoside 100 mL⁻¹) (Fig. 1b).

The results presented here are in good agreement with those reported for Campbell Early grape juice, for which TPC was found at a concentration of 308.2 ± 11.0 mg GAE 100 mL⁻¹ and content of 496.6 ± 15.0 mg expressed as malvidin-3-glucoside 100 mL⁻¹ for total anthocyanins. Minor differences are expected due to different cultivars, climate conditions, soil quality and losses during the juice-making process. Compared to other fruit juices, the analyzed grape juices are rich sources of polyphenol compounds. Pala and Toklucu observed a lower anthocyanin content in pomegranate juice (42.6 ± 1.37 mg cyanidin 3-glucoside 100 mL⁻¹).

**HPLC analysis**

Phenolic compounds are widely distributed in plants, and grapes are well recognized as good sources of polyphenols from several classes. In the present study, 21 phenolic compounds were positively identified and quantified by HPLC (Table 2).

Anthocyanins represent a significant portion of the total polyphenols in grapes. These compounds are among the most important groups of water-soluble pigments, concentrated mainly in the fruit skin, being responsible for the purplish color of the grape and the antioxidant properties. Isabella grape juice showed a higher content of four monoglycosylated anthocyanins (Table 2), but delphinidin 3-O-glucoside was eight times more concentrated in BRS-Cora. Some works have shown prevalence of anthocyanins 3,5-diglucoside in non-vinifera grape cultivars and their hybrids. Diglucosidic anthocyanins were not investigated in the present study; however, as can be noted in Table 2, the phenolic total content (anthocyanins + phenolic acids + flavonoids/proanthocyanidins) was higher in BRS-Cora grape juice, supporting the TPC data. These compounds may be able to avoid certain diseases, thus providing a better life quality.

The major phenolic compounds of BRS-Cora grape juice were delphinidin 3-O-glucoside, proanthocyanidin B1, ferulic acid and caffeic acid. Proanthocyanidin B1, malvidin 3-O-glucoside, syringic acid and catechin were the major ones in Isabella grape juice. In general, the concentration of individual phenolic compounds was different (P ≤ 0.05) between the samples. BRS-Cora grape juice rendered a higher concentration of most of the phenolics quantified, which is in agreement with the trend observed in TPC and anthocyanin analyses.

Among all the phenolic compounds identified, only isorhamnetin and malvidin 3-O-glucoside were not found in BRS-Cora grape juices, and p-coumaric acid and epicatechin gallate were not found in Isabella grape juices. trans-Resveratrol, kaempferol-3-O-glucoside, myricetin and quercetin were not identified in either juice.
Supporting the findings of the present study, Sensoy et al. also found caffeic acid and syringic acid as the major compounds in varieties of red grapes. However, grape juices prepared with Sauvignon Blanc cultivar showed higher catechin and lower caffeic acid concentrations than those found in the present study, within the range of 4.5–7.7 and 0.41–3.7 mg L$^{-1}$, respectively. Such differences may be correlated with the different grape cultivars compared as the present study, which used red grapes, and these authors used Sauvignon Blanc, which is a white grape. However, Ghafoor et al. did not find caffeic acid in grape juice prepared with Campbell Early grape. According to Sun et al., ellagic acid is the major compound observed in different polyphenol profiles of some grape varieties (Fernão Pires, Castelão, Vital, Vinhão, Espadeiro, Azal Tinto). The present study is in accordance with the literature in relation to the variation in polyphenol composition among different grape cultivars. BRS-Cora was developed by Embrapa, Brazil, with the objective of creating new varieties for grape production, with high productivity, sugar content, color intensity and good sensory quality of grape juice, as well as the Vitis labrusca cultivars Isabella and Concord. Therefore, it was expected that BRS Cora grape juice had a higher anthocyanins...
content, as shown in Fig. 1. Additionally, Table 2 shows that BRS Cora also presents a higher content of other phenolic compounds compared to Isabella grape juice, suggesting that the presence of this cultivar in the juice can contribute to improve benefits of grape juice consumption for human health. Furthermore, different techniques used for juice production may generate different polyphenol contents in the final product, but it is not the case for the juices evaluated.

As previously mentioned, phenolic compounds have shown health benefits against LDL oxidation as well as in the reduction of DNA damage and free radical level in blood plasma.45–47 Furthermore, proanthocyanidins, which are among the major phenolic compounds of BRS Cora and Isabella grape juice, have been associated with the prevention of urinary tract infections.48 Grape seed extracts containing catechin, epicatechin and proanthocyanidins also demonstrated positive effects on prevention of Alzheimer’s disease in animal models.49 Sato et al.50 demonstrated that caffeic acid has strong antioxidant potential in vitro and in vivo. Finally, syringic acid has shown a hepatoprotective effect in mice subjected to induced liver injury.51 Thus phenolic compounds identified in BRS-Cora and Isabella will likely provide health benefits for consumers.

Antioxidant activity
Oxidative stress may rise in the cell, affecting the integrity of biomolecules and increasing the risk of oxidative-related diseases. Polyphenols found in berry fruits possess antioxidant properties,52 which may be beneficial against oxidative stress. The in vitro evaluation of antioxidant capacity is helpful in anticipating the potential of food products to prevent oxidative damage in vivo. It is also important for consumers, helping them to make decisions and choose foods to achieve a better diet. Data about the bioavailability of polyphenols are still scarce due to the complexity of this kind of investigation. However, some studies showing the presence of these compounds in blood and tissues and, most recently, metabolomics databases with information on metabolites are becoming available, which may contribute to establishing relationships among different sources of polyphenols, their intake and human health improvement.53–55

In the present study, the antioxidant properties of grape juices were evaluated by oxygen radical absorbance capacity (ORAC), ferric reducing antioxidant power (FRAP) and DPPH radical scavenging activity. ORAC relies on chain-breaking antioxidant capacity against peroxyl radicals,56 FRAP involves the reduction of ferric to ferrous ions via electron transfer,27 and DPPH involves hydrogen donation.25 Peroxyl radicals are reactive oxygen species (ROS), thus being detrimental to health due to the oxidative processes in which they are active. Because of their relatively long half-life,
their effect may occur at both a cellular level and in biological fluids. Ferric ions are also detrimental to biological systems, as they catalyze the oxidation of proteins and lipids. Finally, DPPH is a synthetic radical that has been widely applied to in vitro antioxidant evaluations as a preliminary screen of the antioxidant properties of several food products.\(^7\)

The results of the antioxidant evaluation are summarized in Fig. 2. The antioxidant activity of BRS-Cora grape juice was 2.3-, 3- and 4.5-fold higher than Isabella grape juice, as evaluated by ORAC, FRAP and DPPH, respectively. These results are supported by the higher TPC and anthocyanins found in BRS-Cora grape juice (Fig. 1). Furthermore, a strong positive correlation was found in TPC or anthocyanins and the remaining antioxidant assays (ORAC, FRAP and DPPH) (Table 3).

Antioxidants can stop free radical chain reactions via the donation of hydrogen from their phenolic hydroxyl groups or electrons, thus generating stable end-products that do not take part in further oxidation processes.\(^8\) Therefore, in general, the antioxidant activity of food products reflects their polyphenol content, as demonstrated in the present study. Both grape juices had antioxidant properties in neutralizing peroxyl radicals and in reducing ferric ions, as evaluated by ORAC and FRAP, respectively. Both methods possess biological relevance.\(^9,60\) Thus grape juice consumption will likely have health benefits. Finally, BRS-Cora, which is a relatively new cultivar (it was launched in 2004),\(^44\) showed the highest antioxidant properties in all antioxidant assays when compared to Isabella. However, BRS-Cora is still underused. Thus the grape juice industry should consider the use of BRS-Cora to produce grape juice or use them to produce blends with Isabella, improving not only the color of the juice but also their health benefits.

CONCLUSION

BRS-Cora grape juice showed higher TPC, anthocyanin content and antioxidant potential than Isabella grape juice. Twenty-one phenolic compounds were positively identified and quantified using HPLC-DAD-FLD. Little difference was found in the identity of phenolics between the cultivars; however, their quantification demonstrated significant differences in the grape juices made with BRS-Cora and Isabella grapes. BRS-Cora grape juice had up to two and ten times higher concentrations of gallic acid and epigallocatechin gallate, respectively, than that found in Isabella grape juice. Furthermore, epicatechin gallate was identified and quantified only in BRS-Cora. Gallic acid and polyphenols bearing galloyl groups probably had an influence on the higher antioxidant potential of BRS-Cora grape juice. In this way, the use of BRS-Cora is a good alternative to produce grape juice or in blends with different cultivars. The new cultivar BRS-Cora has been used to improve the color of Brazilian grape juice; however, as new information, the present work demonstrated that the use of BRS-Cora may also offer improvements in the phenolic profile and antioxidant potential of the product, being a potential functional food.

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