

Draft genome of *Enterococcus faecium* CV167, a bacteriocinogenic strain isolated from raw milk in Minas Gerais, Brazil

Yasmin Neves Vieira Sabino,¹ Joice Fátima Moreira Silva,² Bruna Vieira Alonso,³ Isabela Vieira Barbosa,³ Paula Aparecida Azevedo Almeida,⁴ Claudia Oliveira Pinto,⁴ Humberto Moreira Hungaro,⁵ Marcio Roberto Silva,⁴ Guilherme Nunes de Souza,⁴ Geraldo Marcio Costa,² Heloisa Carneiro,⁴ Karina Neoob de Carvalho Castro,⁴ Aline Dias Paiva,⁶ Laura Maria Bruno,⁷ João Batista Ribeiro⁴

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT Here, we describe the draft genome sequence of *Enterococcus faecium* CV167, a strain isolated from raw milk in Brazil that exhibits antimicrobial activity against various pathogens. The draft genome has a completeness index of 99.3%, spans 2,736,418 base pairs, has a guanine–cytosine content of 37.93%, and encodes 2,554 proteins.

KEYWORDS antimicrobial activity, bacteriocins, genome sequence

Given the increasing selection of antibiotic-resistant strains in nature, the search for alternative antimicrobial strategies has become increasingly important. *Enterococcus faecium* CV167, the strain highlighted in this announcement, has demonstrated *in vitro* activity against a variety of pathogens, including *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Streptococcus agalactiae*, *Streptococcus uberis*, *Enterococcus faecalis*, and *Escherichia coli*. The results of the inhibitory activity using the spot-on-lawn assay (1) are shown in Fig. 1. This bacterium was isolated from a raw milk sample stored in a bulk milk tank at a dairy farm in Coronel Xavier Chaves, Minas Gerais, Brazil in February 2022. For bacterial isolation, 1 mL aliquots of milk samples were serially diluted (10^{-1} to 10^{-3}) in phosphate-buffered solution (pH 6.2; Merck, Germany), and 100 μ L was plated onto M17 agar (Sigma-Aldrich, USA). After incubation at 35°C for 48 h, isolated bacterial colonies were streaked onto fresh M17 agar for purification. The isolate designated CV167 was identified as a Gram-positive coccus with no catalase activity. The DNA of the bacteria was then extracted using the phenol–chloroform method (2). DNA quantity and quality were evaluated using NanoDrop 1000 UV/Vis (Thermo Scientific, Massachusetts, EUA), and the sequencing library was prepared with the Illumina DNA Prep Kit. The DNA was sequenced using 300 bp paired-end sequencing on the Illumina NextSeq 2000 platform, generating a sequence depth of 1,169 $\times$. FastQC 0.12.1 (3) was initially used to assess the quality of the sequencing data, which generated a total of 11,863,824 reads. These reads were trimmed using Trimmomatic 0.39 (4) with the following parameters: trailing: 10; leading: 10; sliding window: 4:20; and a minimum length of 50 bp. The trimming process resulted in the removal of only 1.37% of the reads. *De novo* assembly of the short reads into contigs was performed using SPAdes 3.15.4 (5), with the coverage parameter set to “auto” and k-mers of 21, 33, 55, 77, 99, and 127. Contigs shorter than 500 bp were excluded from the final assembly. The assembly quality was evaluated using QUAST 5.2.0 (6). A total of 61 contigs were generated, with a combined length of 2,736,418 bp, a guanine–cytosine (GC) content of 37.93%, and an N50 of 156,530. Genome completeness was assessed using BUSCO 5.5.0 (7), revealing 99.3% completeness for the *Bacilli* class. To confirm the

Editor Julie C. Dunning Hotopp, University of Maryland School of Medicine, Baltimore, Maryland, USA

Address correspondence to João Batista Ribeiro, joao-batista.ribeiro@embrapa.br.

The authors declare no conflict of interest.

Received 14 October 2024

Accepted 21 December 2024

Published 5 February 2025

Copyright © 2025 Sabino et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

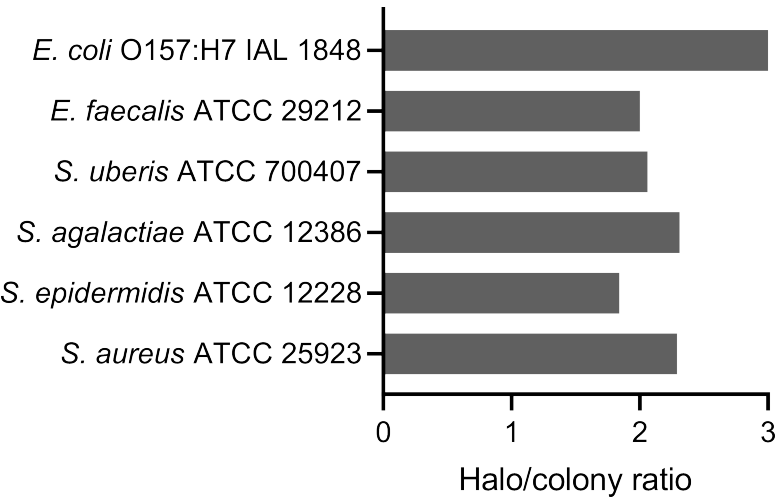


FIG 1 Antimicrobial activity of *Enterococcus faecium* CV167 against Gram-negative and Gram-positive bacteria. The inhibitory activity was evaluated using the spot-on-low technique and expressed as the halo/colony ratio.

TABLE 1 Main genomic features of *Enterococcus faecium* CV167

Genomic features	<i>E. faecium</i> CV167
Genome size	2,736,418 bp
Number of contigs	61
GC content	37.93%
N50	156,530
Protein-coding genes	2,544
Bacteriocin-encoding cluster (antiSMASH)	1
Bacteriocin-encoding cluster (BAGEL4)	3

taxonomic assignment, the assembled genome was analyzed using JSpeciesWS 4.2.1 (8) and *Enterococcus faecium* SRR24 as the reference genome (GenBank accession number [GCA_009734005.2](#)). The analysis revealed 99.45% nucleotide identity and a tetra score of 0.99745. Genome annotation performed using the NCBI Prokaryotic Genome Annotation Pipeline (9) identified 2,554 protein-coding sequences, 11 rRNAs, and 62 tRNAs. Further analysis using BAGEL4 (10) and antiSMASH v6 (11) to assess the potential synthesis of antimicrobial compounds uncovered several clusters encoding secondary metabolites, including enterocin variants. The main genomic features are summarized in Table 1. The draft genome sequence of *Enterococcus faecium* CV167 provides valuable insights into the molecular mechanisms underlying pathogen inhibition. This information is crucial for advancing genetic manipulation techniques and exploring potential industrial applications.

ACKNOWLEDGMENTS

We thank FAPEMIG for providing a fellowship to J.F.M.S. and CNPq for granting fellowships to B.V.A., P.A.A.A., and I.V.B.

This study was supported by the Minas Gerais Research Support Foundation (FAPEMIG, Process APQ 01196-23) and the Brazilian Agricultural Research Corporation (Embrapa, 03.13.14.006.00.00, 10.18.03.061.00.00, and 20.24.00.186.00.00).

AUTHOR AFFILIATIONS

¹Department of Parasitology, Microbiology and Immunology, Universidade Federal de Juiz de Fora, Juiz de Fora, Minas Gerais, Brazil

²Postgraduate Program in Veterinary Sciences, Department of Veterinary Sciences, Universidade Federal de Lavras, Lavras, Minas Gerais, Brazil

³Postgraduate Program in Science and Technology of Milk and Dairy Products, Department of Pharmacy, Universidade Federal de Juiz de Fora, Juiz de Fora, Minas Gerais, Brazil

⁴Embrapa Gado de Leite, Brazilian Agricultural Research Corporation, Juiz de Fora, Minas Gerais, Brazil

⁵Faculty of Pharmacy, Universidade Federal de Juiz de Fora, Juiz de Fora, Minas Gerais, Brazil

⁶Department of Microbiology, Immunology and Parasitology, Universidade Federal do Triângulo Mineiro, Uberaba, Minas Gerais, Brazil

⁷Embrapa Agroindústria Tropical, Brazilian Agricultural Research Corporation, Fortaleza, Ceará, Brazil

AUTHOR ORCIDS

Yasmin Neves Vieira Sabino  <http://orcid.org/0000-0001-6489-374X>

João Batista Ribeiro  <http://orcid.org/0000-0003-1502-4796>

AUTHOR CONTRIBUTIONS

Yasmin Neves Vieira Sabino, Methodology, Writing – original draft, Writing – review and editing | Joice Fátima Moreira Silva, Conceptualization, Investigation, Methodology, Writing – review and editing | Bruna Vieira Alonso, Investigation, Methodology, Writing – review and editing | Isabela Vieira Barbosa, Investigation, Methodology, Writing – review and editing | Paula Aparecida Azevedo Almeida, Investigation, Methodology, Writing – review and editing | Claudia Oliveira Pinto, Conceptualization, Methodology, Writing – review and editing | Humberto Moreira Hungaro, Conceptualization, Investigation, Methodology, Writing – review and editing | Marcio Roberto Silva, Conceptualization, Investigation, Methodology, Writing – review and editing | Guilherme Nunes de Souza, Conceptualization, Investigation, Methodology, Writing – review and editing | Geraldo Marcio Costa, Conceptualization, Funding acquisition, Methodology, Writing – review and editing | Heloisa Carneiro, Methodology, Writing – review and editing | Karina Neoob de Carvalho Castro, Conceptualization, Methodology, Writing – review and editing | Aline Dias Paiva, Conceptualization, Funding acquisition, Methodology, Writing – review and editing | Laura Maria Bruno, Conceptualization, Funding acquisition, Investigation, Methodology, Writing – review and editing | João Batista Ribeiro, Conceptualization, Funding acquisition, Investigation, Methodology, Writing – review and editing

REFERENCES

1. Tagg JR, Dajani AS, Wannamaker LW. 1976. Bacteriocins of gram-positive bacteria. *Bacteriol Rev* 40:722–756. <https://doi.org/10.1128/br.40.3.722-756.1976>
2. Oliveira SD, Santos LR, Schuch DMT, Silva AB, Salle CTP, Canal CW. 2002. Detection and identification of salmonellas from poultry-related samples by PCR. *Vet Microbiol* 87:25–35. [https://doi.org/10.1016/s0378-1135\(02\)00028-7](https://doi.org/10.1016/s0378-1135(02)00028-7)
3. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. Available from: <https://github.com/s-andrews/FastQC>
4. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>
5. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribylski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>
6. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>
7. Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* 31:3210–3212. <https://doi.org/10.1093/bioinformatics/btv351>
8. Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>
9. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
10. van Heel AJ, de Jong A, Song C, Viel JH, Kok J, Kuipers OP. 2018. BAGEL4: a user-friendly web server to thoroughly mine RiPPs and bacteriocins. *Nucleic Acids Res* 46:W278–W281. <https://doi.org/10.1093/nar/gky383>
11. Blin K, Shaw S, Kloosterman AM, Charlop-Powers Z, van Wezel GP, Medema MH, Weber T. 2021. antiSMASH 6.0: improving cluster detection and comparison capabilities. *Nucleic Acids Res* 49:W29–W35. <https://doi.org/10.1093/nar/gkab335>