REVIEW



Odorant binding protein as a management target for *Ceratitis capitata*: a window of opportunities for in vivo/in silico integration

Ramiro P. Guimarães¹ · Vanessa C. Santos² · Beatriz A. G. Paranhos³ · Nathaly C. Aquino⁴ · Ruth R. Nascimento⁴ · Edilson B. Alencar-Filho¹

Received: 8 July 2024 / Accepted: 18 December 2024 © The Author(s), under exclusive licence to Springer Nature Switzerland AG 2025

Abstract

Ceratitis capitata, known as Mediterranean fruit fly, represents one of the main problems to the agricultural production, due the ability to infest a wide variety of fruits, which makes it one of the most worrying pests globally. The infestation of host fruits by the insect compromises their quality and appearance, causing losses to their commercialization and consumption. In order to minimize this problem, fruit growers have adopted the indiscriminate use of conventional insecticides that raise environmental and public health concerns, in addition to promoting resistance in insects. Alternatively, semiochemicals have been explored as a management tool, attracting males and females into traps or promoting repellency in exposed fruits. In this context, in silico approaches, as Molecular Docking, Molecular Dynamics and the Virtual Screening procedures, offer opportunities to identify new molecular entities as potential ligands to Odorant Binding Proteins (OBP), that are involved in olfactory communication of *C. capitata*. In this review, we present a collection of data including the chemistry of olfactory communication of *C. capitata* and some computational tools that can be used in these studies, emphasizing their impact on the behavior of this and other associated insects. We also address theoretical ADME-Tox parameters as initial evaluation criteria to ensure human safety in the environmental applications, as well as the importance of molecular synergism for the effective management of the medfly.

Introduction

The Mediterranean fruit fly, *Ceratitis capitata*, is recognized as one of the most worrying pests due to its ability to adapt to a wide variety of fruits (European and Mediterranean Plant Protection Organization 2022). Its remarkable ecological flexibility and evolutionary capacity make it an invasive and polyphagous pest, with significantly negative

Communicated by Günther Raspotnig.

⊠ Edilson B. Alencar-Filho edilson.beserra@univasf.edu.br

Published online: 03 January 2025

- Postgraduate Program in Biosciences, Federal University of Vale do São Francisco, Petrolina, Pernambuco, Brazil
- Collegiate of Pharmaceutical Sciences, Federal University of Vale do São Francisco, Petrolina, Pernambuco, Brazil
- ³ Laboratory of Biological Control, Embrapa Semiárido, Petrolina, Pernambuco, Brazil
- Chemical Ecology Laboratory, Institute of Chemistry and Biotechnology, Federal University of Alagoas, Maceio, Alagoas, Brazil

economic impacts on the commercialization and industrialization of fruits, in addition to direct damage during preharvest (Papanicolaou et al. 2016; Zucchi 2015). The main crops affected by larvae infestation, originating from eggs deposited in fruits by adult females of C. capitata, include mango, grapes, papaya, pitanga, guava and acerola, creating a way for opportunistic bacteria and fungi. This results in necrosis, rot and premature ripening of the fruits, affecting their appearance and making them unsuitable for commercialization (Santos et al. 2007). Therefore, the incidence of these flies is a global situation, requiring frequent applications of insecticides to control the pest (Bittencourt et al. 2006). However, the indiscriminate use of these products has negative impacts on the environment, leading to the search for attractive or repellent molecules as a new approach to managing the insect (Melo et al. 2010).

In the context above, Molecular Docking and Virtual Screening computational strategies can be used as interesting tools to identify new molecules that can be used in pest control methods (Ferreira et al. 2011a), as well as explain the mechanisms of action of knowled ligands. Molecular Docking strategies are composed by "search algorithms", able to



predicting the best energy, conformation and orientation of a ligand in a specific site of a macromolecular biological target (Kitchen et al. 2004). This technique can be applied, for example, for the identification of molecules with potential affinity with proteins of *Ceratitis capitata* in Virtual Screening campagnes, using virtual libraries of compounds. An important target that emerges in this context is the Odorant Binding Protein (OBP), the structure located in the insect's antennae initially involved in olfactory communication and therefore an important target for species management.

OBPs play an important role in the olfactory system of insects, facilitating the detection of volatile molecules such as pheromones and compounds present in the environment (Rana et al. 2024). These proteins are present in the fluid of olfactory sensills, microstructures located mainly in the antennae and other sensory organs (Benton et al. 2009). With a compact structure, composed predominantly of six α-helices and stabilized by disulfide bridges, OBPs have a hydrophobic central cavity that allows the capture and transport of odorants through an aqueous medium to the olfactory receptors (Al-Jalely and Xu 2021). Their structural conformation protects them from environmental degradation, ensuring efficiency in the transmission of chemical signals (Vieira et al. 2007). In general, the transduction of the odorant signal begins with OBPs, which capture and transport odorant molecules to odorant receptors (ORs) in the membranes of olfactory neurons (Pelosi and Maida 1995).

This entire mechanism of action has some well-defined steps. At the beginning of the recognition process, proteins capture, through the pores of the antennae, odorant molecules present in the environment, interacting with them through Van der Waals forces, hydrophobic interactions, and, in some cases, hydrogen bonds (Leal 2013; Zhang et al. 2018). The binding capacity of OBPs is another notable aspect, being determined by the shape and composition of the hydrophobic cavity, which confers high affinity for specific odorants (Falchetto et al. 2019). Then, the odorant is transported by the OBP through the sensillary fluid, being protected from degradation until it reaches the ORs in the membrane of sensory neurons (Laughlin et al. 2008; Gomez-Diaz et al. 2013). ORs, in turn, are membrane proteins that activate a signaling cascade when an odorant binds to them. The activation of ORs leads to increased levels of cyclic AMP (cAMP), opening of ion channels and depolarization of the olfactory cell, generating an electrical signal that triggers the perception of odor and, consequently, the behavioral response of the insect (Breer 2003).

Near these receptors, odorant release is modulated by factors such as pH (neutral pH), which influences the affinity of the OBP and the subsequent release of the compound, allowing the odorant to bind directly to the OR, activating a signaling cascade that transmits the information to the insect's nervous system, activating the insect's behavioral response

(Zhou 2010; Brito et al. 2016a). This interaction, modulated by factors such as pH, alters the structural conformation of the protein, promoting the controlled release of the odorant (Leal 2013; Rana et al. 2024). Despite the knowledge about the signaling cascade and how the general mechanism occurs, which will be discussed later, the detailed interactions between OBPs and ORs and the dynamics of this transduction are still open questions (Rihani et al. 2021). Future studies should use advanced techniques to clarify how these interactions and variations influence olfactory perception.

OBPs also perform essential functions that go beyond simply capturing odorants. These confer specificity to the olfactory system, allowing insects to discriminate a wide range of chemical compounds, important for certain behaviors such as searching for food, choosing mates for mating, detecting predators, flying, and identifying places for egg deposition (Falchetto et al. 2019; Rana et al. 2024). In addition, OBPs protect odorant molecules from degradation before their interaction with receptors and adapt to different environmental conditions, such as pH and temperature variations, ensuring effective performance in various ecological scenarios (Pelosi et al. 2014; Rihani et al. 2021; Gu et al. 2015). Therefore, the study of these proteins has great practical relevance for the detection of compounds with semiochemical activity and in the development of synthetic pheromones for pest control, chemical biosensors and more effective repellents (Campanacci et al. 2001; Venthur and Zhou 2018), since OBPs represent an essential interface between the external chemical environment and the sensory perception of insects.

Considering that affinity and efficacy events in molecular signaling (physiological response) depends on the structural attributes of a molecule, the Docking could help in identifying molecular architectures with attractive or repellent potential, after experimental confirmation at light of theoretical affinity predictions. In other words, knowing from basic pharmacology that high affinity can result in agonist or antagonist effect depending on the mobilized amino acids, the computational technique can help to elucidate the targeting of repellents or attractors depending on the phenotypic response subsequent to the predictions. In addition, to improve and validate Docking results, Molecular Dynamics (MD) simulations are often used as a subsequent step for ligand-target complexes (De Vivo et al. 2016). These simulations generate the trajectory of atoms over time, allowing the evaluation of the conformational temporal profile of the ligand-protein complex, including their flexibility and the influence of the explicit aqueous medium on the interaction (Alonso et al. 2006). These aspects are importants to accurately predict binding energies and geometries (De Vivo et al. 2016; Alonso et al. 2006). In this paper, an extensive review about the biochemical and phytochemical aspects involved in the olfactory communication of C. capitata are



discussed, emphasizing the window of opportunities widely open for integration with some Computational Chemistry techniques.

Presentation and life cycle of *Ceratitis* capitata

The genus Ceratitis, belonging to the Tephritidae family, comprises approximately 100 cataloged species (White and Elson-Harris 1992; Virgilio et al. 2014). The Ceratitis capitata (Wiedemann) (Diptera: Tephritidae), whose presence dates back to the beginning of the twentieth century in Africa, spread to Mediterranean Europe, Central and South America through the intense global trade of fruits (Nava and Botton 2010; Metcalf 1995; Liquido et al. 1991). It is currently considered as one of the most worrying fruit pests, affecting around 400 fruit varieties (European and Mediterranean Plant Protection Organization 2022). Possessing great evolutionary flexibility in different environments (Papanicolaou et al. 2016; Zucchi 2015), it is an invasive and polyphagous pest, with negative economic impacts on pre-harvest fruit production (McPheron and Steck 1996). In this context, the genus Ceratitis stands out among the main global pests, together with five other genera: Anastrepha, Bactrocera, Rhagoletis, Dacus and Toxotrypana (Fazenda 2023). The genera Ceratitis and Anastrepha are the most economically relevant, since some of their species infest the majority of cultivated fruits (European and Mediterranean Plant Protection Organization 2022). In this regard, the insects becomes particularly destructive because it is capable of infecting around 200 varieties of fruits, from the moment the females pierce the fruits when laying their eggs, consequently allowing larval invasion and even opportunistic microbial invasion (Ouarhach et al. 2022). Insect pheromones are the main compounds that play an important role in intra- and interspecies communication, inducing specific behavioral responses in terms of sexual attraction, mating aggregation and aiding in the identification of the host fruit for egg deposition (Siciliano et al. 2014). After emerging, the adult Ceratitis capitata begins searching for the host fruit to reach sexual maturity, aiming to mate and lay eggs, starting a new cycle (Pereira 2003). The dimensions of the adult of this species vary from 4 to 5 mm in length and 10 to 12 mm in wingspan, with a predominantly yellow coloration, brown-violet eyes, a black thorax on the upper side with symmetrical white designs and a dark yellow abdomen, containing two grayish and yellow transverse stripes (Paranhos et al. 2008).

Considering its life cycle, the infestation process is initiated by the adult female of *C. capitata*, which lays its eggs in the host fruit, compromising its appearance, damaging the pulp and, consequently, reducing its quality for

commercialization and consumption (Vargas et al. 2001; Vontas et al. 2011; Dias-Pini et al. 2022). Fruit flies are holometabolite insects, and iniciate their metamorphosis in the host fruits: from egg, becoming a larva that consumes the fruit and goes to the ground, converting into a pupa, until they become adults. (Thomas et al. 2001). Females, in the sexual maturation phase, lay eggs on the fruit skin, preferably in the maturation stages, in a period that varies from 9 to 16 days after emergence (Malavasi 2009). After 2 days, the eggs hatch, and the larvae, during 6 to 10 days, go through three stages inside the fruit, feeding and producing galleries, depending on environmental conditions (Duarte and Malavasi 2000). The larva, in its last stage, leaves the fruit and buries itself in the soil for the pupal stage, which lasts 10–15 days (Carvalho and Nascimento 2002). Then, the adult insect emerges (Malavasi 2009). After emerging, the Ceratitis capitata adult begins the search for food necessary to reach sexual maturity, aiming to mate and lay eggs, starting a new cycle (Pereira 2003). At this stage, insects seek protein food and sugars for their sexual maturation by lodging themselves in the foliage of plants, mainly in the shaded area where the fruits are (Senar 2016). Mating occurs when the insects reach sexual maturity and, after copulation, the female begins to search for fruits to lay eggs (Raga and Sato 2016). It is at this moment that a new biological cycle of the species begins. The dimensions of the adult in this species vary from 4 to 5 mm in length and 10 to 12 mm in wingspan, with a predominantly yellow color, violet-brown eyes, a black thorax on the upper side with white symmetrical designs and a dark yellow abdomen, containing two stripes grayish and yellow transversals (Paranhos et al. 2008).

The life cycle duration of these flies is significantly influenced by abiotic and biotic factors, such as temperature, relative humidity, physiological conditions and types of host fruits (Arredondo et al. 2010). The availability and population density of hosts constitute biotic factors that impact population dynamics, oviposition preference and the biology of these organisms (Montes et al. 2011). Furthermore, C. capitata demonstrates a remarkable ability to adapt to different hosts and, in the absence of the primary or preferred host, resorts to secondary hosts, facilitating its persistence in available hosts (Leite et al. 2019). The primary host refers to the fruit in which the fly completes its life cycle in a short period, while the secondary host is the alternative fruit, used in the absence of the primary host, which can interfere unfavorably in the different stages of development of these tephritidae (Carvalho and Nascimento 2002). It is important to highlight that, in polyphagous insects, discrimination and selection of the appropriate host represent unpredictable behaviors, especially when these insects turn to other plants in the absence of their primary hosts (Aluja and Mangan 2008). Thus, the biological and demographic cycles of C. capitata populations have been explored in different hosts,



seeking a more in-depth understanding of the different life strategies adopted by this insect (Costa et al. 2011; Zanardi et al. 2011; Leite et al. 2019).

Economic importance and problematics involved

China, India and Brazil are the main fruit producers globally (Vidal 2023). Fruit flies are considered key pests in fruit farming, being responsible for annual losses of approximately one billion dollars worldwide, due to the damage caused and high control costs (Bittencourt et al. 2006; Marques 2023). They prefer juicy fruits, such as papaya, melon, peach, mango, guava and grapes (Haji and Alencar 2000). In grapevines, fruit flies are considered one of the main pests (Haji et al. 2009). In 2023, grape production reached 1,660,124 tons on 74,744 hectares, in Brazil (IBGE 2023a) Considering this economic importance, the risk caused by fruit flies deserves attention and its proper management could further optimize production (Nava and Botton 2010). Their damage in orchards not only makes export difficult, but also leads to the implementation of more stringent quarantine barriers by importing countries (Manrakhan et al. 2022).

The main control method is the chemical, through the application of synthetic insecticides, organophosphates and pyrethroids, whether in the form of toxic bait or spraying (Raga and Sato 2016). However, the indiscriminate use of these pesticides, their toxicity and non-biodegradability of some of them, have significant environmental impacts, in addition to being associated with several diseases (Melo et al. 2010). Given this scenario, as a proposed solution to the problem, alternatives have emerged in Integrated Pest Management (IPM), including the use of biological sources, biopesticides and semiochemicals, among others (Goulart et al. 2015).

Research aimed at understanding the reception of pheromones in C. capitata, for example, has indicated the possibility of developing new applicable attractants or repellents, replacing pesticides (Jang et al. 1989; Siciliano et al. 2014; Falchetto et al. 2019). In particular, non-toxic molecules known as semiochemicals, which are signaling compounds used in insect communication, produced and released in extremely small quantities, have the potential to improve control strategies such as monitoring, mass collection and sexual confusion, contributing to environmental preservation (Arioli et al. 2013). Furthermore, the characterization of these molecules involved in the behaviors of fruit flies sheds light on the understanding of their biology and ecology, providing a solid basis for the development of new pest control tools specific to each species, exploring and interfering in the chemical perception (Scolari et al. 2021).

Olfactory sensitivity

Several approaches to reducing insect pest population levels rely on modulating their behavior. (Camelo et al. 2007). The interpretation of environmental chemical signals plays a crucial role in the survival of these organisms, triggering diverse behavioral responses (Brito et al. 2016). Insects utilize a wide range of semiochemicals, such as pheromones and plant volatiles, to regulate essential behaviors such as identifying food sources, oviposition sites, suitable hosts, discriminating host plants, detecting pheromones, and locating mating partners. (Siciliano et al. 2014; Solari et al. 2018). Plant volatiles, which are complex mixtures of up to hundreds of compounds, play a crucial role in biosignaling of these behaviors (Fraser et al. 2003). Similar to plant volatiles, pheromones, commonly synthesized by male fruit flies, constitute a set of multicompound substances with different volatilities, allowing the transmission of complex messages for feeding, reproduction and defense (Vilela and Della Lucia 2001). In some cases, sexual signaling and pheromone emission begin in the presence of host plant volatiles (Papadopoulos et al. 2006), while in others, host plant volatiles synergistically amplify the response of a given sex to the sexual pheromones of opposite sex (Schmidt-Busser et al. 2009).

Over the years, studies have been dedicated to characterizing the chemical composition of pheromones and identifying their biologically active compounds (Alfaro et al. 2011; Light et al. 1988; Vaníčková et al. 2012; Siciliano et al. 2014). In the case of *C. capitata*, the pheromone compounds involved in sexual intercommunication can be classified as primary constituents ((E,E)- α -farnesene and geranyl acetate), intermediates (myrcene and linalool), and secondary or trace constituents (such as o (E)- β -farnesene and 2,3-butanediol) (Sollai et al. 2018). These compounds regulate the reproductive behavior of fruit flies through molecules released by males (Díaz-Fleischer and Aluja 2003).

Among the tephritid species studied, it is known that virgin males release substances to attract males and females to specific locations close to host plants, where mating and, subsequently, oviposition occur in fruits (Leal and Zucoloto 2008). Volatile compounds can influence the reproductive behavior of males in three distinct situations: increasing calling frequency, enhancing the attraction of females and serving as precursors of chemical signals released by males (López-Guillén et al. 2011). In the case of females, the odors released by males and host fruits function as olfactory cues for locating partners and selecting potential hosts for oviposition (Siderhurst and Jang 2006). The perception of these semiochemicals occurs in antennal olfactory sensilla, present in both males and



females, and which, even in small quantities, are noticed at great distances, depending on the wind speed (Gallo et al. 2002). Therefore, understanding the olfactory sensitivity of *C. capitata* may be valuable for developing more targeted control strategies using biologically active molecules (Reddy and Guerrero 2004). This alternative is considered acceptable in agriculture, compared to other forms of control, making the use of semiochemicals an attractive and viable strategy (Hassanali et al. 2008), as the use of natural or synthetic insect pest pheromones can significantly reduce the use of insecticides, thus minimizing the environmental impact (Ghini and Bettiol 2000).

General aspects of odorant binding protein (OBP)

OBPs are small, water-soluble extracellular transport proteins (Zhou et al. 2010). These biochemical structures, responsible for the odors transport, are located in the insect antennae, specifically in cuticular structures called "sensilla" (Klein 1987). Sensillas have pores that allow the passage of odorous substances that then come into contact with the sensilla lymph, a cytoplasmic liquid where OBP as carrier proteins are highly abundant (up to 10 mM) (Vogt and Riddiford 1981; Vogt et al. 1989). Surrounded by the sensillary lymph are the dendrites of olfactory sensory neurons, which are nerve cells specialized in processing and transmitting odorous stimuli (Fan et al. 2011; Leal et al. 2013). In this environment, OBPs bind to hydrophobic odorant substances and transport them to OR, located in the membranes of dendrites (Pelosi 1994; Biessmann et al. 2010; Pelletier et al. 2010). By transporting volatile compounds under ideal pH conditions to the ORs, the olfactory message is converted by ORs into behavioral responses (Zhou et al. 2010) (Fig. 1). Detailing the signaling cascade, the ORs are G-protein-coupled receptor (GPCR). Each receptor is specialized to binding specific classes of odorants or pheromones. Upon binding the ligand, the OR undergoes a conformational change that activates the associated G-protein, releasing an α-subunit that activates adenylyl cyclase, an enzyme that catalyzes the conversion of ATP into cyclic AMP (cAMP). The increase in cAMP levels leads to the activation of Protein Kinase A (PKA), that can phosphorylate a variety proteins within the olfactory sensory neuron. The cAMP production also leads to the opening of cation channels, allowing Na⁺ and Ca²⁺ ions to flow, depolarizing the olfactory sensory neuron and generating an action potential. The depolarization is the initial electrical signal in the process of odor detection (Fan et al. 2011).

Additionally, several studies, performed on different species of insects, revealed that these proteins play not only the role of passive transporters of odorants, but are also involved

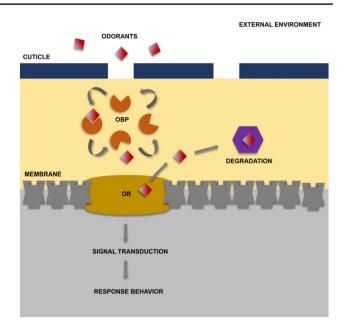


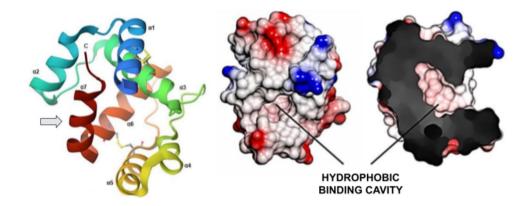
Fig. 1 Schematic representation of the recognition of volatile compounds by the odorant protein. Source: Adapted from Sanchez-Garcia et al. (2009)

in the discrimination of semiochemicals, functioning as the first molecular filter in the selection process (Chang et al. 2015; Leal 2005; Yin et al. 2015). The OBP primary sequence of insects, for example, determines the binding site specificity for different ligands, as evidenced by several molecular structures obtained through X-ray crystallography and NMR spectroscopy (Tsitsanou et al. 2012). Considering the main interest of this review, the OBP of Ceratits capitata is deposited in the online Protein Data Bank (PDB ID: 6HHE), of which the three-dimensional structure was determined from studies involving X-ray diffraction. (Falchetto et al. 2019). In this sense, the number of candidate genes for a structural approach targeting the olfactory system continues to grow consistently, especially after the completion of genomic sequencing of several insect species, such as Drosophila melanogaster and Anopheles gambiae (Adams et al. 2000; Holt et al. 2002).

The three-dimensional architecture of insect OBPs generally has a molecular weight ranging from 15 to 20 kDa and is composed of a domain of six α -helices, which form a hydrophobic cavity (González-González et al. 2019; Sun et al. 2018a, b; Vogt 2002). This cavity is stabilized by three disulfide bridges, which interconnect conserved cysteines, conferring stability to the structure (Leal et al. 1999). In the case of the *C. capitata* OBP, there is a distinct structural characteristic: the presence of a seventh α -helix in the C-terminal region (Fig. 2). This additional helix plays a crucial role in defining the central hydrophobic binding cavity of the odorant molecule, allowing greater stability of the bound molecules compared to OBPs from other insects, which



Fig. 2 Three-dimensional structure of the odorant protein from *Ceratitis capitata* and the representation of the hydrophobic binding cavity, where the ligand interacts with the target macromolecule. The arrow is indicating the seventh alpha-helix in red of the protein. Source: Adapted from Falcheto et al. (2019)



typically have only six α -helices. Furthermore, the seventh α -helix enables the closure of the cavity when the odorant compound binds to the protein, a phenomenon influenced by the pH of the hemolymph, which ensures the stability and functioning of the protein (Falchetto et al. 2019).

Furthermore, the OBP is genetically more expressed in females and plays a crucial role in intersexual olfactory communication (Falchetto et al. 2019). In C. capitata, the male synergistically releases the intersexual communication pheromone in trace quantities, with the aim of attracting the female for mating, at the same time, it also attracts males, thus facilitating the encounter with females (Siciliano et al. 2014). This process results in increased pheromone concentrations, making it more effective in attracting a greater number of females under specific physiological conditions (Prokopy and Hendrichs 1979; Jang et al. 1989). The increase in concentrations is strongly influenced by the sexual maturity of males, with the glands responsible for pheromone production becoming more active in the reproductive phase (Heath et al. 1993). Pheromone production in males is also regulated by circadian rhythms, with peaks in release occurring during the hours of the day most conducive to mating, usually in the late afternoon. This synchronization maximizes the chances of reproductive success, ensuring that males release pheromones at times of greatest female activity (Landolt and Sivinski 1992). In addition, environmental factors such as the presence of receptive females and lighting conditions influence the amount of pheromone released. Males adjust their release in response to external stimuli, such as the perception of pheromones from other males, increasing competition for female attention (Jang and Light 1996).

As already mentioned, the predominant compound in the pheromone released by *C. capitata* males is (E,E)- α -farnesene (Gonçalves et al. 2006; Jang et al. 1989; Merli et al. 2018). This component demonstrates a remarkable specificity and affinity with the Odorant Binding Protein (OBP) when compared to the five main compounds of the Mediterranean fly pheromone (Falchetto et al. 2019).

Therefore, the six volatiles that make up the male sex pheromone of *C. capitata* are: α -farnesene and geranyl acetate (major components), linalool and β -myrcene (intermediate components), β -farnesene and 2,3-butanediol (minor/trace components) (Sollai et al. 2018).

These compounds play a crucial role in several complex behaviors, such as cutting, aggregation, associative learning, search for and acceptance of food, in addition to host plant recognition by insects (Biolchini et al. 2017). (E,E)-αfarnesene is also an active component in antennal and behavioral pheromones in other tephritid species, such as Anastrepha ludens and Anastrepha suspended (Rocca et al. 1992). More specifically, (E,E)- α -farnesene is a volatile compound released by plants, especially in ripe fruits, and acts as an attractant for insects such as fruit flies, guiding them to food sources or egg-laying sites (Bruce and Pickett 2011). In contrast, some male insects emit alpha-farnesene as signs of sexual attraction or to mark territory, demonstrating the versatility of the compound in different biological contexts (Schwab et al. 2008). However, it is important to highlight that male fruit flies do not produce alpha-farnesene; they obtain the compound from the plant. The (E,E)- α -farnesene found in ripe fruits is used by males of some species of fruit flies as a component in their sexual pheromones (He and Zhang 2012). They collect (E,E)- α -farnesene and other volatile compounds from fruits to incorporate into their own pheromone emissions, which help attract females for mating (Witzgall et al. 2008). This behavior highlights the importance of the interaction between the compound produced by the plant and the mating strategy of insects.

Fruit flies detect alpha-farnesene through highly sensitive OR on their antennae, which can capture very low concentrations of the compound (Bruce and Pickett 2011). This high sensitivity is crucial for flies to locate the source of the odor, playing a vital role in their survival and reproductive success (Cha et al. 2012). In addition, the activation or deactivation of olfactory receptors (OR) on fruit fly antennae is influenced by the concentration of volatile compounds, the age of the insect and its physiological state (Schwab et al.



2008). The effectiveness of (E,E)- α -farnesene as an attractant for fruit flies, for example, depends on its concentration. Very low concentrations may not activate ORs, while very high concentrations may cause repulsion or temporary desensitization. The ideal concentration range to attract fruit flies generally varies from parts per billion (ppb) to parts per million (ppm), balancing attraction and avoiding receptor saturation (Cha et al. 2012).

Considering this information, the manipulation of chemical communication emerges as a promising option to optimize control strategies for Mediterranean fly populations (Keil 1999). Insects' sense of smell plays a crucial role in understanding plant-host interactions, intersexual communication between male and female, and oviposition promoted by females (Gadenne et al. 2016). Therefore, to contain the populations of these insects, control methods that use sexual pheromone compounds in the agricultural sector can be employed (Zarbin et al. 2009), aiming to increase the effectiveness of capture systems for these flies (Tan et al. 2014).

In this scenario, structural bioinformatics and chemoinformatics stand out as crucial tools for the in silico identification of promising ligands capable of interfering with the olfactory system of *C. capitata*, especially in females, which play an essential role in the fruit infestation process (Carey and Carlson 2011; Suckling et al. 2013; Pelosi et al. 2014).

Compounds that bind to odorant protein

Compounds that interact with OBPs, triggering behavioral responses, are often found in fruit extracts and absorbed by males to compose the pheromone (He and Zhang 2012; Witzgall et al. 2010; Jacobson et al. 1973). These compounds, absorbed by males, play a vital role in intersexual communication, while those released by the fruits themselves facilitate the location of host plants, being essential for insects to meet their nutritional needs and find suitable sites for oviposition (Visser 1986; Knudsen et al. 1993). Male fruit flies, Ceratitis capitata, use specialized organs called accessory glands to absorb and process the volatile substances that make up the pheromone (Jang and Light 1996). In addition, males also have specialized structures called peritrema, which help in the emission of pheromones and in the detection of chemical substances in the environment (He and Zhang 2012). Geranyl acetate, (E,E)-α-farnesene, linalool (Baker et al. 1985; Jang et al. 1989), limonene, (E)-βocimene, indole (Cossé et al. 1995) and myrcene (Ohinata et al. 1977) are examples of volatile compounds absorbed and released by these structures (Jang and Light 1996).

Female fruit flies, in turn, detect these pheromones using ORs located on their antennae (Jang and Light 1991). Antennae are highly specialized sensory organs, equipped with olfactory neurons capable of capturing and processing

chemical signals such as pheromones, which trigger specific behaviors, such as attraction for mating (Witzgall et al. 2010). These responses are in agreement with findings reported for C. capitata and other tephritid species, where some key components of the male sex pheromone (geranyl acetate and (E,E)- α -farnesene) and host plant volatiles elicit behavioral changes in females (Fein et al. 1982; Jang et al. 1989; Jang and Light 1996; Light et al. 1988a; Robacker et al. 1986; Robacker and Hart 1987; Pers et al. 1984). Siciliano et al. (2014) also points out that Ceratitis females have a greater sensitivity to recognize the compound (E,E)-α-farnesene (Siciliano et al. 2014). Corroborating this information, this compound and other similar ones are also identified in the pheromones of other genera, such as Anastrepha, which belongs to the Tephritidae family (known as fruit flies) (Belliard et al. 2022).

In the species Anastrepha fraterculus, the compound limonene was identified in the sexual pheromone released by the male, directly influencing the responses of females and altering their way of reacting to homospecific and heterospecific pheromonal mixtures (Cáceres et al. 2009; Břízová et al. 2013). This same species also revealed the presence of α -pinene and (E,E)- α -farnesene in the antennae of females (Milet-Pinheiro et al. 2015). A comparative study of the volatile compounds released by Anastrepha suspensa and Anastrepha ludens identified five new compounds in the pheromonal mixture, including (E,E)- α -farnesene and limonene (Rocca et al. 1992). In Anastrepha striata, linalool, myrcene, limonene, (Z)- β -ocimene, (E)- β -ocimene, terpineol and (Z,E)- α -farnesene were found (Cruz-López et al. 2015a). Finally, the chemical composition of the pheromone from Anastrepha obliqua males indicated that linanol came from the host, being found exclusively in aeration extracts and not in salivary gland extracts (Gonçalves et al. 2013).

Thus, through these studies it is possible to observe that different species of fruit flies present similarities in the composition of the pheromonal mixture with *C. capitata*, and can, consequently, promote these communications and be used as standards in the selection of new attractive molecules or repulsive (Falchetto et al. 2019).

Natural compounds that interfere with the olfactory sensitivity of insects

Some studies highlight the influence of natural substances on the olfactory sensitivity of insects, in general. Thus, the correlation of other species with *C. capitata* is possible in some extension, because insect OBPs share some common characteristics in terms of size, the constitution of alpha helices, highly conserved disulfide bonds and an internal cavity for binding volatile compounds (Falchetto et al. 2019). This may be a challenge in the search for selectivity, but it



also encourages more detailed research to find small amino acid changes, pockets or specific prominences in the odorant receptor topology that may allow more specific interactions in a specific species. Furthermore, thinking about future application, it is important to study the environmental mapping of the most frequent insects in geographic regions of pest management. Despite these contradictions, regarding the molecular characterization of this review, we have that the benzyl benzoate compound, for example, provided greater attraction for C. capitata males compared to trimedlure (reference attractant) (El-Metwally et al. 2019). It is an aromatic ester that has a balsamic, herbal and oily smell (Menenses et al. 2020), present in clove oil. The same compound was also tested in *Drosophila melanogaster* (fruit fly) larvae, indicating an attractive olfactory response (Khurana and Siddiqi 2013). Rocca et al. (1992), carrying out a comparative study of the volatile compounds released by Anastrepha suspensa (Caribbean fruit fly) and Anastrepha ludens (Mexican fruit fly), identified five new compounds in the pheromonal mixture, including β -bisabolene and (E,E)- α farnesene, with intense attractive activity. Aluja et al. (2020) also point out that of the 14 components present in the pheromone of A. ludens and A. obliqua males, β-bisabolene acted in conjunction with the other molecules, modifying the behavior of the larvae of these tephritid polyphagic pests. In a quantitative analysis of pheromone blends in males of A. suspensa, β-bisabolene, together with solid suspension, anastrephin and epianastrephin, stood out as one of the main components of the pheromone, playing a significant role as an attractant for females. (Ponce et al. 1993). Similarly, in another study, in the pheromone of A. suspensa males, in addition to the presence of α -farnesene, β -bisabolene was also found, promoting intersexual communication (Lima-Medonça et al. 2014). In agreement, essential oils from citrus fruits with β-bisabolene were attractive in stimulating oviposition by *C. capitata* females in the host fruit (Ioannou et al. 2012). (+)- α -curcumene, an aromatic sesquiterpene, was reported to stimulate the antennae of female Aedes aegypti together with other sesquiterpenes (β-sesquiphellandrene, zingiberene and β-bisabolene) extracted from ginger essential oil (Zingiber officinale, Zingiberaceae) (Campbell 2009). Ceratitis capitata and A. aegypti are from the order Diptera, which makes it possible to correlate work on these two species (Luu-Dam et al. 2021). Liu et al. (2022) also highlight the presence of α -curcumene as a volatile compound extracted from a northeastern Chinese plant (Rhododendron dauricum), acting on insect OBPs. The germacrene B present in the composition of the oil from the leaves of Commiphora leptophloeos (imburana) was recognized as a source of compounds with repellent and larvicidal activities against the dengue mosquito (Silva et al. 2015). Furthermore, in essential oils extracted from tangerine (Citrus reticulata) and tea tree (*Melaleuca alternifolia*), germacrene B also has repellent activity against *Drosophila suzukii* (spotted wing fly) (Bedini et al. 2020) (Fig. 3).

β-longipinene showed a repulsive response in antennae of female Aedes aegypti when used in a synthetic mixture containing 1-octen-3-ol, (-)-isolongifolene, (+)-longifolene and 3-caryophyllene with 100 ng of each substance (Zhang et al. 2011). α-copaene isolated from clove oil was 5 to 6 times more attractive than trimedlure, the main commercial attractant for C. capitata males, confirming the interaction of this substance with the odorant protein (Lull et al. 2023). Vaníčková et al. (2012a) found α-copaene in the mixture of C. capitata sexual pheromones promoting an attractive response. The volatiles in ginger root oil, which mainly contain α-copaene, also modified the behavior of several species of fruit flies of the genus Anastrepha, named as Anastrepha fraterculus, A. ludens, A. obliqua and Anastrepha serpentina (Wiedemann) (Dip., Tephritidae) (Ruiz et al. 2021). α -copaene was also one of the main volatile constituents found in guava, along with α-caryophyllene promoting attractive olfactory responses in the fruit fly genus Bactrocera (Jaleel et al. 2021). In contrast, this same compound showed no evidence of repellency to A. aegypti mosquitoes (Dekker et al. 2011). Regarding α-humulene, Silva et al. (2015) revealed that the presence of this compound caused changes in the behavior of A. aegypti and presented high repulsive activity at low concentrations (5 ppm). Jaleel et al. (2019) reported that through olfactometer tests, the mixture of β -caryophyllene and α -humulene was of good attractiveness to females of Bactrocera correcta (Asian fruit fly). In C. capitata, males when exposed for three days to the aroma of α -humulene performed significantly fewer matings than males that were not exposed, consequently exhibiting a signaling suppressive activity in intersexual communication (Shelly and Nishimoto 2015). In contrast, males of A. fraterculus (South American fruit fly) when exposed to a mixture of seven compounds released by guava, including α-humulene, increased their courtship behavior, accentuating the mating process (Bachmann et al. 2015).

β-chamigrene was found among the volatiles of orange, lemon, bergamot and apple fruits, together with limonene, attracting C. capitata females to the host fruit discovery behavior and in the fruit's susceptibility to oviposition infestation (Antonatos et al. 2023). The essential oil of Melaleuca alternifolia had α-selinene, α-humulene and α-copaene in its composition and triggered olfactory responses to the emissions of these volatiles in C. capitata males (Tabanca et al. 2020). The essential oil of Magnolia citrata, composed of α-selinene, α-humulene, β-caryophyllene and other molecules, exhibited moderately strong attraction for males of C. capitata (Luu-Dam et al. 2021). The essential oil of Cupressus funebris (Chinese weeping cypress) showed repellent activity against adult yellow fever mosquitoes, A. aegypti, having in its



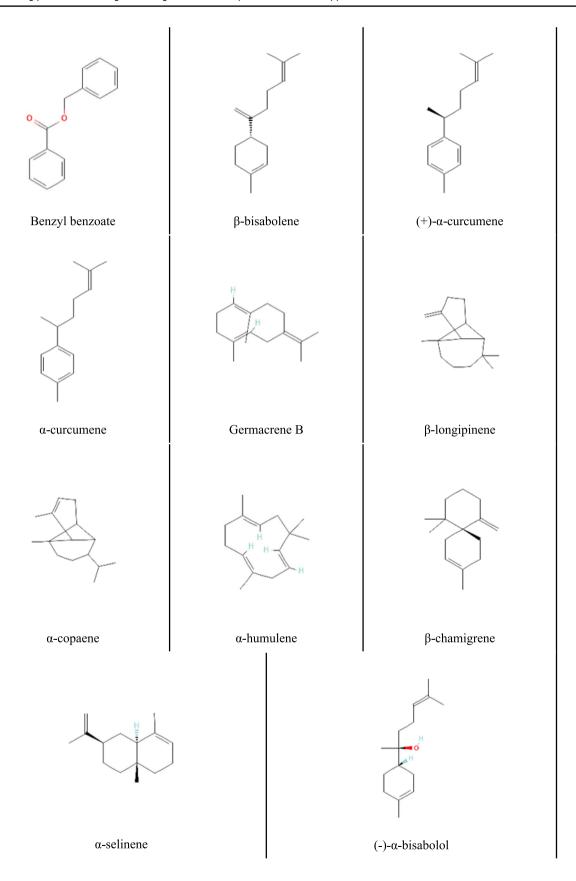


Fig. 3. 2D attractive and repellent compounds (Source: PubChem)

composition the sesquiterpene β -chamigrene, as well as α -selinene, γ -bisabolene, among others (Carroll et al. 2011). α -selinene also triggered antennal responses in A. aegypti females, together with α -humulene, determining them, therefore, as active compounds present in the essential oil of the leaf of *Commiphora leptophloeos* (imburana do sertão) (Silva et al. 2015).

(-)-α-bisabolol is a monocyclic sesquiterpene natural product, also called levomenol, colorless viscous oil with a pleasant aroma, weak floral odor with a tertiary hydroxyl group and a six-membered ring, two trisubstituted unsaturations, has the minimum formula C₁₅H₂₆O and molecular weight of 222.37 g (Nemoto et al. 1993). In nature, it occurs in enantiomeric (+) and (-) forms, with only the (-) enantiomer having biological activities (Megwer and Schornow 2001). Furthermore, regarding the control capacity promoted by this compound, in a research involving six constituents of an essential oil, (-)- α -bisabolol indicated significant repellency to the eastern fruit fly, Bactrocera dorsalis, when applied separately in the Y-tube olfactometer (Jaffar and Lu 2022). This species is also a fly from the Tephritidae family like C. capitata, possibly pointing to the action of this compound as a repellent against Mediterranean fruit flies. Four terpenes including (-)-α-bisabolol showed affinity with the odorant protein (AaegOBP1) of A. aegypti, also demonstrating protective-repellent activity comparable to DEET (N,N-diethyl-3-methylbenzamide) (Portilla-Pulido et al. 2020). (-)- α -bisabolol extracted from the essential oil of German, Roman and Chinese chamomile flowers also promoted repulsive activity for A. aegypti with an action similar to DEET (Ali et al. 2023).

DEET is the most used insect repellent in the world and is characterized by blocking the electrophysiological responses of olfactory sensitivity to attractive odors, mainly in Anopheles gambiae and Drosophila melanogaster (Ditzen et al. 2008). In an affinity study, it was possible to indicate that the interaction of DEET with the OBP takes place in the hydrophobic cavity, the characteristic binding region of insect OBPs (Tsitsanou et al. 2012). Unlike an attractive substance that, when transported by the OBP to the OR, promotes the activation of the biosignaling process, DEET, when transported to the OR, blocks it, preventing the recognition of another molecule, which consequently causes, the removal of the insect (Ditzen et al. 2008). Despite the repellent effect, the main concern with widely used synthetic compounds is the insect's potential to develop resistance or insensitivity (Degenaro et al. 2013; Wagman et al. 2015), as well as concerns related to possible toxicity and carcinogenicity for mammals (Legeay et al. 2016). Thus, in recent years, research into new insect repellents has focused on natural products, as some plant-derived repellents target multiple odor receptors (Thireou et al. 2018). Therefore, (-)- α -bisabolol emerges as a promising repellent compound,

with the potential to integrate control formulations, effectively replacing DEET.

Nesterkina et al. (2018) also highlighted the importance of the hydroxyl group in the repellent activity of the compounds, highlighting the lack of repellency of ρ-Cymene, devoid of the OH- group, unlike menthol, which exhibited repellent activity very close to that of DEET. This highlight reinforces the role of (-)-α-bisabolol, which has repellent activity, due to the presence of the OH- group in its composition, contributing to the effectiveness of its repellency. Over time, repulsive activity has been associated with the structural aspects of compounds, mainly in relation to functional groups, as evidenced by older and well-established works. Garson and Winnike (1968), for example, pointed out that compounds containing amides, imides, phenols, alcohols, hydroxy ethers, glycols and hydroxy esters were active repellents, while the original hydrocarbons were weak repellents. Bunker and Hirschfelder (1925) and Roadhouse (1953) corroborate this information, highlighting that the presence of the amide and phenol group are one of the main chemical groups that promote repellency. Christophers (1947) further reported that the repellent activity of alcohol groups, along with aldehydes and phenols, raised the hypothesis that the repulsive activity was related to the positioning of the -OH groups. Given this, it can be said that these compounds that interfere with the olfactory sensitivity of insects, whether in an attractive or repellent way, can boost the identification of compounds, through virtual screening, that bind to the odorant protein of C. capitata, acting as excellent potential for the management of these flies.

Molecular docking and virtual screening

Virtual screening is an in silico method used to examine large databases of compounds, aiming to select molecules with high affinity in target proteins, for subsequent in vitro biochemical evaluation (Ferreira et al. 2011). Through its search and selection mechanisms, virtual screening has established itself as an effective, fast and economically viable technique, allowing the reduction of the number of compounds that require experimental evaluation (Ferreira et al. 2011). There are several approaches to virtual screening, and choosing the most appropriate one depends on the availability of information (Pinto et al. 2022). The strategies of this method are related to molecular database investigation tools, which make use of computational models and algorithms to filter compounds based on desired characteristics (Schneider 2010).

To achieve this, the strategies adopted for screening are based on the structure of bioactive ligands (LBVS—Ligand-Based Virtual Screening) or on the structure of the biological receptor (SBVS—Structure-Based Virtual Screening)



(Guido et al. 2008). In the last process, virtual screening employs molecular docking, a technique for predicting the preferred binding orientation of a molecule to a receptor by calculating binding energies (Morris and Lim-Wilby 2008). This computational approach makes it possible to couple thousands or even millions of compounds to a target protein, identifying potential inhibitors or modulators of signaling pathways (Trott and Olson 2009; Lee et al. 2015; Kurcinski et al. 2019). This is made possible by optimizing docking algorithms, allowing ligands to access the target protein and revealing favorable binding sites and potential adverse interactions (Mikovski et al. 2018).

The results of the docking-based screening are scored based on several criteria. The first consists of the Binding Energy Score, which evaluates the strength of the interaction between the ligand and the target, usually calculating variation in energy of the system when the ligand is bound to the target, providing the changes in enthalpy and entropy (Morris et al. 2009). The second is the Affinity Score, which reflects the affinity and stability of the ligand for the target, which can be associated with the dissociation constant (Kd). A lower score generally indicates a better affinity (Morris et al. 2009). The third is the Fit or Fit Score, which evaluates the spatial geometry of the ligand in the binding pocket of the target, often considering the overlap with a standard geometry. The RMSD (Root Mean Square Deviation) is the common metric for this score, used to measure the distance between the ideal position and the calculated position of the ligand (Goodsell and Olson 2000). The fourth and final criterion is the Specific Interaction Score, which indicates the specific interactions between functional groups of the ligand and the amino acid residues of the target protein, such as hydrogen bonds, ionic interactions and hydrophobic interactions (Jones et al 1997).

The assembly of molecular libraries is also a crucial step in compound screening, involving the creation of a diverse set of molecules to test interactions with a specific target. The assembly of these libraries can be done in several ways. Chemical Compound Libraries consist of the set of chemical molecules that may include small compounds, peptides or other chemical entities, and can be assembled by direct chemical synthesis, combinatorial, or by purchasing ready-made compounds from specialized suppliers (Graaf and Benson 2011). Virtual Compound Libraries can be assembled using molecule generation algorithms and molecular modeling techniques to create a diverse set of structures, thus obtaining a set of molecules generated by computational simulation, as fragments or derivatives of known compounds (Bender et al. 2021). Natural Substance Libraries can be made by direct extraction from natural sources or by chemical synthesis based on known natural structures, forming collections of compounds derived from natural sources, such as plants or microorganisms (Newman and Cragg 2007). In addition to these, it is possible to assemble Peptide or Protein Libraries and DNA or RNA Libraries that respectively collect a set of peptides or proteins, and a set of DNA or RNA sequences, often used to study genetic interactions or gene functions (Krumpe and Mori 2007; Wu et al. 1999).

In molecular docking, two computational steps are established: (i) the molecular docking algorithm, which evaluates the different conformations and degrees of freedom of the ligand according to the properties of the interaction site, generating a list of poses that best complement the cavity, and (ii) the scoring functions, which perform approximate calculations to evaluate the electrostatic complementarity and van der Waals interactions between the ligand and the receptor (Gohlke and Klebe 2002). In this sense, Autodock VINA is the most widely used molecular docking program today, employing the grid box concept to identify the binding site and reduce the analysis time for each molecule (Eberhardt et al. 2021). The grid-based method calculates and stores the energy potentials between ligand atoms at various points in the interaction cavity region of the molecular target (Trott and Olson 2009). This pre-calculated data is stored in libraries by AutoGrid and then AutoDock VINA uses it to speed up interaction calculations (Kitchen et al. 2004). On the other hand, there are now a wide variety of paid and free online programs and platforms available for performing molecular docking and, with this tool, virtual screening. Among the most cited programs, we can highlight the GOLD (Jones et al. 1997), Dock (Gschwend and Kuntz 1996), Molegro (Bitencourt-Ferreira et al. 2019), SwissDock (Grosdidier et al. 2011), DockThor (Guedes et al. 2024), among others.

In pioneering work with the organism *Bactrocela dorsalis*, Jayanthi et al. (2014) proposed a useful virtual screening protocol for this oriental fruit fly, combining molecular docking algorithm with molecular dynamics simulation technique in identifying potential ligands for its OBP modeled via homology. Despite using a relatively very short equilibration time by current standards (700 ps), they achieved an excellent correlation between in silico and in vitro affinity data, showing that this association would be promising, including specific isoforms of OBPs, for other organisms.

Considering the particularities and objectives of virtual screening and molecular docking, both are important tools for: 1- Selecting molecules that act as pesticides on specific targets, seeking selectivity; 2- Elucidate classes of compounds with greater affinity for biological targets for subsequent in vitro/in vivo application; 3- Explain the molecular mechanism of action of compounds with previously determined activity. Examples of these applications in the context of pest control and management, extending to *C. capitata*, can be found in the works of Renthal (2024), Wang et al.

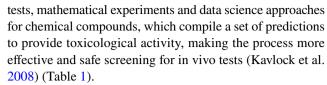


(2024), Liggri et al. (2023), Tiwari and Sowdhamini (2023) e Thireou et al. (2018).

In silico ADME-Tox parameters

ADMETox (Absortion, Distribution, Metabolism, Excretion and Toxicology) parameters for bioactive compounds are considered nowdays as extremely important, because they help to evaluate the viability and safety of a compound to be exposed to humans or animals before its introduction to the market (Yu and Adedoyin 2003). Knowledge about these parameters can save time and resources, avoiding the development of candidates for human, animal or environmental applications, preventing toxicity problems among others (Bandeira et al. 2023). Furthermore, it is estimated that research into the development of new products based on bioactive molecules entails costs in the order of millions of dollars, reinforcing the need to use methodologies that reduce the number of molecules that need to be evaluated on the bench (Tetko et al. 2006). Therefore, ADME-Tox parameters play a crucial role in the chemical products development process, contributing to the selection of more promising and safe candidates for humans and animals exposed (Yu and Adedovin 2003). It is a fact that the platforms and software dedicated to evaluating ADMET-Tox properties have human beings as their main target. However, considering the context of molecules applied to insect management, the alerts provided by these theoretical filters can be quite valuable in predicting situations involving humans and animals exposed to environmental products. Still in this sense, the construction of platforms dedicated to the prediction of physicochemical properties aimed specifically at the interaction with insects presents itself as a field that is still unexplored and capable of important contributions. Characteristics such as the external coating of certain family or considerations about the process of distribution through the hemolymph and insect metabolism could be explored in the construction of useful predictive models.

Here are some of the main reasons why these parameters can be important considering the possibility of human exposure: A (Absorption), evaluates the ability of a compound to be absorbed in the human body by analyzing whether it is effective or not; D (Distribution) indicates whether the compound reaches some type of tissue such as the ability to cross biological barriers, such as the blood–brain barrier; M (Metabolism) highlights whether the given compound will undergo transformation when metabolized; E (Excretion), refers to the elimination of the compound by the body; and Tox (Toxicity) assesses whether the compound is safe to apply, analyzing whether it promotes adverse side effects (Geerts and Heyden 2011). Thus, the evaluations of these parameters in silico are defined as the set of computational



The ADME-Tox properties (absorption, distribution, metabolism, excretion and toxicity) of the selected compounds are predicted using the online platforms pkCSM (Pires et al. 2015) or SwissADME (Daina et al. 2017), with the purpose to determine whether a given molecule will pose a risk to humans when handled in the laboratory or applied to the fruit after an in vivo test. For this, a simplified canonical system of molecular input lines (SMILES) is used to represent all chemical structures submitted to the platform (Yamari et al. 2023) (Table 2).

Molecular dynamics simulations

Molecular dynamics (MD) simulation is a computational technique that models the movement of atoms and molecules over time (Barbhuiya and Das 2023). This approach becomes particularly powerful when analyzing ligand-receptor interactions, taking into account both the flexibility of the ligand and the molecular target (Brooijmans and Kuntz 2003). Furthermore, it enables researchers to simulate the dynamic behavior, energy and interactions of individual atoms and molecules (Chen et al. 2018, 2023). This technique is classically based on the application of Newtonian laws, focusing on atomic movement over time, where the trajectory of molecules can be calculated based on variations in the energy and position of the atoms (Verli 2014). The behavior of each atom is analyzed based on the differential form of the equation of Newton's second law (Cunha 2013).

The Root Mean Square Deviation (RMSD) is a metric also frequently used in the area of computational chemistry, especially in molecular docking and molecular dynamics studies (Maruyama et al. 2023). In MD, RMSD is used to monitor how the conformation of a ligand changes over time during a simulation (Brüschweiler 2003). This can be valuable for understanding the flexibility of the ligand and how it adapts and stabilizes in the binding site over time (Verli 2014). The RMSD calculation is performed by the following formula: $RMSD = \sqrt{\frac{1}{N}\sum_{i=1}^{N}(r_i - r_i^0)^2}$, where \mathbf{r}_i are the positions of the atoms in the simulated structure, \mathbf{r}_i^0 are the corresponding positions in the reference structure, and N is the total number of atoms (Kuzmanic and Zagrovic 2010). The RMSD result is plotted as a function of time to evaluate the stability of the simulation (Maiorov and Crippen 1994). The graph may show initial phases of equilibration followed by a stable phase, or it may indicate significant fluctuations (Verli 2014). Small initial spikes in the RMSD can be



Table 1 The main semiochemical molecules that act in interfering with the behavior of organisms close to Ceratitis capitata

Molecule	Molecular formula Function	Function	Organisms	Behavior influenced	Synergy with other compounds	References
Benzyl benzoate	C14H12O2	Attractant	Ceratitis capitata	Increased attraction to males	No.	Menenses et al. (2020); El- Metwally et al. (2019)
		Attractant	Drosophila melanogaster	Attractive olfactory response	No	Khurana and Siddiqi (2013)
β-bisabolene	C15H24	Attractant	Anastrepha suspensa, A. ludens, A. obliqua	Attraction and behavior modification in larvae and females	Yes, with α-copaene	Rocca et al. (1992); Aluja et al. (2020); Lima-Medonça et al. (2014); Ponce et al. (1993); Ioannou et al. (2012)
$(+)$ - α -curcumene C15 H_{22}	$C15H_{22}$	Stimulant	Aedes aegypti	Antennal stimulation	No	Campbell (2009)
α-curcumene	C15H22	Attractant	Ceratitis capitata	Possible correlation with olfactory response	No	Liu et al. (2022)
Germacrene B	C15H ₂₄	Repellent, larvicide	Aedes aegypti, Drosophila suzukii	Repellent and larvicidal activity	No	Silva et al. (2015); Bedini et al. (2020)
β-longipinene	C15H ₂₄	Repellent	Aedes aegypti	Repulsive response in anten- nae	No	Zhang et al. (2011)
α-copaene	C15H ₂₄	Attractant	Ceratitis capitata	Superior attraction compared to other attractants	Yes, with β-bisabolene	Lull et al. (2023); Vaníčková et al. (2012a)
		Attractant	Anastrepha fraterculus, A. ludens, A. obliqua	Behavior modification and attraction	Yes, with β-bisabolene	Ruiz et al. (2021)
α-humulene	C15H ₂₄	Repellent/attractant	Aedes aegypti, Ceratitis capitata, Bactrocera correcta, A. fraterculus	Repellent activity and modification in mating behavior	Yes, with α-copaene	Silva et al. (2015); Shelly and Nishimoto (2015); Bachmann et al. (2015); Jaleel et al. (2019)
β-chamigrene	C15H24	Attractant/repellent	Ceratitis capitata, Aedes aegypti	Attraction to females, repellent activity	No	Antonatos et al. (2023); Carroll et al. (2011)
α-selinene	C15H24	Attractant/repellent	Ceratitis capitata, Aedes aegypti	Activation of antennal responses, moderate attraction	No	Silva et al. (2015); Tabança et al. (2020); Luu-Dam et al. (2021)
(-)-α-bisabolol	C15H260	Repellent	Bactrocera dorsalis, Aedes aegypti	Significant repellent activity	No	Jaffar and Lu (2022); Portilla- Pulido et al. (2020); Ali et al. (2023)



Table 2 Summary of the predicted functions by the platforms for each parameter

Parameter	PREDICTION FUNCTIONS
A (absorption)	Water solubility Caco2 permeability Intestinal absorption (human) Skin Permeability P-glycoprotein substrate P-glycoprotein I inhibitor P-glycoprotein II inhibitor
D (Distribution)	VDss (human) Fraction unbound (human) BBB permeability CNS permeability
M (Metabolism)	CYP2D6 substrate CYP3A4 substrate CYP1A2 inhibitior CYP2C19 inhibitior CYP2C9 inhibitior CYP2D6 inhibitior CYP2D6 inhibitior CYP3A4 inhibitior
E (Excretion)	Total Clearance Renal OCT2 substrate
TOX (Toxicity)	AMES toxicity Oral Rat Acute Toxicity (LD50) Max. tolerated dose (human) Hepatotoxicity Skin Sensitisation

expected during equilibration and if the RMSD stabilizes at a consistent value, the simulation is probably in equilibrium (Maruyama et al. 2023). Large continuous oscillations may indicate problems in the simulation (Shaw 2010).

It can be stated that MD seeks to reproduce, in the computational environment, conditions that are close to the natural environment at the macroscopic level (Gunsteren et al. 2006). Solvation is a crucial aspect to be considered in this type of simulation, since water plays a significant role in many biological reactions, influencing, for example, the hydration necessary for the functioning of some receptor proteins (Guerra et al. 2016). Thus, there is substantial relevance attributed to computer simulation models to reproduce the biological environment more accurately (Berendsen et al. 1984), offering insights into mechanical, thermal and chemical properties, as well as the effects of external factors, such as temperature and pressure (Zhang et al. 2022).

However, it is essential to note that MD simulations have limitations, requiring precise force field parameters, adequate computational resources and appropriate modeling assumptions (Barbhuiya and Das 2023). One of the main limitations lies in the inability to analyze ligands outside of a local minimum energy state (Brooijmans and Kuntz 2003). This considerably restricts the events that DM can analyze, especially when observing the behavior of the ligand in different temperature ranges, making it sensitive and dependent

on robust energy minimization analysis algorithms (Brooijmans and Kuntz 2003).

One of the algorithms used to more accurately demonstrate the stability of the formed complexes is the application of the Molecular Mechanics Poisson-Boltzmann surface area method (MM-PBSA) (Xu et al. 2023). MM/PBSA is a method that calculates the binding energy between a protein and a ligand (Genheden and Ryde 2015), evaluating the energy terms of the DM force field and approximating the polar energy terms with the Poisson–Boltzmann (PB) equation ($G = E_{bond} + E_{VdW} + E_{ele} + G_{pol} + G_{np} - T.S$), and non-polar energy terms using the Solvent Accessible Surface Area (SASA) method (Gallo et al. 2022). This last algorithm makes it possible to identify how the surface area of a protein interacts with water molecules (solvent) (Mazola et al. 2015).

Protein obtention, softwares and platforms in structure based molecular design

In silico methods are computational techniques that can be used to identify potential ligands for OBPs. Some platforms can be used to model or find deposited receptor structures, such as PDB (Protein Data Bank) and UniProt, which provide 3D structures of important target proteins (Berman et al. 2000; Wang et al. 2021). Homology modeling is another widely used technique, based on the premise that proteins with similar structures have similar functions (Hameduh et al. 2020). In this context, known olfactory proteins are used as models to build structures of new proteins of interest, comparing sequences with databases of proteins with known structures (Alfonso-Prieto and Capelli 2023). Tools such as MODELLER (Sali and Blundell 1993) and Swiss-Model (Waterhouse et al. 2018) are often used to predict the three-dimensional structures, facilitating the assessment of their odorant-binding capacity based on structural similarities. Alternatively, the structure can be modeled using AlphaFold 2 (https://alphafold.ebi.ac.uk/), platform that revolutionized the prediction of 3D protein structures, with amino acid sequences obtained from NCBI (Jumper et al. 2021).

Phylogenetic analysis is also a valuable method, comparing protein sequences between different species to identify potential odorant targets, exploring the evolutionary relationship and identifying groups of receptors with conserved functions. Tools such as Clustal Omega (Sievers et al. 2011) and MEGA (Tamura et al. 2021) are used to build phylogenetic trees and perform comparative analyses. Furthermore, machine learning and neural network methods have been gaining prominence in bioinformatics to predict interactions between proteins and ligands, using large data sets to efficiently identify new interactions (Dhakal et al. 2022;



Lemkul 2018). After training the models, they can identify new odorant-binding proteins based on recognized patterns, accelerating the discovery of interactions without the need for time-consuming laboratory experiments (Guo and Yamaguchi 2022).

Accuracy in protein structural determination is extremely important, since the 3D conformation and binding sites of the target molecules are necessary for the correct anchoring of potential ligands (Rodrigues et al. 2012). Techniques such as X-ray crystallography, Nuclear Magnetic Resonance (NMR), and cryo-electron microscopy (Cryo-EM) are responsible for assembling these structures with high resolution and reliability, allowing their use for screening and docking processes (Berman et al. 2000). X-Ray crystallography is the most widely used technique for determining protein structures (Ambrosio and Franchini 2017; Shi 2014). Analysis of the diffraction pattern generated by the passage of X-rays through the crystal allows the reconstruction of the atomic structure with high precision, and good resolutions are considered as smaller than 2 Å (Feiten 2022). In addition, NMR (Nuclear Magnetic Resonance) is also used to provide structural information on proteins in solution, but with a lower resolution than X-ray crystallography (Figueiredo and Marsaioli 2007; Cavanagh et al. 2007). The geometric information used in NMR to solve the three-dimensional (3D) structures of proteins resides in short interproton distance data (less than 5 Å) (Oschkinat et al. 1988). Cryo-EM (Cryo-Electron Microscopy), in turn, is a powerful method for resolving the three-dimensional structures of biological macromolecules, mainly due to technological advances that have made it possible to achieve lower resolutions (Yip et al. 2020). One of the main advantages of this technique is that it allows the study of larger protein complexes (Sawh-Gopal et al. 2023).

In contrast, in silico methods based on structure predictions, generated by homology modeling based on amino acid sequences (SwissModel, for example), have some limitations. These models generally present uncertainties in some fundamental regions, such as binding sites or the unreliability of amino acid rotation and arrangement, which can consequently result in incorrect interactions (Jumper et al. 2021). This can end up compromising the virtual screening process, since these limitations can influence the accuracy of molecular docking simulations, leading to the identification of false positive or non-biologically active compounds (Cheng et al. 2007). Therefore, the use of three-dimensional proteins already available in databases and obtained by the techniques mentioned above becomes more reliable to offer a solid basis for the identification of bioactive compounds (Dror et al. 2012; Jumper et al. 2021).

After obtaining the three-dimensional structure, the prediction of binding sites can be performed. There are algorithms capable of identifying probable odor binding sites, or the technique for experimental determination of the macromolecule already considers a standard complexed ligand. For example, tools such as SiteHound (Hernandez et al. 2009), GRaSP (Santana et al. 2020) and FPocket (Guilloux et al. 2009) analyze the three-dimensional structure of proteins to identify cavities that can serve as binding sites for molecular docking studies (Morris et al. 2009).

Once the protein structure has been obtained, using a completely in silico method or with the aid of experimental techniques, its subsequent processing can be carried out. Programs such as Chimera software (Petersen et al. 2004) can be used, for example, to remove water molecules from experimental techniques, if necessary. The virtual platform ABPS (Adaptive Poisson–Boltzmann Solver), in turn, can perform additional electrostatic analysis, adjusting the pH media and aminoacid protonation states for a more realistic simulation (Lee et al. 2002; Brito et al. 2016a; Jurrus et al. 2017).

A step towards validating docking strategies based on the structure of macromolecular targets is the obtention of the ROC (Receiver Operating Characteristic Curve) curve. For example, the Drug Design online platform (http://stats. drugdesign.fr/) is a free tool that can be used in the molecular modeling field. The ROC curve is a validation process, that indicate if a macromolecule and the docking algorithm used for screening has the capacity to differentiate molecules that promote a real biological response from compounds that bind and do not promote any type of response (Emperieur-Mot et al. 2016; Davis and Goadrich 2006). Therefore, true positive ligands, molecules that bind at the binding site and promote behavioral responses, must be initially identified through a literature review (Kim et al. 2023). These molecules may already come with their structure optimized to a minimal energy or this geometry can be obtained in programs such as ArgusLab or PyRx (Dallakyan and Olson 2014; Thompson 2004). Based on these defined true ligands, a library of, for example, 50 decoys (false positives) for each true ligand must be assembled using, for example, the DUD-E platform and after fitting all these false positives, 10 decoys corresponding to each real ligand, randomly selected, will be used to build the ROC curve (Mysinger et al. 2012). The AUC (Area Under the Curve) value closest to 1, obtained after building the curve, indicates that the fitting model has an excellent ability to differentiate true positive ligands from false positives (Emperieur-Mot et al. 2016). The molecular docking parameters for the aforementioned step and for the rest of the virtual screening can be defined in docking programs interface (Morris et al. 2009) and the docking itself can be performed. Softwares and platforms as AutoDock, Vina (Eberhardt et al. 2021; Trott and Olson 2009), SwissDock (Grosdidier et al. 2011) and DockThor (Guedes et al. 2024), are examples widely used.



Considering specifically the molecular docking in the virtual screening process, online molecular databases make this process possible. Some examples of databases also considering the specific interaction with odorant binding proteins, are: NuBBE (Nucleus of Bioassays Biosynthesis and Ecophysiology of Natural Products), which was created with the aim of bioprospecting new natural molecules with antifungal, antitumor, antioxidant, anticholinesterase, anti-inflammatory and antiparasitic potential (Pilon et al. 2017). The ZINC database, which is a free database with more than 230 million compounds in 3D format, ready for docking (Irwin and Shoichet 2005). LOTTUS, which is a database for storage, research and analysis of Natural Products (NPs), currently contains more than 200 thousand molecules extracted from plants (Rutz et al. 2022). Essential Oil (EssOilDB) and Essential Oil University (EOUDB), which are databases of molecules isolated from essential oils (Kumari et al. 2014a, b; Pappas 2024), and Pherobase, a platform composed of several databases that provide comprehensive information on more than 6500 pheromones and semiochemical compounds (El-Sayed 2024). In addition, one of the best known and most used is PubChem, operated and maintained by the National Center for Biotechnology Information (Kim et al. 2016).

Within the specificity of communication by semiochemicals, after virtual screening, the molecules can be selected as possible candidates based on criteria such as lowest binding energy and vapor pressure, which indicate the compound as volatile, an important character to interact with OBP (Mourão 2021; Gomes et al. 2022). The calculation of enthalpy of vaporization, boiling point and vapor pressure values of the most promising molecules found in each database can be performed (Pence and Williams 2010; Mucha et al. 2023). The visualization of the interactions for the most promising compounds from screening, complexed with target, can be made by softwares as BIOVIA Discovery Studio, PyMol and LigPlot+, that provide 2D diagrams of the interactions between the atoms of the ligands with amino acid residues of the protein, as well as can demonstrate this fit in a three-dimensional format (Schrödinger and DeLano 2020; BIOVIA 2015; Wallace et al. 1996).

Considering the Molecular Dynamics procedures, some of the main software used are: GROMACS 5.1.2 (Hess et al. 2008; Abraham et al. 2015), AMBER (Pearlman et al. 1995), CHARMM (Brooks et al. 2009), and GPUGRID (Rodríguez-Espigares et al. 2020). Below we will describe a minimal step-by-step process that can later be detailed and explored for a basic molecular dynamics routine, as per the guidelines in our laboratory.

The topological parameters for ligands, necessary for running molecular dynamics, can be acquired through online platforms, such as ATB (Automated Topology Builder) (Stroet et al. 2018). ATB is a web-accessible server that

provides topologies and parameters for a wide variety of molecules, ideal for application in molecular simulations, computational design of compounds, and refinement of structures obtained by X-ray diffraction (Malde et al. 2011). Another procedure necessary in the dynamic simulation, often neglected by docking, is the solvation. The complexes are solvated with water molecules in a box (cubic, monoclinic, orthorombic, etc.), considering periodic boundary conditions to treat the surface problem (Berendsen et al. 1984). This method allows the simulation of a more realistic biological aqueous environment and closer to the real interaction situation through the influence of water molecules in the system (Jorgensen et al. 1983; Namba et al. 2008).

Also within the simulation, the Particle Mesh Ewald algorithm is used as another parameter, in this case, to treat long-range electrostatic interactions (Darden et al. 1993). The method requires charge neutrality of the molecular system to accurately calculate the Coulomb interaction, which consists of the electrostatic interaction between electrically communicated particles (Kolafa and Perram 1992). The bond lengths can be maintained in equilibrium using the P-LINCS algorithm (Hess et al. 2008). This method is called parallel linear constraint solver (P-LINCS), which allows the restriction of all bonds in macromolecules, protecting the energy conservation properties and promoting improvements in the restriction of decoupled angle constraints (Hess 2007). It is worth noting that this entire system must be carried out by controlling energy minimization, through the application of other joint algorithms such as NVT (Constant temperature, constant volume) and NPT (Constant temperature, constant pressure), in conjunction with the modified Berend-sen thermostat and a Parrinello-Rahman barostat, to control pressure and temperature during the simulation (Parrinello and Rahman 1981). In detail, Berend-sen consists of an algorithm to resize particle velocities in molecular dynamics to control the simulation temperature (Berendsen et al. 1984). While the Parrinello-Rahman barostat considers that the inertia tensor of the simulation cell is spherical and constant in time, considering the irrotational fluctuation movements of the cell (Podio-Guidugli 2010).

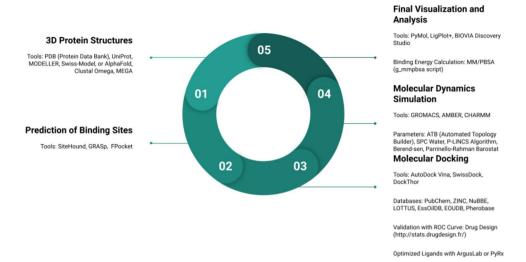
Finally, the average value of the binding energy for all complexes can be estimated calculating the free energies of the monomers and the ligand–protein complex in each stage submitted to the simulation (Kollman et al. 2000). For example, the MM/PBSA method can be used through the 'g_mmpbsa' script (Kumari et al. 2014a) (Fig. 4).

In vivo, in vitro, in silico perspectives

Techniques for testing repellent or attractive compounds in fruit flies, such as *Ceratitis capitata*, include laboratory (in vitro) and field (in vivo) experiments. In the laboratory, flies



Fig. 4 Diagram of the main stages of OBP identification, Docking, Virtual Screening, and Molecular Dynamics simulation using computational methods



are exposed to different concentrations of the compounds in controlled boxes, called choice boxes or wind tunnels, in order to observe their behavior in response to the compounds (Scolari et al. 2021). Confirmation of whether or not attraction occurred, or whether the compound was repellent or not, is assessed by the amount of time and frequency of the flies' visits to the sources of the molecules (Tabanca et al. 2019). In field experiments, the compounds are applied in traps or devices, and effectiveness is measured by the capture of flies, as well as by proven changes in the population in areas that were treated versus those that were not (Manoukis et al. 2015). Therefore, it can be said that these methods make it possible to determine the attractive or repellent properties of compounds in pest management.

There are some limitations associated with the field use of the compounds as discussed in this review, one of which is the short period of protection provided by the molecule, especially when its volatility is high and the application is directly on the fruit (Thireou et al. 2018). To overcome this issue, it is important to develop slow release formulations (Ghayempour and Montazer 2016), add fixatives to extend the protection time (Tawatsin et al. 2001) and create devices for continuous spatial release (Dame et al. 2014), which represent potential modifications to extend the effectiveness of these molecules. A strategy proposed by Drapeau et al. (2009) to increase repellent activity over time involves the use of microemulsions with surfactants associated with highly volatile compounds against *A. aegypti*.

It is important to highlight that, depending on the type of compound or activity provided, synergism between molecules is a crucial factor to be considered in the management of *C. capitata*. Mixtures made with up to six compounds imitate the actions of pheromones, which are also made up of a set of molecules (Milet-Pinheiro et al. 2015). This alternative is even more effective when volatiles are combined with known pheromone constituents, as the

synergism between these semiochemicals has been shown to produce more efficient mixtures in controlling insect pests (Reddy and Guerrero 2004a). The synergism of mixtures, unlike the individual application of compounds, promotes a higher level of perception, achieved only by the joint action of these volatiles, forming what researchers call a "complete bouquet" that is detected by the insect, inducing the desired behavior when a mixture is applied (Cruz-López et al. 2015a).

Furthermore, each mixture is interpreted as a new signal, so the addition or removal of a component from the formulation can result in a new mixture, triggering a completely different behavioral response from the previous one (Beyaert et al. 2010). Therefore, it is clear that natural compounds are not ephemeral, and the chemistry behind the joint formulation of these molecules is crucial to prolong the effectiveness of control and ensure its total effectiveness (Carroll et al. 2011). In some cases, as mentioned in the studies above, it is also possible to test the use of the compound in isolation, which can vary significantly depending on the type of assay used for these analyzes (Jaffar and Lu 2022; Ali et al. 2023).

The insect's great capacity for chemical recognition makes the in silico exploration of molecular criteria for ligand-target affinity a challenge and, at the same time, a fruitful way to identify interaction patterns and propose new promising scaffolds. Furthermore, chemical characteristics intrinsic to each compound as well as blends of molecules with known attractive or repellent properties can be studied through artificial intelligence and pattern recognition algorithms, generating models with orthogonal variables that can be useful in the development of products for pest management.

Author contributions All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by Ramiro Guimarães Passos, Edilson Beserra de Alencar Filho and Vanessa Costa Santos. The first draft of the manuscript was



written by Ramiro Guimarães Passos and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript."

Funding This work was supported by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq); Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES); Centro Nacional de Processamento de Alto Desempenho da UFC (CENAPAD-UFC); Author R.G.P. has received research support from Fundação de Amparo à Pesquisa do Estado de Pernambuco (FACEPE);

Data availability Not applicable.

Declarations

Conflict of interest The authors have no relevant financial or non-financial interests to disclose.

References

- Abraham M, Murtola T, Schulz R, Pall S (2015) GROMACS: High performance molecular simulations through multi-level parallel-ism from laptops to supercomputers. SoftwareX 1:19–25. https://doi.org/10.21105/joss.00608
- Adams M, Celniker S, Holt R, Evans C et al (2000) The genome sequence of *Drosophila melanogaster*. Science 287(5461):2185–2195. https://doi.org/10.1126/science.287.5461.2185
- Al-Jalely BH, Xu W (2021) Olfactory sensilla and olfactory genes in the parasitoid wasp *Trichogramma pretiosum* Riley (Hymenoptera: Trichogrammatidae). InSects 12(11):998. https://doi.org/ 10.3390/insects12110998
- Alfaro C, Vacas S, Zarzo M, Navarro-Llopis V, Primo J (2011) Solid phase microextraction of volatile emissions of *Ceratitis capitata* (Wiedemann) (Diptera: Tephritidae): influence of fly sex, age, and mating status. J Agric Food Chem 59:298–330. https://doi.org/10.1021/jf104183c
- Alfonso-Prieto M, Capelli R (2023) Machine learning-based modeling of olfactory receptors in their inactive state: human OR51E2 as a case study. J Chem Inf Model 63(10):2911–2917. https://doi.org/10.1021/acs.jcim.3c00380
- Ali A, Tabanca N, Raman V, Avonto C et al (2023) Chemical composition of essential oils from German, Roman, and Chinese chamomile flowers and their biological activities against three economically important insects. Rec Nat Prod 17(4):595–614
- Alonso H, Bliznuk A, Greadym J (2006) Combining docking and molecular dynamic simulations in drug design. Med Res Rev 26(5):531–568
- Aluja M, Mangan RL (2008a) Fruit fly (Diptera: Tephritidae) host status determination: critical conceptual, methodological, and regulatory considerations. Annu Rev Entomol 53(1):473–502. https://doi.org/10.1146/annurev.ento.53.103106.093350
- Aluja M, Cabagne G, Altúzar-Molina A, Pascacio-Villafañ C et al (2020) Host plant and antibiotic effects on scent bouquet composition of Anastrepha ludens and Anastrepha obliqua calling males, two polyphagous tephritid pests. InSects. https://doi.org/ 10.3390/insects11050309
- Ambrosio ALB, Franchini KG (2017) Cristalografia macromolecular: a biologia sob a ótica dos raios X. Ciênc Cult 69(3):45–49. https://doi.org/10.21800/2317-66602017000300009
- Antonatos S, Anastasaki E, Balayiannis G, Michaelakis A et al (2023) Identification of volatile compounds from fruits aroma and citrus essential oils and their effect on oviposition of *Ceratitis capitata*

- (Diptera: Tephritidae). Environ Entomol 52(3):327–340. https://doi.org/10.1093/ee/nvad024.short
- Arioli C, Botton M, Mafraneto A, Molinari F et al (2013) Feromônios sexuais no manejo de insetos-praga na fruticultura de clima temperado. Empresa Pesqui Agropecu Ext Rural Santa Catarina: Bol Téc 159:1–58
- Arredondo J, Díaz-Fleischer F, Pérez-Staples D (2010) Biología y Comportamiento. In: Montoya P, Hernández E (eds) Moscas de la fruta: fundamentos y procedimientos para su manejo. IICA, México, DF, pp 91–106
- Bachmann G, Segura D, Devescovi F, Juárez L et al (2015) Male sexual behavior and pheromone emission is enhanced by exposure to guava fruit volatiles in *Anastrepha fraterculus*. PLoS ONE. https://doi.org/10.1371/journal.pone.0124250
- Baker R, Herbert R, Grant G (1985) Isolation and identification of the sex pheromone of the Mediterranean fruit fly, *Ceratitis capitata* (Wied.). J Chem Soc Chem Commun. https://doi.org/ 10.1039/c39850000824
- Bandeira L, Pinto L, Carneiro C (2023) Pharmacometrics: the already-present future of precision pharmacology. Ther Innov Regul Sci 57(1):57-69. https://doi.org/10.1007/s43441-021-00238-7
- Barbhuiya S, Das B (2023) Molecular dynamics simulation in concrete research: a systematic review of techniques, models and future directions. J Build Eng 76:10301447. https://doi.org/10.1016/j.jobe.2023.10301447
- Bedini S, Cosci F, Tani C, Pierattini E et al (2020) Essential oils as post-harvest crop protectants against the fruit fly *Drosophila suzukii*: bioactivity and organoleptic profile. InSects 11(8):508
- Belliard S, Bachmann G, Fernández P, Hurtado J et al (2022) Identification of host plant volatile stimulants of *Anastrepha fraterculus* male courtship behavior. Frontiers. https://doi.org/10.3389/fevo.2022.943260
- Bender BJ, Gahbauer S, Luttens A et al (2021) A practical guide to large-scale docking. Nat Protoc 16(11):4799–4832. https://doi.org/10.1038/s41596-021-00597-z
- Benton R, Vannice KS, Gomez-Diaz C, Vosshall LB (2009) Variant ionotropic glutamate receptors as chemosensory receptors in Drosophila. Cell 136(1):149–162. https://doi.org/10.1016/j.cell.2008.12.001
- Berendsen H, Postma J, Gunsteren W, DiNola A et al (1984) Molecular dynamics with coupling to an external bath. J Chem Phys 81:3684–3690. https://doi.org/10.1063/1.448118
- Berman H, Westbrook J, Feng Z, Gilliland G et al (2000) The protein data bank. Nucleic Acids Res 28(1):235–242. https://doi.org/10.1093/nar/28.1.235
- Beyaert I, Wäschke N, Scholz A, Varama M et al (2010) Relevance of resource-indicating key volatiles and habitat odour for insect orientation. Anim Behav 79(5):1077–1086. https://doi.org/10.1016/j.anbehav.2010.02.001
- Biessmann H, Andronopoulou E, Biessmann M, Douris V et al (2010) The *Anopheles gambiae* odorant binding protein 1 (AgamOBP1) mediates indole recognition in the antennae of female mosquitoes. PLoS ONE. https://doi.org/10.1371/journ al.pone.0009471
- Biolchini M, Murru E, Anfora G, Loy F et al (2017) Fat storage in Drosophila suzukii is influenced by different dietary sugars in relation to their palatability. PLoS ONE. https://doi.org/10.1371/journal.pone.0183173
- BIOVIA (2015) Discovery Studio Modeling Environment. Dassault Syst. Release, San Diego, 4
- Bitencourt-Ferreira G, de Azevedo WF Jr (2019) Molegro virtual docker for docking. Methods Mol Biol 2053:149–167. https://doi.org/10.1007/978-1-4939-9752-7_10
- Bittencourt MAL, Cova AKW, Silva ACM, Silva VES et al (2006) Espécies de moscas-das-frutas (Tephritidae) obtidas em



- armadilhas McPhail no Estado da Bahia, Brasil. Semin Ciênc Agrár 27(4):561-564
- Breer H (2003) Olfactory receptors: molecular basis for recognition and discrimination of odors. Anal Bioanal Chem 377(3):427–433. https://doi.org/10.1007/s00216-003-2113-9
- Brito N, Moreira M, Melo A (2016a) A look inside odorant-binding proteins in insect chemoreception. J Insect Physiol 95:51–65. https://doi.org/10.1016/j.jinsphys.2016.06.003
- Břízová R, Mendonça A, Vaníčková L, Lima-Mendonça A et al (2013) Pheromone analyses of the *Anastrepha fraterculus* (Diptera: Tephritidae) cryptic species complex. Fla Entomol 96:1107–1115. https://doi.org/10.1653/024.096.0346
- Brooijmans N, Kuntz I (2003) Molecular recognition and docking algorithms. Annu Rev Biophys Biomol Struct 32:335–373. https://doi.org/10.1146/annurev.biophys.32.110601.141803
- Brooks B, Brooks C, Mackerell A, Nilsson L et al (2009) CHARMM: the biomolecular simulation program. J Comput Chem 30(10):1545–1614. https://doi.org/10.1002/jcc.21287
- Bruce TJ, Pickett JA (2011) Perception of plant volatile blends by herbivorous insects: finding the right mix. Phytochemistry 72(13):1605–1611
- Brüschweiler R (2003) Efficient RMSD measures for the comparison of two molecular ensembles: root-mean-square deviation. Proteins 50(1):26–34. https://doi.org/10.1002/prot.10286
- Bunker C, Hirschfelder A (1925a) Mosquito repellents. Am J Trop Med Hyg 5:359–383. https://doi.org/10.5555/19251000617
- Cáceres C, Segura D, Vera M, Wornoayporn V et al (2009) Incipient speciation revealed in *Anastrepha fraterculus* (Diptera: Tephritidae) by studies on mating compatibility, sex pheromones, hybridization, and cytology. Biol J Lin Soc 97:152–165. https://doi.org/10.1111/j.1095-8312.2008.01154.x
- Camelo LA, Landolt PJ, Zack RS (2007) A kairomone based attractand-kill system effective against alfalfa looper (Lepidoptera: Noctuidae). J Econ Entomol 100:366–374
- Campanacci V, Krieger J, Bette S, Sturgis JN, Lartigue A, Cambillau C, Breer H, Tegoni M (2001) Revisiting the specificity of *Mamestra brassicae* and *Antheraea polyphemus* pheromone-binding proteins with a fluorescence binding assay. J Biol Chem 276(23):20078–20084. https://doi.org/10.1074/jbc. M100713200
- Campbell C (2009) Analyses of essential and edible oils, and constituents therein, as candidate repellents for the yellow fever mosquito *Aedes aegypti* L. (Diptera: Culicidae) (Master of Pest Management). https://summit.sfu.ca/_flysystem/fedora/sfu_migrate/9858/ETD4904.pdf
- Carey A, Carlson J (2011) Insect olfaction from model systems to disease control. Proc Natl Acad Sci 108(32):12987–12995. https://doi.org/10.1073/pnas.1103472108
- Carroll J, Tabanca N, Kramer M, Elejalde N et al (2011a) Essential oils of *Cupressus funebris*, *Juniperus communis*, and *J. chinensis* (Cupressaceae) as repellents against ticks (Acari: Ixodidae) and mosquitoes (Diptera: Culicidae) and as toxicants against mosquitoes. J Vector Ecol 36(2):258–268
- Carvalho RDS, Nascimento AS (2002) Criação e utilização de Diachasmimorpha longicaudata para controle biológico de moscas-dasfrutas (Tephritidae). In: Parra JRP et al (eds) Controle Biológico no Brasil: Parasitoides e Predadores. Editora Manole, Barueri, pp 165–179
- Cavanagh J, Fairbrother WJ, Palmer AG, Skelton NJ (2007) Protein NMR spectroscopy: principles and practice, 2nd edn. Academic Press
- Cha DH et al (2012) Effect of apple cultivar, maturity, and phagostimulants on the attraction of apple maggot fly, *Rhagoletis pomonella*, to synthetic apple volatiles. J Chem Ecol 38(4):319–329
- Chang H, Liu Y, Yang T, Pelosi P et al (2015) Pheromone binding proteins enhance the sensitivity of olfactory receptors to sex

- pheromones in *Chilo suppressalis*. Sci Rep. https://doi.org/10.1038/srep13093
- Chen Z, Pei J, Li R, Xiao F (2018) Performance characteristics of asphalt materials based on molecular dynamics simulation: a review. Constr Build Mater 189(6):695–710. https://doi.org/10. 1016/j.conbuildmat.2018.08.150
- Chen J, Chu X, Cheng Y, Shu Q et al (2023) Application of molecular dynamics simulations in interface interaction of clay: current status and perspectives. Asia-Pac J Chem Eng 18(5):e2955. https://doi.org/10.1002/apj.2955
- Cheng Y (2015) Single-particle cryo-EM at crystallographic resolution. Cell 161(3):450–457. https://doi.org/10.1016/j.cell.2015.03.049
- Cheng TM, Blundell TL, Fernandez-Recio J (2007) pyDock: Electrostatics and desolvation for effective scoring of rigid-body protein–protein docking. Proteins Struct, Funct Bioinform 68(2):503–515. https://doi.org/10.1002/prot.21419
- Christophers S (1947) Mosquito repellents; being a report of the work of the mosquito repellent inquiry, Cambridge, 1943–5. J Hyg 45(2):176–231
- CONAB (Companhia Nacional de Abastecimento) (2023) Boletim Hortigranjeiro, 9(6). Retrieved from file:///C:/Users/Compaq/ Documents/Boletim-Hortigranjeiro-Junho-2023.pdf
- Cossé A, Todd J, Millar J, Martínez L et al (1995) Electroantennographic and coupled gas chromatographic-electroantennographic responses of the Mediterranean fruit fly, *Ceratitis capitata*, to male-produced volatiles and mango odor. J Chem Ecol 21:1823– 1836. https://doi.org/10.1007/BF02035168
- Costa A, Amorim F, Anjos-Duarte C, Joachim-Bravo I (2011) Influence of different tropical fruits on biological and behavioral aspects of the Mediterranean fruit fly *Ceratitis capitata* (Wiedemann) (Diptera, Tephritidae). Rev Bras Entomol 55(3):355–360
- Cruz-López L, Malo E, Rojas J (2015a) Sex pheromone of *Anastre-pha striata*. J Chem Ecol 41(5):458–464. https://doi.org/10.1007/s10886-015-0599-7
- Cunha RA (2013) Caracterização molecular do mecanismo de interação de quitosanas com bicamadas lipídicas compostas de dipalmitoilfosfatidilcolina (Dissertação de Mestrado). Instituto de Química, Universidade Federal de Uberlândia, Uberlândia MG. https://bdtd.ibict.br/vufind/Record/UFU_664c8b329923973 70a08ab67a6dac05c
- Daina A, Michielin O, Zoete V (2017) SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. Sci Rep. https://doi.org/ 10.1038/srep42717
- Dallakyan S, Olson A (2014) Small-molecule library screening by docking with PyRx. Methods Mol Biol 1263:243–250. https://doi.org/10.1007/978-1-4939-2269-7_19
- Dame D, Meisch M, Lewis C, Kline D et al (2014) Field evaluation of four spatial repellent devices against Arkansas rice-land mosquitoes. J Am Mosq Control Assoc 30:31–36
- Darden T, York D, Pedersen L (1993) Particle mesh Ewald: an N●log(N) method for Ewald sums in large systems. J Chem Phys 98:10089–10092. https://doi.org/10.1063/1.464397
- Davis J, Goadrich M (2006) The relationship between Precision-Recall and ROC curves. In: Proceedings of the 23rd International Conference on Machine Learning (ICML 2006), pp 233–240
- De Vivo M, Masetti M, Bottegoni G, Cavalli A (2016) Role of molecular dynamics and related methods in drug discovery. J Med Chem 59(9):4035–4061
- Degenaro M, McBride C, Seeholzer L, Nakagawa T et al (2013) orco mutant mosquitoes lose strong preference for humans and are not repelled by volatile DEET. Nature 498(7455):487–491
- Dekker T, Ignell R, Ghebru M, Glinwood R et al (2011) Identification of mosquito repellent odours from *Ocimum forskolei*. Parasites Vect. https://doi.org/10.1186/1756-3305-4-183



- Dhakal A, McKay C, Tanner JJ, Cheng J (2022) Artificial intelligence in the prediction of protein-ligand interactions: recent advances and future directions. Brief Bioinform 23(1):bbab476. https://doi.org/10.1093/bib/bbab476
- Dias-Pini N, Maciel G, Souza A, Nascimento J et al. (2022) Incidência de Ceratitis capitata (Wiedemann) (Diptera: Tephritidae) e Parasitoides Associados em Frutos de Acerola, Malpighia emarginata DC, em Paraipaba, CE. Empresa Brasileira de Pesquisa Agropecuária. Retrieved from https://www.infoteca.cnptia.embra pa.br/infoteca/bitstream/doc/1147134/1/BP-235.pdf
- Díaz-Fleischer F, Aluja M (2003) Influence of conspecific presence, experience, and host quality on oviposition behavior and clutch size determination in *Anastrepha ludens* (Diptera: Tephritidae).
 J Insect Behav 16:537–554. https://doi.org/10.1023/A:10273 07424150
- Ditzen M, Pellegrino M, Vosshall L (2008) Insect odorant receptors are molecular targets of the insect repellent DEET. Science 319(5871):1838–1842
- dos Santos JAN et al (2007) Fruticultura nordestina: desempenho recente e possibilidades de políticas. Banco do Nordeste do Brasil
- Drapeau J, Verdier M, Touraud D, Krockel U et al (2009) Effective insect repellent formulation in both surfactantless and classical microemulsions with a long-lasting protection for human beings. Chem Biodivers 6(6):934–947. https://doi.org/10.1002/cbdv.200800225
- Dror RO, Dirks RM, Grossman JP, Xu H, Shaw DE (2012) Biomolecular simulation: a computational microscope for molecular biology. Annu Rev Biophys 41:429–452. https://doi.org/10.1146/annurev-biophys-042910-155245
- Duarte A, Malavasi A (2000) Tratamentos quarentenários. In: Malavasi A, Zucchi RA (eds) Moscas-das-frutas de importância econômica no Brasil: conhecimento básico e aplicado. Holos, Ribeirão Preto, pp 187–192
- Eberhardt J, Santos-Martins D, Tillack A, Forli S (2021) AutoDock Vina 1.2.0: new docking methods, expanded force field, and python bindings. J Chem Inf Model 61(8):3891–3898. https://doi.org/10.1021/acs.jcim.1c00501
- El-Metwally M, Mostafa M, Ghanim N (2019) Effect of trimedlure diluted with certain oils against Mediterranean fruit fly, *Ceratitis capitata* males under filed conditions. J Entomol Zool Stud 7(2):326–332
- El-Sayed A (2024) The pherobase: database of pheromones and semiochemical. Pherobase. https://www.pherobase.net
- Emperieur-Mot C, Zagury J, Montes M (2016) Screening exploreran interactive tool for the analysis of screening results. J Chem Inf Model 56(12):2281–2286. https://doi.org/10.1021/acs.jcim. 6b00317
- European and Mediterranean Plant Protection Organization (2022) *Ceratitis capitata* distribution. https://gd.eppo.int/taxon/CERTCA/distribution. Accessed 19 Jan 2024
- Falchetto M, Ciossani G, Scolari F, Cosimo A et al (2019) Structural and biochemical evaluation of *Ceratitis capitata* odorant-binding protein 22 affinity for odorants involved in intersex communication. Insect Mol Biol 28:431–443. https://doi.org/10.1111/imb. 12559
- Fan J, Francis F, Liu Y, Chen J et al (2011) An overview of odorantbinding protein functions in insect peripheral olfactory reception. Genet Mol Res 10:3056–3069
- Farneti B et al (2017) Impact of volatile compounds on the sensory perception of apple quality: a review. Trends Food Sci Technol 69:266–274
- Fazenda L (2023) Influência de moscas-das-frutas (Diptera: Tephritidae) na produtividade de pimenta malagueta, no semiárido Moçambicano. Rev Verde 18(1):17–23

- Fein BL, Reissig WH, Roelofs WL (1982) Identification of apple volatiles attractive to the apple maggot fly. J Chem Ecol 8:1473–1487. https://doi.org/10.1007/BF00987690
- Feiten MC (2022) A determinação de estruturas tridimensionais de enzimas pela cristalografia de raios-X. Ciênc Exatas Tecnol 9(2). https://orcid.org/0000-0001-9522-7168
- Ferreira R, Oliva G, Andricopulo A (2011) Integração das técnicas de triagem virtual e triagem biológica automatizada em alta escala: oportunidades e desafios em P&D de fármacos. Quím Nova 10. Retrieved from https://www.scielo.br/j/qn/a/wCtk9GFDMs rNCpScP8kn8Jr/
- Figueiredo IM, Marsaioli AJ (2007) Mapeamento das interações proteína-ligante através de técnicas de RMN de ¹H utilizando detecção do ligante. Quim Nova 30(7):1597–1605. https://doi.org/10.1590/S0100-40422007000700019
- Fraser AM, Mechaber WL, Hildebrand JG (2003) Electroantennographic and behavioral responses of the sphinx moth *Manduca sexta* to host plant headspace volatiles. J Chem Ecol 29:1813–1833. https://doi.org/10.1023/A:1024898127549
- Gadenne C, Barrozo R, Anton S (2016) Plasticity in insect olfaction: to smell or not to smell? Annu Rev Entomol 61:317–333. https://doi.org/10.1146/annurev-ento-010715-023523
- Gallo D, Nakano O, Silveira Neto S, Carvalho RDL, Batista GC, Berti Filho E, Omoto C (2002) Entomologia Agrícola. FEALQ, Piracicaba
- Gallo G, Barcick U, Coelho C, Salardani M et al (2022) A proteomics-MM/PBSA dual approach for the analysis of SARS-CoV-2 main protease substrate peptide specificity. Peptides. https://doi.org/10. 1016/j.peptides.2022.105140
- Garson L, Winnike M (1968) Relationships between insect repellency and chemical and physical parameters: a review. J Med Entomol 5(3):339–352. https://doi.org/10.1093/jmedent/5.3.339
- Geerts T, Heyden Y (2011) In silico predictions of ADME-Tox properties: drug absorption. Comb Chem High Throughput Screen 14(5):339–361. https://doi.org/10.2174/138620711795767764
- Genheden S, Ryde U (2015) The MM/PBSA and MM/GBSA methods to estimate ligand-binding affinities. Expert Opin Drug Discov 10:449–461. https://doi.org/10.1517/17460441.2015.1032936
- Ghayempour S, Montazer M (2016) Micro/nanoencapsulation of essential oils and fragrances: focus on perfumed, antimicrobial, mosquito-repellent and medical textiles. J Microencapsul 33(6):497–510
- Ghini R, Bettiol W (2000) Proteção de plantas na agricultura sustentável. Geography. Retrieved from https://api.semanticscholar.org/ CorpusID:128380862
- Gimero A, Ojeda-Montes M, Tomás-Hernández S, Cereto-Massagué et al (2019) The light and dark sides of virtual screening: what is there to know? Int J Mol Sci 20(6):1375
- Gohlke H, Klebe G (2002) Approaches to the description and prediction of the binding affinity of small-molecule ligands to macromolecular receptors. Angew Chem 41(15):76–2644. https://doi.org/10.1002/1521-3773(20020802)41:15%3c2644::AID-ANIE2 644%3e3.0.CO:2-O
- Gomes J, Rocha L, Leal C, Filho E (2022) Virtual screening of molecular databases for potential inhibitors of the NSP16/NSP10 methyltransferase from SARS-CoV-2. J Mol Struct. https://doi.org/10.1016/j.molstruc.2022.131108
- Gomez-Diaz C, Reina JH, Cambillau C, Benton R (2013) Pheromone ligands for sensory neurons are not conformationally activated odor-binding proteins. PLoS Biol 11:e1001546
- Gonçalves G, Santos J, Silva C, Santos E et al (2006) Occurrence of fruit flies (Diptera: Tephritidae) in the state of Alagoas, Brazil. Fla Entomol 89(1):93–94. https://doi.org/10.1653/0015-4040(2006)89[93:OOFFDT]2.0.CO;2
- Gonçalves G, Silva C, Mendonça A, Vaníčková L et al (2013) Pheromone communication in *Anastrepha obliqua* (Diptera:



- Tephritidae): a comparison of the volatiles and salivary gland extracts of two wild populations. Fla Entomol 96(4):1365–1374. https://doi.org/10.1653/024.096.0416
- González-González A, Rubio-Melendez M, Ballesteros G, Ramirez C et al (2019) Sex- and tissue-specific expression of odorant-binding proteins and chemosensory proteins in adults of the scarab beetle *Hylamorpha elegans* (Burmeister) (Coleoptera: Scarabaeidae). Zoolog Sci. https://doi.org/10.7717/peerj.7054
- Goodsell DS, Olson AJ (2000) Structural symmetry and protein function. Annu Rev Biophys Biomol Struct 29:105–136. https://doi.org/10.1146/annurev.biophys.29.1.105
- Gouchll HK, Schmidt CH, Gilbert LJ (1957) Evaporation of repellents from skin and cloth. Entomol Soc America Bull 3:36–37
- Goulart H, Lima F, Morais R, Bernardo V (2015) Feromônios: Uma Alternativa Verde para o Manejo Integrado de Pragas. Rev Virtual de Quím 1(20). Retrieved from https://docs.ufpr.br/~rbg/assets/files/Feromonios%20NO%20MIP.pdf
- Grosdidier A, Zoete V, Michielin O (2011) SwissDock, a proteinsmall molecule docking web service based on EADock DSS. Nucleic Acids Res 39:270–277. https://doi.org/10.1093/nar/ gkr366
- Gschwend DA, Kuntz ID (1996) Orientational sampling and rigidbody minimization in molecular docking, revisited: on-the-fly optimization and degeneracy removal. J Comput-Aided Mol des 10(2):123–132
- Gu SH, Zhou JJ, Gao S et al (2015) Identification and comparative expression analysis of odorant binding protein genes in the tobacco cutworm *Spodoptera litura*. Sci Rep 5:13800. https://doi.org/10.1038/srep13800
- Guedes IA, Pereira da Silva MM, Galheigo M, Krempser E, de Magalhães CS, Correa Barbosa HJ, Dardenne LE (2024) DockThor-VS: a free platform for receptor-ligand virtual screening. J Mol Biol. https://doi.org/10.1016/j.jmb.2024.168548
- Guerra A, Denis S, Goff O, Sicardi V et al (2016) Development and validation of a new dynamic computer-controlled model of the human stomach and small intestine. Biotechnol Bioeng 113(6):1325–1335. https://doi.org/10.1002/bit.25897
- Guido R, Oliva G, Andricopulo A (2008) Virtual screening and its integration with modern drug design technologies. Curr Med Chem 15:37–46. https://doi.org/10.2174/092986708783330790
- Gunsteren W, Bakowis D, Baron R, Chandrasekhar I et al (2006) Biomolecular modeling: goals, problems, perspectives. Angew Chem Int Ed 45(25):4064–4092. https://doi.org/10.1002/anie. 200502655
- Guo Z, Yamaguchi R (2022) Machine learning methods for proteinprotein binding affinity prediction in protein design. Front Bioinform 2:1065703. https://doi.org/10.3389/fbinf.2022.1065703
- Hameduh T, Haddad Y, Adam V, Heger Z (2020) Homology modeling in the time of collective and artificial intelligence. Comput Struct Biotechnol J 18:3494–3506. https://doi.org/10.1016/j.csbj.2020.
- Hassanali A, Herren H, Khan Z, Pickett J et al (2008) Integrated pest management: the push–pull approach for controlling insect pests and weeds of cereals, and its potential for other agricultural systems including animal husbandry. Philos Trans R Soc B 363:611–621. https://doi.org/10.1098/rstb.2007.2173
- He XY, Zhang Q (2012) Roles of alpha-farnesene and its derivatives in chemical communication of the apple maggot fly, *Rhagoletis pomonella*. J Chem Ecol 38(4):319–329
- Heath RR et al (1993) Sex pheromone components of male Mediterranean fruit fly: the role of the major components in female attraction. J Chem Ecol 19(4):661–671
- Hernandez M, Ghersi D, Sanchez R (2009) SITEHOUND-web: a server for ligand binding site identification in protein structures. Nucleic Acids Res 37:W413–W416. https://doi.org/10.1093/nar/gkp281

- Hess B (2007) P-LINCS: a parallel linear constraint solver for molecular simulation. J Chem Theory Comput 4(1):116–122. https://doi.org/10.1021/ct700200b
- Hess B, Kutzner C, Spoel D, Lindahl E (2008) GROMACS 4: algorithms for highly efficient, load-balanced, and scalable molecular simulation. J Chem Theory Comput 4(3):435–447. https://doi.org/10.1021/ct700301q
- Holt R, Subramanian G, Halpern A, Sutton G et al (2002) The genome sequence of the malaria mosquito *Anopheles gambiae*. Science 298(5591):49–129. https://doi.org/10.1126/science.1076181
- Huang W, Lin Z, Gunsteren W (2011) Validation of the GROMOS 54A7 force field with respect to β-peptide folding. J Chem Theory Comput 7(5):1237–1243. https://doi.org/10.1021/ct100747y
- Ioannou C, Papadopoulos N, Kouloussis N, Tananaki C et al (2012) Essential oils of citrus fruit stimulate oviposition in the Mediterranean fruit fly *Ceratitis capitata* (Diptera: Tephritidae). Physiol Entomol 37(4):330–339. https://doi.org/10.1111/j.1365-3032. 2012.00847.x
- Irwin J, Shoichet B (2005) ZINC-a free database of commercially available compounds for virtual screening. J Chem Inf Model 45(1):177-182
- Jacobson M, Ohinata K, Chambers DL, Jones WA, Fujimoto MS (1973) Insect sex attractants: 13—Isolation, identification and synthesis of sex pheromones of the Mediterranean fruit fly. J Med Chem 16:248–251. https://doi.org/10.1021/jm00262a023
- Jaffar S, Lu Y (2022) Toxicity of Some essential oils constituents against oriental fruit fly, *Bactrocera dorsalis* (Hendel) (Diptera: Tephritidae). InSects 13(10):954
- Jaleel W, He Y, Lü L (2019) The response of two Bactrocera species (Diptera: Tephritidae) to fruit volatiles. J Asia-Pac Entomol 22:758–765
- Jaleel W, Saeed R, Shabbir M, Azad R et al (2021) Olfactory response of two different Bactrocera fruit flies (Diptera: Tephritidae) on banana, guava, and mango fruits. J King Saud Univ Sci 33(5):101455
- Jang EB, Light DM (1991) Behavioral responses of female Mediterranean fruit flies to the sex pheromone and other attractants in a laboratory flight tunnel. J Insect Behav 4(6):751–762
- Jang E, Light D (1996) Olfactory semiochemicals of tephritids. In: McPheron BA, Steck GJ (eds) Fruit fly pests: a world assessment of their biology and management. St. Lucie Press, Delray Beach, pp 73–90
- Jang E, Light D, Flath R, Nagata J et al (1989a) Electroantennogram responses of the Mediterranean fruit-fly, *Ceratitis capitata* to identified volatile constituents from calling males. Entomol Exp Appl 50:7–19. https://doi.org/10.1111/j.1570-7458.1989.tb023 07.x
- Jayanthi KP, Kempraj V, Aurade RM, Roy TK, Shivashankara KS, Verghese A (2014) Computational reverse chemical ecology: virtual screening and predicting behaviorally active semiochemicals for *Bactrocera dorsalis*. BMC Genom 19(15):209. https:// doi.org/10.1186/1471-2164-15-209.PMID:24640964;PMCID: PMC4003815
- Jones G et al (1997) Development and validation of a genetic algorithm for flexible docking. J Mol Biol 267(3):727–748. https://doi.org/10.1006/jmbi.1996.0897
- Jorgensen W, Chandrasekhar J, Madura J, Impey R et al (1983) Comparison of simple potential functions for simulating liquid water. J Chem Phys. Retrieved from https://www.semanticscholar.org/paper/Comparison-of-simple-potential-functions-for-liquid-Jorgensen-Chandrasekhar/3e91f8caa53385dc4256634638431d10a8c9173a
- Jumper J, Evans R, Pritzel A, Green T, Figurnov M, Ronneberger O, Hassabis D (2021) Highly accurate protein structure prediction with AlphaFold. Nature 596(7873):583–589. https://doi.org/10. 1038/s41586-021-03819-2



- Jurrus E, Engel D, Star K, Monson K et al (2017) Improvements to the APBS biomolecular solvation software suite. Protein Sci 27(1):112–128. https://doi.org/10.1002/pro.3280
- Kavlock R, Ankley G, Blancato J, Breen M et al (2008) Computational toxicology: a state of the science mini review. Toxicol Sci 103(1):14–27. https://doi.org/10.1093/toxsci/kfn287
- Keil T (1999) Morphology and development of the peripheral olfactory organs. Insect Olfaction. https://doi.org/10.1007/978-3-662-07911-9 2
- Khurana S, Siddiqi O (2013) Respostas olfativas de larvas de Drosophila. Chem Senses 38(4):315–323. https://doi.org/10.1093/chemse/bjs144
- Kim S, Chen J, Cheng T, Gindulyte A et al (2023a) PubChem 2023 update. Nucleic Acids Res 51:1373–1380. https://doi.org/10. 1093/nar/gkac205
- Kitchen D, Decornez H, Furr J, Bajorath J (2004a) Docking and scoring in virtual screening for drug discovery: methods and applications. Nat Rev Drug Discov 3(11):935–949. https://doi.org/10.1038/nrd1549
- Klein U (1987) Sensillum-lymph proteins from antennal olfactory hairs of the moth Antheraea polyphemus (Saturniidae). Insect Biochem 17:1193–1204. https://doi.org/10.1016/0020-1790(87)90093-X
- Knudsen J, Tollsten L, Bergström L (1993) Floral scents: a checklist of volatile compounds isolated by head-space techniques. Phytochemistry 33:253–280. https://doi.org/10.1016/0031-9422(93)85502-I
- Kolafa J, Perram J (1992) Cutoff Errors in the Ewald summation formulae for point charge systems. Mol Simul. https://doi.org/ 10.1080/08927029208049126
- Kollman PA, Massova I, Reyes C, Kuhn B, Cheatham TE (2000) Calculating structures and free energies of complex molecules: combining molecular mechanics and continuum models. Acc Chem Res 33:889–897. https://doi.org/10.1021/ar000033j
- Krumpe LR, Mori T (2007) Potential of phage-displayed peptide library technology to identify functional targeting peptides. Expert Opin Drug Discov 2(4):525–537. https://doi.org/10.1517/17460441.2.4.525
- Kumari R, Kumar R, Lynn A (2014a) G-mmpbsa-a GROMACS tool for high-throughput MM-PBSA calculations. J Chem Inf Model 54:1951–1962. https://doi.org/10.1021/ci500020m
- Kumari S, Pundhir S, Priya P, Jeena G et al (2014b) EssOilDB: a database of essential oils reflecting terpene composition and variability in the plant Kingdom. Database (Oxford). https://doi.org/10.1093/database/bau120
- Kurcinski M, Badaczewska-Dawid A, Kolinski M, Kolinski A et al (2019) Flexible docking of peptides to proteins using CABSdock. Protein Sci 29:211–222. https://doi.org/10.1002/pro. 3827
- Kuzmanic A, Zagrovic B (2010) Determination of ensemble-average pairwise root mean-square deviation from experimental B-factors. Biophys J 98:861–871. https://doi.org/10.1016/j.bpj.2009. 11.002
- Landolt PJ, Sivinski JM (1992) Effect of time of day, adult age, and host availability on sexual signaling and sexual selection in the Caribbean fruit fly, *Anastrepha suspensa*. Environ Entomol 21(4):687–693
- Laughlin JD, Ha TS, Jones DNM, Smith DP (2008) Activation of pheromone-sensitive neurons is mediated by conformational activation of pheromone-binding protein. Cell 133(7):1255–1265. https://doi.org/10.1016/j.cell.2008.04.046
- Le Guilloux V, Schmidtke P, Tuffery P (2009) Fpocket: an open source platform for ligand pocket detection. BMC Bioinform 10:168. https://doi.org/10.1186/1471-2105-10-168
- Leal W (2005) Pheromone reception. Top Curr Chem 240:1–36. https://doi.org/10.1007/b98314

- Leal WS (2013) Odorant reception in insects: roles of receptors, binding proteins, and degrading enzymes. Annu Rev Entomol 58:373–391
- Leal T, Zucoloto F (2008) Selection of artificial hosts for oviposition by wild Anastrepha obliqua (Macquart) (Diptera, Tephritidae): Influence of adult food and effect of experience. Rev Bras Entomol 52. Retrieved from https://www.researchgate.net/publication/250034295_Selection_of_artificial_hosts_for_oviposition_by_wild_Anastrepha_obliqua_Macquart_Diptera_Tephritidae_Influence of adult food and effect of experience
- Leal W, Nikonova L, Peng G (1999) Disulfide structure of the pheromone binding protein from the silkworm moth *Bombyx mori*. FEBS Lett 464:85–90. https://doi.org/10.1016/S0014-5793(99) 01683-X
- Leal W, Choo Y, Xu P, Da Silva C et al (2013) Differential expression of olfactory genes in the southern house mosquito and insights into unique odorant receptor gene isoforms. Proc Natl Acad Sci USA 110:18704–18709. https://doi.org/10.1073/pnas.1316059110
- Lee D, Damberger F, Peng G, Horst R et al (2002) NMR structure of the unliganded *Bombyx mori* pheromone-binding protein at physiological pH. FEBS Lett 531(2):314–318. https://doi.org/10.1016/S0014-5793(02)03548-2
- Lee H, Heo L, Lee M, Seok C (2015) GalaxyPepDock: a proteinpeptide docking tool based on interaction similarity and energy optimization. Nucleic Acids Res 42:5–431. https://doi.org/10. 1093/nar/gkv438
- Legeay S, Clere N, Hilairet G et al (2016) The insect repellent N, N-diethyl-m-toluamide (DEET) induces angiogenesis via allosteric modulation of the M3 muscarinic receptor in endothelial cells. Sci Rep. https://doi.org/10.1038/srep28546
- Leite S, Costa D, Ribeiro A, Moreira A et al (2019) Oviposition preference and biological performance of *Ceratitis capitata* in Anacardiaceae, Cactaceae and Vitaceae fruit. Agric Entomol 86:1–8
- Liggri P, Pérez-Garrido A, Tsitsanou K, Dileep K et al (2023) 2D finger-printing and molecular docking studies identified potent mosquito repellents targeting odorant binding protein 1. Insect Biochem Mol Biol. https://doi.org/10.1016/j.ibmb.2023.103961
- Light DM, Jang EB, Dickens JC (1988a) Electroantennogram response of the Mediterranean fruit fly, *Ceratitis capitata*, to a spectrum of plant volatiles. J Chem Ecol 14:159–180. https://doi.org/10.1007/BF01022539
- Lima-Medonça A, Medonça A, Sant'Ana A, Nascimento R (2014) Semioquímicos de moscas das frutas do gênero Anastrepha. Quím Nova 37(2):293–301
- Liquido N, Shinoda L, Cunningham T (1991) Host plants of the Mediterranean fruit fly (Diptera: Tephritidae): an annotated world review. Entomological Society of America Miscellaneous Publications, Lanham
- Liu J, Gui P, Wang X, Chen H et al (2022) Study on phytochemical and pharmacological activities of four Rhododendron plants endemic to Northeast China. J Agric Food Res 7:100255
- López-Guillén G, López LC, Malo EA, Rojas JC (2011) Olfactory responses of *Anastrepha obliqua* (Diptera: Tephritidae) to volatiles emitted by calling males. Florida Entomologist 94(4):874–881
- Lull C, Gil-Ortiz R, Cantín A (2023) A chemical approach to obtaining α-copaene from clove oil and its application in the control of the medfly. Appl Sci 13(9):5622
- Luu-Dam N, Tabanca N, Estep A, Nguyen D (2021) Insecticidal and attractant activities of *Magnolia citrata* leaf essential oil against two major pests from Diptera: *Aedes aegypti* (Culicidae) and *Ceratitis capitata* (Tephritidae). Molecules 26(8):2311
- Maiorov V, Crippen G (1994) Significance of root-mean-square deviation in comparing three-dimensional structures of globular



- proteins. J Mol Biol 235(2):625–634. https://doi.org/10.1006/jmbi.1994.1017
- Malavasi A (2009) Biologia, ciclo de vida, relação com o hospedeiro, espécies importantes e biogeografia de tefritideos. In: Malavasi A, Virginio JF (eds) Biologia, monitoramento e controle de moscas-das-frutas. Biofábrica Moscamed Brasil, Juazeiro, pp 1–5
- Malde A, Zue L, Brisa M, Stroet M et al (2011) An automated force field topology builder (ATB) and repository: version 1.0. J Chem Theory Comput 7(12):4026–4037. https://doi.org/10.1021/ct200 196m
- Manoukis NC, Siderhurst MS, Jang E (2015) Field estimates of attraction of Ceratitis capitata to trimedlure and Bactrocera dorsalis (Diptera: Tephritidae) to methyl eugenol in varying environments. Environ Entomol 44(3):695–703. https://doi.org/10.1093/ee/nvv020
- Manrakhan A, Daneel J, Stephen P, Hattingh V (2022) Cold tolerance of immature stages of *Ceratitis capitata* and *Bactrocera dorsalis* (Diptera: Tephritidae). J Econ Entomol 115(2):482–492
- Maruyama Y, Igarashi R, Ushiku Y, Mitsutake A (2023) Analysis of protein folding simulation with moving root mean square deviation. J Chem Inf Model 63(5):1529–1541. https://doi.org/10.1021/acs.jcim.2c01444
- Mazola Y, Guirola O, Palomares S, Chinea G et al (2015) A comparative molecular dynamics study of thermophilic and mesophilic β-fructosidase enzymes. J Mol Model 21(8):228. https://doi.org/10.1007/s00894-015-2711-7
- McPheron BA, Steck GJ (eds) (1996) Fruit Fly Pests: a world assessment of their biology and management. St. Lucie Press, Delray Beach
- Megwer M, Schornow HG (2001) Organic-chemical drugs and their synonyms, vol 2. Wiley, Berlim, p 1178
- Melo R, Brito L, Petrere V, Angelotti F et al (2010) Pesticidas e seus impactos no ambiente. EMBRAPA. https://www.alice.cnptia.embrapa.br/alice/bitstream/doc/875500/1/Roseli.pdf
- Menenses A, Balen M, Jasper E, Korte I et al (2020) Enzymatic synthesis of benzyl benzoate using different acyl donors: Comparison of solvent-free reaction techniques. Process Biochem 92:261–268. https://doi.org/10.1016/j.procbio.2020.01.018
- Merli D, Mannucci B, Bassetti F, Corana F et al (2018) Larval diet affects male pheromone blend in a laboratory strain of the Medfly, *Ceratitis capitata* (Diptera: Tephritidae). J Chem Ecol 44:339–353. https://doi.org/10.1007/s10886-018-0939-z
- Metcalf E (1995) Biography of the medfly. In: Morse JG, Metcalf RL, Carey JR, Dowell RV (eds) The Mediterranean fruit fly in California: defining critical research. University of California-Riverside, pp 43–48
- Mikovski D, Basso J, Silva P, Ribas J (2018) Química medicinal E A Sua Importância No Desenvolvimento De Novos Fármacos. Rev Saúde Desenvolv 12(13):29–43
- Milet-Pinheiro P, Navarro D, De Aquino N, Ferreira L et al (2015a) Identification of male-borne attractants in *Anastrepha fraterculus* (Diptera: Tephritidae). Chemoecology 25(3):115–122. https://doi.org/10.1007/s00049-014-0180-3
- Montes S, Raga A, Boliani A, Santos P (2011) Dinâmica populacional e incidência de moscas-das-frutas e parasitoides em cultivares de pessegueiros (*Prunus persica* L. Batsch) no município de Presidente Prudente – SP. Rev Bras Frutic 33(2):402–411. https://doi. org/10.1590/S0100-29452011005000052
- Morris G, Lim-Wilby M (2008) Molecular docking. Methods Mol Biol 443:82–365. https://doi.org/10.1007/978-1-59745-177-2_3
- Morris G, Huey R, Lidstrom W, Sanner M et al (2009) AutoDock4 and AutoDockTools4: automated docking with selective receptor flexibility. J Chem Inf Model 16(30):2785–2791. https://doi.org/10.1021/ci1001272

- Mourão E (2021) Triagem virtual de metabólitos secundários de plantas da caatinga na busca por potenciais agentes para o controle de Aedes aegypti. Dissertação Universidade Federal do Vale do São Francisco. Petrolina PE, pp 1–118. Retrieved from http://www.univasf.edu.br/~tcc/00001c/00001c82.pdf
- Mucha M, Szyk B, Pamula H, Mucha K et al (2023) About us: Omni Calculator. Retrieved from https://www.omnicalculator.com/ about-us
- Mysinger M, Carchia M, Irwin J, Schoichet B (2012) Directory of useful decoys, enhanced (DUD-E): better ligands and decoys for better benchmarking. J Med Chem 55(14):6582–6594. https://doi.org/10.1021/jm300687e
- Namba Ā, Silva V, Šilva C (2008) Dinâmica molecular: teoria e aplicações em planejamento de fármacos. Eclét Quím 33(4):13–24
- Nava DE, Botton M (2010) Bioecologia e controle de *Anastrepha* fraterculus e Ceratitis capitata em pessegueiro. Embrapa, Pelotas
- Nemoto H, Shiraki M, Nagamochi M, Fukumoto K (1993) A concise enantiocontrolled total synthesis of (-)-a-bisabolol and (+)-4-epia-bisabolol. Tetrahedron 34(31):4939–4942
- Nesterkina M, Bernier U, Tabanca N, Kravchenko I (2018) Repellent activity of monoterpenoid esters with neurotransmitter amino acids against yellow fever mosquito, *Aedes aegypti*. Open Chem. https://doi.org/10.1515/chem-2018-0015/html
- Newman DJ, Cragg GM (2007) Natural products as sources of new drugs over the last 25 years. J Nat Prod 70(3):461–477. https://doi.org/10.1021/np068054v
- Ohinata K, Jacobson M, Nakagawa S, Fujimoto M et al (1977) Mediterranean fruit fly: laboratory and field evaluations of synthetic sex pheromones. J Environ Sci Health, Part A 12(3):67–68
- Oschkinat H, Griesinger C, Kraulis PJ, Sørensen OW, Ernst RR, Gronenborn AM, Clore GM (1988) Three-dimensional NMR spectroscopy of a protein in solution. Nature 332(6162):374–376. https://doi.org/10.1038/332374a0
- Ouarhach A et al (2022) Evaluation of insecticidal activity of *Lavan-dula coronopifolia* essential oil against the Mediterranean fruit fly *Ceratitis capitata* Wiedemann (Diptera: Tephritidae). S Afr J Bot 149:748–753
- Paluch G, Bartholomay L, Coats J (2010a) Mosquito repellents: a review of chemical structure diversity and olfaction. Pest Manag Sci 66(9):925–935. https://doi.org/10.1002/ps.1974
- Papadopoulos N, Shelly T, Niyazi N et al (2006) Olfactory and behavioral mechanism underlying enhanced mating competitiveness following exposures to ginger root oil and orange oil in males of the Mediterranean fruit fly, *Ceratitis capitata* (Diptera: Tephritidae). J Insect Behav 19(3):403–418. https://doi.org/10.1007/s10905-006-9031-6
- Papanicolaou A, Schetelig M, Arensburger P, Atkinson P et al (2016)
 The whole genome sequence of the Mediterranean fruit fly, Ceratitis capitata (Wiedemann), reveals insights into the biology and adaptive evolution of a highly invasive pest species. Genome Biol 17:1–31
- Pappas R (2024) About EOU. Essential oils. https://essentialoils.org/
- Paranhos BAJ, Nascimento AS, Barbosa FR, Viana R, Sampaio R, Malavasi A, Walder JMM (2008) Técnica do inseto estéril: nova tecnologia para combater a mosca-das-frutas, Ceratitis capitata, no Submédio do Vale do São Francisco. Comun Téc Embrapa Semiárido 137:1–6
- Parrinello M, Rahman A (1981) Polymorphic transitions in single crystals: a new molecular dynamics method. J Chem Phys 52:7182–7190. https://doi.org/10.1063/1.328693
- Pearlman D, Case D, Caldwell J, Ross W et al (1995) AMBER, a package of computer programs for applying molecular mechanics, normal mode analysis, molecular dynamics and free energy



- calculations to simulate the structural and energetic properties of molecules. Comput Phys Commun 91:1–41
- Pelletier J, Guidolin A, Syed Z, Cornel A et al (2010) Knockdown of a mosquito odorant-binding protein involved in the sensitive detection of oviposition attractants. J Chem Ecol 36:245–248. https://doi.org/10.1007/s10886-010-9762-x
- Pelosi P (1994) Odorant-binding proteins. Crit Rev Biochem Mol Biol 29:199–228. https://doi.org/10.3109/10409239409086801
- Pelosi P, Maida R (1995) Odorant-binding proteins in insects. Comp Biochem Physiol b: Biochem Mol Biol 111(3):503–514. https:// doi.org/10.1016/0305-0491(95)00019-5
- Pelosi P, Mastrogiacomo R, Iovinella I, Tuccori E et al (2014) Structure and biotechnological applications of odorant-binding proteins. Appl Microbiol Biotechnol 98:61–70. https://doi.org/10.1007/s00253-013-5383-y
- Pence H, Williams A (2010) ChemSpider: an online chemical information resource. J Chem Educ 87(11):1123–1124. https://doi.org/10.1021/ed100697w
- Pereira J (2003) O comportamento da Mosca-do-Mediterrâneo, *Ceratitis capitata* (Wied.) (Díptera: Tephritidae) em Pegões (Dissertação de mestrado, Instituto Superior de Agronomia, Universidade de Lisboa)
- Petersen E, Goddard T, Huang C, Couch G et al (2004) UCSF Chimera–a visualization system for exploratory research and analysis. J Chem Inf Model 25(13):1605–1612. https://doi.org/10.1021/ci030064+
- Pilon A, Valli M, Dametto A, Pinto M et al (2017) NuBBEDB: an updated database to uncover chemical and biological information from Brazilian biodiversity. Sci Rep. https://doi.org/10.1038/s41598-017-07451-x
- Pinto G, Hendrikse N, Stourac J, Damborsky J et al (2022) Virtual screening of potential anticancer drugs based on microbial products. Semin Cancer Biol 86:1207–1217. https://doi.org/10.1016/j.semcancer.2022.03.013
- Pires D, Blundell T, Ascher D (2015) pkCSM: predicting small-molecule pharmacokinetic and toxicity properties using graph-based signatures. J Med Chem 58(9):4066–4072. https://doi.org/10. 1021/acs.jmedchem.5b00104
- Podio-Guidugli P (2010) On (Andersen-)Parrinello-Rahman molecular dynamics, the related metadynamics, and the use of the Cauchy-Born rule. J Elast 100:145–153. https://doi.org/10.1007/s10659-010-9250-0
- Ponce WP, Nation JL, Emmel TC, Smittle BJ, Teal PEA (1993) Quantitative analysis of pheromone production in irradiated Caribbean fruit fly males, *Anastrepha suspensa* (Loew). J Chem Ecol 19:3045
- Portilla-Pulido J, Castillo-Morales R, Rodríguez M, Duque J et al (2020) Design of a repellent against *Aedes aegypti* (Diptera: Culicidae) using in silico simulations with AaegOBP1 protein. J Med Entomol 57(2):463–476
- Prokopy R, Hendrichs J (1979) Mating-behavior of *Ceratitis capitata* (Diptera, Tephritidae) on a field-caged host tree. Ann Entomol Soc Am 72:642–648. https://doi.org/10.1093/aesa/72.5.642
- Raga A, Sato M (2016) Controle Químico de Moscas-das-Frutas. Instituto Biológico: Documento técnico, pp 1–14. Retrieved from http://www.biologico.sp.gov.br/uploads/docs/dt/moscas_das_frutas.pdf
- Rana A, Sharma D, Choudhary K, Kumari P, Ruchika K, Yangchan J, Kumar S (2024) Insight into insect odorant binding proteins: an alternative approach for pest management. J Nat Pestic Res 8:100069. https://doi.org/10.1016/j.napere.2024.100069
- Reddy GVP, Guerrero A (2004a) Interactions of insect pheromones and plant semiochemicals. Trends Plant Sci 9(5):253–261. https://doi.org/10.1016/j.tplants.2004.03.009

- Renthal R (2024) Arthropod repellent interactions with olfactory receptors and ionotropic receptors analyzed by molecular modeling. Curr Res Insect Sci 5:100082
- Rihani K, Ferveur JF, Briand L (2021) The 40-year mystery of insect odorant-binding proteins. Biomolecules 11(4):509. https://doi.org/10.3390/biom11040509
- Roadhouse L (1953a) Laboratory studies on insect repellency. Can J Zool 31(5):535–546. https://doi.org/10.1139/z53-038
- Robacker DC, Hart WG (1987) Electroantennograms of male and female Caribbean fruit flies (Diptera: Tephritidae) elicited by chemicals produced by males. Ann Entomol Soc Am 80:508–512. https://doi.org/10.1093/AESA/80.4.508/title/electroant ennograms_of_male_and_female_caribbean_fruit_flies_dipte ra_tephritidae_elicited_by_chemicals_produced_by_males1
- Robacker DC, Chapa BE, Hart WG (1986) Electroantennograms of Mexican fruit flies to chemicals produced by males. Entomol Exp Appl 40:123–127. https://doi.org/10.1111/j.1570-7458. 1986.tb00492.x
- Rocca J, Nation J, Strekowski L, Battiste M (1992) Comparison of volatiles emitted by male Caribbean and Mexican fruit flies. J Chem Ecol 18:223–244. https://doi.org/10.1007/BF00993755
- Rodrigues R, Mantoani S, Almeida J, Pinsetta F, Semighini E, Barreto da Silva V, Silva C (2012) Virtual screening strategies in drug design. Rev Virtual Quím 4(6):739–762. https://doi.org/10.5935/1984-6835.20120055
- Rodríguez-Espigares I, Torrens-Fontanals M, Tiemann J, Aranda-García D et al (2020) GPCRmd uncovers the dynamics of the 3D-GPCRome. Nat Methods 17:777–787
- Ruiz M, Juárez M, Barud F, Goane L et al (2021) Lemon and Schinus polygama essential oils enhance male mating success of Anastrepha fraterculus. Entomol Exp Appl 169(2):172–182. https:// doi.org/10.1111/eea.13005
- Rutz A, Sorokina M, Galgonek J, Mietchen D et al (2022) The LOTUS initiative for open knowledge management in natural products research. Elife. https://doi.org/10.7554/elife.70780
- Šali A, Blundell TL (1993) Comparative protein modelling by satisfaction of spatial restraints. J Mol Biol 234:779–815
- Sánchez-Gracia A, Vieira FG, Rozas J (2009) Molecular evolution of the major chemosensory gene families in insects. Heredity 103:208–216. https://doi.org/10.1038/hdy.2009.55
- Sawh-Gopal A, Ishemgulova A, Chua EYD, Aragon MF, Mendez JH, Eng ET, Noble AJ (2023) Cryo-electron microscopy screening automation across multiple grids using Smart Leginon. Biochemistry. https://doi.org/10.3791/66007
- Schmid N, Eichenberger A, Choutko A, Riniker S et al (2011) Definition and testing of the GROMOS force-field versions 54A7 and 54B7. Eur Biophys J 40:843–856. https://doi.org/10.1007/s00249-011-0700-9
- Schmidt-Busser D, Arx M, Guerin P (2009) Host plant volatiles serve to increase the response of male European grape berry moths, *Eupoecilia ambiguella*, to their sex pheromone. J Comp Physiol 195(12):1432–1351. https://doi.org/10.1007/s00359-009-0464-1
- Schneider G (2010) Virtual screening: an endless staircase? Nat Rev Drug Discov 9:6–273. https://doi.org/10.1038/nrd3187
- Schrödinger L, DeLano W (2020) PyMOL. http://www.pymol.org/ pymolScolari
- Schwab W, Davidovich-Rikanati R, Lewinsohn E (2008) Biosynthesis of plant-derived flavor compounds. Plant J 54(4):712–732
- Scolari F, Valerio F, Benelli G, Papadopoulos NT, Vaníčková L (2021) Tephritid fruit fly semiochemicals: current knowledge and future perspectives. InSects 12(5):408. https://doi.org/10.3390/insects12050408
- Shaw D (2010) Atomic-level characterization of the structural dynamics of proteins. Science 330(6002):341–346. https://doi.org/10.1126/science.1187409



- Shelly T, Nishimoto J (2015) Exposure to the plant compound α-humulene reduces mating success in male Mediterranean fruit flies (Diptera: Tephritidae). Ann Entomol Soc America 108(3):215–221
- Shi Y (2014) A glimpse of structural biology through X-ray crystallography. Cell 159(5):995–1014. https://doi.org/10.1016/j.cell. 2014.10.051
- Siciliano P, He X, Woodcock C, Pickett J et al (2014) Identification of pheromone components and their binding affinity to the odorant binding protein CcapOBP83a-2 of the Mediterranean fruit fly, Ceratitis capitata. Insect Biochem Mol Biol 48:51–52
- Siderhurst M, Jang E (2006) Female-biased attraction of oriental fruit fly, *Bactrocera dorsalis* (Hendel), to a blend of host fruit volatiles from *Terminalia catappa* L. J Chem Ecol 32:34–2513. https://doi.org/10.1007/s10886-006-9160-6
- Sievers F, Wilm A, Dineen D, Gibson TJ, Karplus K, Li W, Lopez R, McWilliam H, Remmert M, Söding J, Thompson JD, Higgins DG (2011) Fast, scalable generation of high-quality protein multiple sequence alignments using clustal omega. Mol Syst Biol 7:539. https://doi.org/10.1038/msb.2011.75
- Silva R, Milet-Pinheiro P, Silva P, Silva A et al (2015) (E)-caryophyllene and α-humulene: *Aedes aegypti* oviposition deterrents elucidated by gas chromatography-electrophysiological assay of *Commiphora leptophloeos* leaf oil. PLoS ONE. https://doi.org/10.1371/journal.pone.0144586
- Solari P, Maccioni R, Marotta R, Catelani T et al (2018) The imbalance of serotonergic circuitry impairing the crop supercontractile muscle activity and the mitochondrial morphology of PD PINK1B9 Drosophila melanogaster are rescued by *Mucuna pruriens*. J Insect Physiol 111:32–40
- Sollai G, Solari P, Crnjar R (2018) Olfactory sensitivity to major, intermediate and trace components of sex pheromone in *Ceratitis capitata* is related to mating and circadian rhythm. J Insect Physiol 110:23–33. https://doi.org/10.1016/j.jinsphys.2018.08.002
- Stroet M, Caron B, Visscher K, Geerke D et al (2018) Automated topology builder Versão 3.0: Predição de entalpias livres de solvatação em água e hexano. J Chem Theory Comput 14(11):5834–5845. https://doi.org/10.1021/acs.jctc.8b00768
- Suckling D, Dymock J, Park K, Wakelin R et al (2013) Communication disruption of guava moth (*Coscinoptycha improbana*) using a pheromone analog based on chain length. J Chem Ecol 39:1161–1168. https://doi.org/10.1007/s10886-013-0353-5
- Sun L, Xu W, Li Q, Gao Y, Sun Y, Zhang Y (2018a) Identification of an odorant-binding protein (OBP) gene closely associated with olfactory function in the blowfly, *Lucilia Cuprina*. Insect Sci 25(2):205–216
- Sun JS, Larter NK, Chahda JS, Rioux D, Gumaste A, Carlson JR (2018b) Humidity response depends on the small soluble protein Obp59a in Drosophila. Elife 7:e39249
- Tabanca N, Masi M, Epsky ND, Nocera P, Cimmino A, Kendra PE, Niogret J, Evidente A (2019) Laboratory evaluation of natural and synthetic aromatic compounds as potential attractants for male Mediterranean fruit fly, *Ceratitis capitata*. Molecules 24(13):2409. https://doi.org/10.3390/molecules24132409
- Tabanca N, Niogret J, Kendra P, Epsky N (2020) TLC-based bioassay to isolate Kairomones from tea tree essential oil that attract male Mediterranean fruit flies, *Ceratitis capitata* (Wiedemann). Biomolecules 10(5):683
- Tamura K, Stecher G, Kumar S (2021) MEGA11: molecular evolutionary genetics analysis version 11. Mol Biol Evol 38(7):3022–3027. https://doi.org/10.1093/molbev/msab120
- Tan K, Nishida R, Jang E, Shelly T (2014) Pheromones, male lures, and trapping of tephritid fruit flies. Trapping and the detection, control, and regulation of tephritid fruit flies. Springer, Dordrecht. https://doi.org/10.1007/978-94-017-9193-9_2

- Tawatsin A, Wratten S, Scott R, Thavara U et al (2001) Repellency of volatile oils from plants against three mosquito vectors. J Vector Ecol 26(1):76–82
- Tetko I, Bruneau P, Mewes H, Rohrer D et al (2006) Can we estimate the accuracy of ADME–Tox predictions. Drug Discov Today 11(15–16):700–707. https://doi.org/10.1016/S1359-6446(06) 02230-3
- Thireou T, Kythereoti G, Tsitsanou K, Koussis K et al (2018) Identification of novel bioinspired synthetic mosquito repellents by combined ligand-based screening and OBP-structure-based molecular docking. Insect Biochem Mol Biol 98:48–61. https://doi.org/10.1016/j.jbmb.2018.05.001
- Thomas D, Holler T, Heath R, Salinas E et al (2001) Trap-lure combinations for surveillance of Anastrepha fruit flies (Diptera: Tephritidae). Fla Entomol 84(3). https://journals.flvc.org/flaent/article/view/59657
- Thompson M (2004) Molecular docking using ArgusLab, an efficient shape-based search algorithm and the a score scoring function. ACS Meeting, Philadelphia
- Tiwari V, Sowdhamini R (2023) Structural modelling and dynamics of full-length of TLR10 sheds light on possible modes of dimerization, ligand binding, and mechanism of action. Curr Res Struct Biol 5:100097
- Trott O, Olson A (2009) AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading. J Comput Chem 31(2):455–461. https://doi.org/10.1002/jcc.21334
- Tsitsanou K, Thireou T, Drakou C, Koussis K et al (2012) Anopheles gambiae odorant binding protein crystal complex with the synthetic repellent DEET: implications for structure-based design of novel mosquito repellents. Cell Mol Life Sci 69:283–297. https://doi.org/10.1007/s00018-011-0745-z
- Valerio F, Benelli G, Papadopoulos N et al (2021) Tephritid fruit fly semiochemicals: current knowledge and future perspectives. InSects 12(5):408
- Van der Graaf PH, Benson N (2011) Systems pharmacology: bridging systems biology and pharmacokinetics-pharmacodynamics (PKPD) in drug discovery and development. Pharm Res 28(7):1460–1464. https://doi.org/10.1007/s11095-011-0467-9
- Van Der Pers JN, Haniotakis GE, King BM (1984) Electroantennogram responses from olfactory receptors in Dacus oleae. Entomol Hell 2:47–53
- Vaníčková L, Do Nascimento R, Hoskovec M, Jezkova Z et al (2012a)
 Are the wild and laboratory insect populations different in semiochemical emission? The case of the medfly sex pheromone. J
 Agric Food Chem 60:7168–7176. https://doi.org/10.1021/jf301
 474d
- Vargas R, Ramadan M, Hussain T, Mochizuki N (2001) Comparative demography of six fruit fly (Diptera: Tephritidae) parasitoids (Hymenoptera: Braconidae). Biol Control 25:30–40. https://doi.org/10.1016/S1049-9644(02)00046-4
- Venthur H, Zhou J-J (2018) Odorant receptors and odorant-binding proteins as insect pest control targets: a comparative analysis. Front Physiol 9:1163. https://doi.org/10.3389/fphys.2018.01163
- Verli H (2014) Bioinformática: da biologia à flexibilidade molecular. SBBq, São Paulo
- Vidal M (2023) Fruticultura. Escritório Técnico de Estudos Econômicos do Nordeste ETENE (280). Retrieved from https://www.bnb.gov.br/s482-dspace/bitstream/123456789/1761/1/2023_CDS_280.pdf
- Vieira FG, Sánchez-Gracia A, Rozas J (2007) Comparative genomic analysis of the odorant-binding protein family in 12 Drosophila genomes: purifying selection and birth-and-death evolution. Genome Biol 8(11):R235. https://doi.org/10.1186/ gb-2007-8-11-r235



- Vilela EF, Della Lucia TMC (2001) Feromônios de insetos: biologia, química e emprego no manejo de pragas, 2nd edn. Holos, Ribeirão Preto
- Virgilio M, White I, De Meyer M (2014) A set of multi-entry identification keys to African frugivorous flies (Diptera, Tephritidae). ZooKeys 428:97–108. https://doi.org/10.3897/zookeys.428.7366
- Visser J (1986) Host odour perception in phytophagous insects. Annu Rev Entomol 21:121–144. https://doi.org/10.1146/annurev.en. 31.010186.001005
- Vogt R (2002) Odorant binding protein homologues of the malaria mosquito *Anopheles gambiae*; possible orthologues of the OS-E and OS-F OBPs of *Drosophila melanogaster*. J Chem Ecol 28:2371–2376. https://doi.org/10.1023/A:1021009311977
- Vogt R, Riddiford L (1981) Pheromone binding and inactivation by moth antennae. Nature 293:161–163. https://doi.org/10.1038/ 293161a0
- Vogt R, Köhne A, Dubnau J, Prestwich G (1989) Expression of pheromone binding proteins during antennal development in the gypsy moth *Lymantria dispar*. J Neurosci: off J Soc Neurosci 9(9):3332–3346. https://doi.org/10.1523/jneurosci.09-09-03332.1989
- Vontas J, Hernández-Crespo P, Margaritopoulos J, Ortega F et al (2011) Insecticide resistance in tephritid flies. Pestic Biochem Physiol 100(3):199–205
- Wagman J, Achee N, Grieco J (2015) Insensitivity to the spatial repellent action of transfluthrin in *Aedes aegypti*: a heritable trait associated with decreased insecticide susceptibility. PLoS Negl Trop Dis. https://doi.org/10.1371/journal.pntd.0003726
- Wallace AC, Laskowski RA, Thornton JM (1996) LIGPLOT: a program to generate schematic diagrams of protein-ligand interactions. Protein Eng 8(2):127–134
- Wang Y, Wang Q, Huang H, Huang W, Chen Y, McGarvey PB, Wu CH, Arighi CN, UniProt Consortium (2021) A crowdsourcing open platform for literature curation in UniProt. Plos Biol 19(12):e3001464. https://doi.org/10.1371/journal.pbio.3001464
- Wang L, Hou M, Liang C, Xu Q, Lu Y, Zhao Z (2024) Role of odorant binding protein C12 in the response of *Tribolium castaneum* to chemical agents. Pestic Biochem Physiol 201:105861
- Waterhouse A, Bertoni M, Bienert S, Studer G, Tauriello G, Gumienny R, Heer FT, de Beer TAP, Rempfer C, Bordoli L, Lepore R, Schwede T (2018) SWISS-MODEL: homology modelling of protein structures and complexes. Nucleic Acids Res 46(W1):W296–W303
- White IM, Elson-Harris MM (1992) Fruit flies of economic significance: their identification and bionomics. CAB International, Wallingford
- Witzgall P et al (2008) Sex pheromone of the apple maggot fly: a compound critical for attracting the insect. Chem Senses 33(8):647–655
- Witzgall P, Kirsch P, Cork A (2010) Sex pheromones and their impact on pest management. J Chem Ecol 36(1):80–100
- Wu H, Nie Y, Huse WD, Watkins JD (1999) Humanization of a murine monoclonal antibody by simultaneous optimization of framework and CDR residues. J Mol Biol 294(1):151–162. https://doi.org/ 10.1006/jmbi.1999.3141

- Wüthrich K (2003) NMR studies of structure and function of biological macromolecules (Nobel Lecture). J Biomol NMR 27(1):13–39
- Xu Y, Zhou F, Li Y (2023) Diffusive limit of the Vlasov–Poisson– Boltzmann System without Angular Cutoff. Mathematics Subject Classification. https://arxiv.org/abs/2312.16588
- Yamari I, Abchir O, Siddique F, Zaki H et al (2023) The anticoagulant potential of *Lippia alba* extract in inhibiting SARS-CoV-2 Mpro: density functional calculation, molecular docking analysis, and molecular dynamics simulations. Sci Afr. https://doi.org/10.1016/j.sciaf.2023.103900
- Yin J, Choo Y, Duan H, Leal W (2015) Selectivity of odorant-binding proteins from the southern house mosquito tested against physiologically relevant ligands. Front Physiol. https://doi.org/10.3389/fphys.2015.00056
- Yip KM, Fischer N, Paknia E, Chari A, Stark H (2020) Atomic-resolution protein structure determination by cryo-EM. Nature 587(7832):157–161. https://doi.org/10.1038/s41586-020-2833-4
- Yu H, Adedoyin D (2003) ADME–Tox in drug discovery: integration of experimental and computational technologies. Drug Discov Today 8(18):852–861. https://doi.org/10.1016/S1359-6446(03) 02828-9
- Zanardi O, Nava D, Botton M, Grützmacher A, Junior Machota R et al (2011) Desenvolvimento e reprodução da moscado-mediterrâneo em caquizeiro, macieira, pessegueiro e videira. Pesq Agrop Bras 46(7):682–688. https://doi.org/10.1590/S0100-204X201100
- Zarbin P, Rodrigues M, Lima E (2009) Insect pheromones: technology and challenges for a competitive agriculture in Brazil. Quim Nova 32(3):722–731. https://doi.org/10.1590/S0100-4042200900 0300031
- Zhang Q, Margaryan A, Schneidmiller R (2011) Methods for repelling insects using sesquiterpene hydrocarbons and their derivatives. United States Patent. https://patentimages.storage.googleapis.com/75/55/1a/a4004204fe518c/US8057829.pdf
- Zhang SF, Zhang Z, Kong XB, Wang HB, Liu F (2018) Dynamic changes in chemosensory gene expression during the *Dendrolimus punctatus* mating process. Front Physiol 8:1127. https://doi.org/10.3389/fphys.2017.01127
- Zhang G, Han J, Chen Y, Xiong J et al (2022) Generation mechanism and dual-dynamics simulation of surface patterns in single-point diamond turning of single-crystal copper. J Manuf Process 75:1023–1038. https://doi.org/10.1016/j.jmapro.2022.02.023
- Zhou JJ (2010) Odorant-binding proteins in insects. Vitam Horm 83:241–272. https://doi.org/10.1016/S0083-6729(10)83010-9
- Zhou T, Lu Y, Wang B (2010) Integrating TTF and UTAUT to explain mobile banking user adoption. Comput Hum Behav 24:760–767. https://doi.org/10.1016/j.chb.2010.01.015
- Zucchi R (2015) Mosca do mediterrâneo, Ceratitis capitata (Wiedemann). FEALQ, Piracicaba

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

