

# **Modelling pesticide uptake by potatoes through an ordinary differential equation coupled with a dispersion-advection equation**

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## **1 Introduction**

Potato plants are cultivated in more than one hundred countries across different continents because of their extraordinary ability to adapt to different weather and soil conditions, surpassed only by wheat, rice and corn, being consumed by more than one billion people. With a global production of 324 million tons in 2010, potato is the fourth most important food crop (or fifth if we consider sugar cane, a plant that is not always consumed as food) and it is one of the most important vegetables consumed worldwide [1]. Potato plants' agronomic efficiency guarantees the use of several types of soils destined to food production, which contributes to increase the total potato cropping area in a global scenario of fast population growth and economic development. All over the world, the main limiting factor to potato cropping is the plant's susceptibility to a great number of pests and diseases, some of them capable of causing serious damage to potato production. This obliges the use of many and varied types of pesticides that may cause serious environmental and alimentary problems [2-4].

Even when taking into account that the most recent agronomic management techniques, suggested by the integrated production systems, reduce risks of environment and food contamination it is very important that managers, technicians and researchers learn how to estimate the cumulative potential of pesticides in potatoes, which will enable them to recommend new products and technologies in order to have economically and environmentally sustainable productions. The United Nations Food and Agriculture Organization is currently promoting the tuber as an efficient food crop (potatoes are low-fat, high-carbohydrate food) that can improve food security in developing countries. Approximately 80% of the potato plant can be used for human consumption, a significantly higher percentage than that of cereals like corn and wheat.

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Several studies indicate the presence of pesticides and organic substances in potatoes [5-15] but none of them present a theoretically acceptable model that one can use to estimate pesticide concentrations in potato and in soil solution by the ordinary differential equation (ODE) coupled with a dispersion-advection equation (DAE) system. Three potato-specific plant uptake models were recently published [13,14,16], using Fick's second law for modelling the diffusive flow of a pesticide through potato tissues. These three models enable the estimation of pesticide bioconcentration factors in potato tubers, by using a known concentration of pesticide in soil. Trapp and co-workers [16] suggest that the transfer of pesticide mass occurs through the potato tissues, predominantly by the soil solution. This model makes it possible to estimate the diffusivity of organic substances in soil for potatoes, and helps significantly to elaborate useful mathematical models to determine the potato's concentration of non-ionic organic pesticides in the soil solution. Paraíba and Kataguirí [13], based on the work of Trapp and co-workers [16], presented a model that estimates the bioconcentration factor (BCF) of pesticides in potatoes supposing that the pesticide in the soil solution is absorbed by the potato by passive diffusion, following Fick's second law and the pesticides' degrade in the soil according to a first-order kinetic equation. Juraske and co-workers [14] also presented and evaluated a dynamic model of first-order kinetic equation for uptake of pesticides in potatoes, with measurements performed in a field trial in the region of Boyacá, Colombia.

It is extremely difficult to assess pesticides' environmental fate, as some of these substances are quite mobile that can take different pathways before they are effectively removed from the soil system. From the water and potato contamination point of view, soil plays a central role in pesticides uptake and transport. Soil is the main geologic compartment where degradation can occur and pesticides are likely to be attenuated before they leach to groundwater or uptake to potato. The soil capability to adsorb (sorption + adsorption) pesticides is affected by the occurrence of organic matter and soil mineral constituents, and soil ionic charge. Additionally, pesticides potential to leach to groundwater is affected by soil permeability, soil texture, water table fluctuations, and water content. Pavlis and co-workers [17] present a cross-analysis of different models of pesticide fate and the environmental indicators to underscore their relative strengths, weaknesses, similarities and dissimilarities.

The objective of this work is to present a model that simulates pesticide uptake by potatoes using a system with two differential equations, where one equation describes pesticide uptake by potatoes and the other one describes the pesticide lixiviation in soil. It was assumed that uptake and lixiviation occur, respectively, by diffusion of the pesticide through

potato tissues and by diffusion and advection of the pesticide in soil profile. Uptake was described by an ordinary differential equation (ODE) that was coupled to a dispersion-advection equation (DAE). These equations describe the kinetics of the pesticide in potatoes and in soil profile, respectively. The coupled equations (ODE-DAE) were numerically solved through a finite difference method, programmed in Matlab<sup>®</sup> system, and used to simulate uptake of pesticides by potatoes.

## 2 Materials and methods

### 2.1 Mathematical modelling of pesticide uptake by potatoes, using an ODE coupled with a DAE

#### 2.1.1 Model development

The pesticides' uptake by potato from the soil solution, by passive diffusion, and the dilution by growth and enzymatic metabolism of pesticide in potato were modelled by

$$\frac{dC_p}{dt} = k_u C_w(\delta_r, t) - k_p C_p \quad (1)$$

where  $C_p$  ( $\text{mg kg}^{-1}$ ) is the pesticide concentration in the potato,  $C_w = C_w(\delta_r, t)$  ( $\text{mg L}^{-1}$ ) is the pesticide concentration in soil solution in a day  $t$  (day),  $\delta_r$  (m) is the average depth of a potato inside the soil,  $k_p$  ( $\text{day}^{-1}$ ) is the dilution and metabolism rate, and  $k_u$  ( $\text{L kg}^{-1} \text{day}^{-1}$ ) is the rate of pesticide uptake by the potato. The initial concentration of pesticide in potatoes is assumed to be null, that is,  $C_p(0) = 0$ .

The pesticide uptake rate of soil solution was estimated supposing a passive diffusion through potato tissues with the diffusion coefficient as in [16], given by  $k_u = \frac{23D_p}{(r_p)^2 \rho_p k_{sw}}$ , where  $D_p$  ( $\text{m}^2 \text{day}^{-1}$ ) is the effective diffusion coefficient of pesticide through potato tissues,  $r_p$  (m) is the average radius of the potato,  $\rho_p$  ( $\text{kg L}^{-1}$ ) is the density of the potato, and  $k_{sw}$  (dimensionless) is the soil-water partition coefficient of the pesticide. Potato density is necessary to define correctly the pesticide uptake rate with a measured unit of  $\text{L kg}^{-1} \text{day}^{-1}$ , while the pesticide uptake rate is inversely proportional to potato density, see [18].

The dimensionless soil-water partition coefficient of the pesticide was calculated by  $k_{sw} = \rho_s f_{oc} k_{oc} + f_w + f_a k_{aw}$ , where  $\rho_s$  ( $\text{L kg}^{-1}$ ) is the total soil density. The  $f_{oc}$ ,  $f_w$  and  $f_a$  coefficients are the volumetric fractions of organic carbon, water and air of the soil,

respectively. The parameter  $k_{oc}$  ( $\text{L kg}^{-1}$ ) is the pesticide organic carbon-water partition coefficient and it was estimated by the expression given by  $k_{oc} = 10^{(0.1+0.81\log k_{ow})}$ , see [19,20], where  $k_{ow}$  is the pesticide's octanol-water partition coefficient and  $k_{aw}$  (dimensionless) is the pesticide's air-water partition coefficient that is estimated by  $k_{aw} = \frac{P_v P_m}{S_w R_g (273+T)}$ , where ( $T = 25^\circ\text{C}$ ) is the air temperature, ( $R_g = 8.314\text{Pa m}^3\text{mol}^{-1}$ ) is the universal gas constant,  $P_v$  (Pa) is the pesticide's vapour pressure,  $P_m$  ( $\text{g mol}^{-1}$ ) is its molar mass and  $S_w$  ( $\text{g m}^{-3}$ ) is its water solubility.

The pesticide's coefficient of effective diffusion through potato tissues was estimated by  $D_p = p_w \tau_w D_w$ , where  $\tau_w$  (dimensionless) is a coefficient of soil tortuosity and  $p_w$  (dimensionless) is the volumetric fraction of pesticide dissolved in the water phase of potato tissue, calculated by  $p_w = w_p / k_{pw}$ , in which  $w_p$  is the pore water fraction in the potato tissue,  $k_{pw}$  (dimensionless) is the potato-water partition coefficient of the pesticide, estimated by the equation  $k_{pw} = w_p + f_{chp} k_{chw} + 0.82 L_p (k_{ow})^{0.77}$  [16], where  $f_{chp}$  and  $L_p$  are respectively the volumetric fractions of carbohydrate and lipid of the potato tissue and  $k_{chw}$  is the pesticide's carbohydrate-water partition coefficient [21]. As an approximation, we thus assume  $k_{chw}$  to be 0.1 for compounds with  $\log k_{ow} < 0$ ; 0.2 for  $0 \leq \log k_{ow} < 1.0$ ; 0.5 for  $1.0 \leq \log k_{ow} < 2.0$ ; 1 for  $2.0 \leq \log k_{ow} < 3.0$  for; 2 for  $3.0 \leq \log k_{ow} < 4.0$  and 3 for  $\log k_{ow} \geq 4.0$  [21]. The tortuosity factor is calculated using the method of Millington and Quirk, see [22,23], and is given by  $\tau_w = (f_w)^{10/3} / (f_w + f_a)^2$ . The pesticide diffusivity in soil solution,  $D_w$  ( $\text{m}^2\text{day}^{-1}$ ), was estimated by  $D_w = \frac{4.93 \times 10^{-6} T_w \sqrt{\phi_w w_m}}{\xi_w (v_m)^{0.6}}$ , where ( $T_w = 298^\circ\text{K}$ ) is the average soil temperature,  $\phi_w = 2.6$  is an association term for the solvent (water),  $w_m = 18\text{g mol}^{-1}$  is the molar mass of water,  $\xi_w = 8.9 \times 10^{-1}\text{cp}$  is the water viscosity and  $v_m$  ( $\text{cm}^3\text{mol}^{-1}$ ) is the molar volume of the pesticide, see [23].

Pesticide concentration in soil solution was described by a dispersion-advection equation (DAE) given by

$$D_e \frac{\partial^2 C_w}{\partial z^2} - v_w \frac{\partial C_w}{\partial z} - R_f \left( \frac{\partial C_w}{\partial t} + k_s C_w \right) = 0 \quad (2)$$

where  $D_e$  ( $\text{m}^2\text{day}^{-1}$ ) is the soil solution's coefficient of effective dispersion,  $C_w = C_w(z, t)$  ( $\text{mg L}^{-1}$ ) is the pesticide concentration in soil solution,  $R_f$  is the retardation factor of pesticide in soil,  $v_w$  ( $\text{m day}^{-1}$ ), is the speed of water flow in soil pores,  $\rho_w$  ( $\text{kg L}^{-1}$ ) is the wet density of the soil and  $k_s$  ( $\text{day}^{-1}$ ) is the pesticide's degradation rate in soil, which in turn is estimated by  $k_s = \ln(2)/t_{1/2}$ , where  $t_{1/2}$  (day) is the pesticide's half-life in the soil. The retardation factor was estimated by  $R_f = 1 + \frac{\rho_s f_{oc} k_{oc} + f_a k_{aw}}{f_w}$ .

According to Jury et al (1992), the coefficient of effective dispersion is the sum of the pesticide's molecular diffusion in soil solution plus the soil solution's hydrodynamic dispersion, estimated by  $D_e = \tau_w D_w + \alpha_s v_w$ , where  $f_w$  is the volumetric fraction of water in soil in the field capacity and  $\alpha_s$  (m) is a factor expressing water dispersion in the soil. The term  $\alpha_s v_w$  ( $\text{m}^2\text{day}^{-1}$ ) is the coefficient of hydrodynamic dispersion and  $\tau_w$  is a tortuosity coefficient, see [23,24].

The thin top layer of the soil that initially contains the pesticide is considered as part of the soil profile and its existence is incorporated to the initial conditions. The modelled soil profile has a form of  $z = 0$  up to  $z = \varepsilon$ . The initial conditions are then given by

$$C_w(z, 0) = \frac{10^{-4} \times ad}{\varepsilon(\rho_s k_{oc} f_{oc} + f_w)}, \quad 0 < z < \varepsilon \quad (3)$$

and

$$C_w(z, 0) = 0, \quad \varepsilon < z < +\infty \quad (4)$$

where  $ad$  ( $\text{g ha}^{-1}$ ) is the pesticide application dose and  $\varepsilon$  (m) is the width of the top layer of soil surface that describes the initial area, where takes place the incorporation of pesticides applied to the soil. The model given by Equation 2 needs two boundary conditions; in the definition of the boundary condition, it is assumed that the water added on top of the soil is free from pesticides and that there is not a concentration gradient in the soil layer of infinite depth. Then, the boundary conditions for Equation 2 are given by

$$v_w C_w(0, t) - D_e \frac{\partial C_w(0, t)}{\partial z} = 0, \quad t \geq 0 \quad (5)$$

and

$$\frac{\partial C_w(+\infty, t)}{\partial z} = 0, \quad t \geq 0 \quad (6)$$

where all the terms have already been defined. Numerical solutions of the coupling between the pesticide concentration in potatoes, as given by the ODE defined by Equation 1, and the pesticide concentration in soil solution as given by the DAE, defined by Equations 2-6, were obtained by using a finite difference method similar to those used by Contreras and co-workers [25].

### 2.1.2 Input data

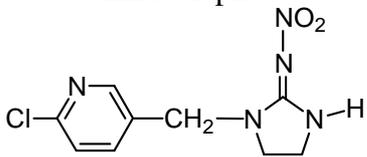
We have selected the imidacloprid insecticide to simulate the effects that pesticide leaching and degradation in the soil have upon pesticide accumulation in potatoes. Imidacloprid (1-[(6-chloro-3-pyridinyl)methyl]-N-nitro-2-imidazolidinimine) is a neonicotinoid insecticide that acts biochemically as an antagonist by binding to postsynaptic nicotinic receptors in the central nervous system of insects. It is a systemic insecticide with translaminar activity and with contact and stomach action. Readily taken up by the plant, it is then distributed acropetally with good root-systemic action. Imidacloprid is one of the most widely used insecticides and it can be applied by soil injection, tree injection, application to the skin, broadcast foliar, ground application as granular or liquid formulation or as a pesticide-coated seed treatment. In potatoes, it is recommended for the control of aphids (*Myzus persicae*), thrips (*Thrips palmi*), beetles (*Diabrotica speciosa*) and other insects. It is also effective against insects that attack barley, beans, broccoli, cabbage, cauliflower, chicory, chrysanthemum, citrus, coffee, corn, cotton, cucumbers, eggplant, garlic, gerbera, grapes, kale, lettuce, melon, oats, onions, peanuts, peppers, pineapple, poinsettia, pumpkin, rice, sorghum, soybean, squash, sugar cane, sunflower, tobacco, tomatoes, watermelon and wheat. Doses used for foliar applications are between 25-100 g ha<sup>-1</sup>, while for seed treatment the doses are 50-175 g/100 kg<sup>-1</sup> for seeds in general, and 350-700 g/100 kg<sup>-1</sup> for cotton seeds [26]. Metabolites of imidacloprid have a broad range of toxicities, with some of them showing stronger insecticidal activity than the parent compound and several of them demonstrating chronic toxicity to honeybees in dosing studies, see [27,28].

Imidacloprid has been reported to persist in different concentrations in stems and roots of plants, depending on the application method [29]. Thus, the pesticide can be an important hazard to the environment because it is categorized as moderately toxic and with a high potential to leach into groundwater [30,31]. Imidacloprid can persist for a period longer than 90 days under dry or cold conditions, and is moderately to highly mobile in soils with low clay or organic matter content. Because it does not adsorb strongly to soil particles and has a

lengthy half-life of more than 90 days and moderate to high solubility in water ( $610 \text{ mg L}^{-1}$ ), it has a high potential for groundwater contamination [32-35]. Imidacloprid was found in fresh potatoes sold in farmers' markets in Alberta, Canada, at concentrations ranging from 15-31  $\mu\text{g kg}^{-1}$  [15].

Table 1 presents the physicochemical properties of imidacloprid necessary for the model given by Equations 1-6 to simulate both the uptake of insecticide imidacloprid by potatoes and the lixiviation in soil. Table 2 presents the values of potato and soil which were used in numerical simulation by Equations 1-6. In these simulations an implicit numerical scheme was used similar to the one used by Freijer and co-workers [24], with an integration time step of  $5.0 \times 10^{-3}$  days and an integration depth step of  $1.0 \times 10^{-2}$  meters. In the initial conditions given by Equations 3-4, the depth of the superficial soil layer was assumed to be  $\varepsilon = 1.0 \times 10^{-2}$  meters, in a soil profile of 1.0 meter of depth ( $0.01 \leq z \leq 1.0$ ). The total time adopted for the simulation was 120 days, which corresponds to the longest period until potato harvest, after pesticide is applied in the soil on day zero, and an initial imidacloprid concentration of  $100 \text{ g ha}^{-1}$  (Equation 3) was assumed for the soil solution.

Table 1. Imidacloprid physicochemical characteristics.

imidacloprid	
Common name	imidacloprid
Molecular structure	
CAS RN	138261-41-3
Field use and class	Insecticide and neonicotinoid
Chemical Abstracts name	1-[(6-chloro-3-pyridinyl)methyl]-N-nitro-2-imidazolidinimine
IUPAC name	1-(6-chloro-3-pyridylmethyl)-N-nitroimidazolidin-2-ylideneamine
Molecular formula	$\text{C}_9\text{H}_{10}\text{ClN}_5\text{O}_2$
Half-life in the soil: $t_{1/2}$	120 days
Molar volume: $V_m$	$160.1 \text{ cm}^3 \text{ mol}^{-1}$
Molecular weight: $P_m$	$255.67 \text{ g mol}^{-1}$
Octanol-water partition coefficient: $K_{ow}$	3.71 ( $\log K_{ow} = 0.57$ )
Organic carbon partition coefficient: $K_{oc}$	$k_{oc} = 10^{(0.1+0.81 \log K_{ow})} \text{ L kg}^{-1}$
Vapor pressure: $P_v$	$4.0 \times 10^{-10} \text{ Pa}$
Water solubility: $S_w$	$610 \text{ g m}^{-3}$

Sources: Available at <https://scifinder.cas.org/scifinder/view/scifinder/scifinderExplore.jsf>; <http://www.syrres.com/what-we-do/databaseforms.aspx?id=386> and [26].

Table 2. Physicochemical parameters for potato plants and soil applied to the ODE-DAE model to estimate concentrations of pesticides in potatoes.

Parameter	Symbol	Value	Unit
potato water volumetric content <sup>a</sup>	$w_p$	0.778	L L <sup>-1</sup>
potato lipid volumetric content <sup>a</sup>	$L_p$	0.001	L L <sup>-1</sup>
potato carbohydrate volumetric content <sup>a</sup>	$f_{chp}$	0.154	L L <sup>-1</sup>
pesticide dissipation in potato <sup>a</sup>	$k_p$	0.142	day <sup>-1</sup>
potato density <sup>b</sup>	$\rho_p$	1.4	kg L <sup>-1</sup>
average potato sphere ray <sup>a</sup>	$r_p$	0.03	m
soil-organic carbon volumetric fraction <sup>a</sup>	$f_{oc}$	0.02	L L <sup>-1</sup>
soil-water volumetric fraction <sup>a</sup>	$f_w$	0.25	L L <sup>-1</sup>
soil-air volumetric fraction <sup>a</sup>	$f_a$	0.12	L L <sup>-1</sup>
soil density on dry base <sup>a</sup>	$\rho_s$	1.1	kg L <sup>-1</sup>
maximum depth of potato plants roots	$\delta_r$	0.10	m
initial concentration of pesticide in soil solution	$C(0,0)$	1.0	mg L <sup>-1</sup>
initial concentration of pesticide in potato	$C_p(0)$	0.0	mg kg <sup>-1</sup>
water speed in soil pores	$v_w$	0.031	m day <sup>-1</sup>
longitude of dispersion in the soil	$\alpha_s$	0.005	m
soil porosity	$\sigma_s$	0.37	L L <sup>-1</sup>
soil tortuosity factor	$\tau_w$	0.183	-

<sup>a</sup>See [16]; <sup>b</sup>Available at <http://www.starch.dk/isi/starch/tm5www-potato.asp>

### 3. Numerical simulation, results and discussion

The value of octanol-water partition coefficient of 3.71 ( $\log k_{ow} = 0.57$ ) indicates that imidacloprid has a low lipid affinity. The value of organic carbon-water partition coefficient,  $3.64 \text{ L kg}^{-1}$  ( $k_{oc}$ ), indicates that imidacloprid has a low affinity with soil organic matter, which means that its translocation from soil solution to potato cannot be difficult. The value of  $6.77 \times 10^{-14}$  obtained for the air-water partition coefficient ( $k_{aw}$ ) using data from Table 1,

indicates that imidacloprid is not volatile at the temperature of 25°C [36]. The value of 2.60 for the pesticide uptake rate ( $k_u$ ) indicates good imidacloprid mobility in the potato by diffusion [16]. The value of 0.81 for the potato to water partition coefficient ( $k_{pw}$ ) indicates that imidacloprid has a moderate affinity with potato material, see [13,16].

The imidacloprid concentration in potatoes over time, as simulated by the ODE-ADE model, is illustrated in Figure 1. Initially, the compound concentration is null, then it increases continuously up to a maximum value, after which it decreases with time. This concentration pattern is due to the compartment system described by the ODE model and it was assumed for the pesticide in potato dissipation kinetic of first order, Equation 1. At the 6th day ( $t_{max} = 6.7$ ) after imidacloprid application at the first day ( $t = 0$ ) to the soil, its concentration in potatoes reaches the simulated maximum value of  $0.15 \text{ mg kg}^{-1}$  ( $C_p(t_{max}) = 0.15 \text{ mg kg}^{-1}$ ). In similar simulations with the azoxystrobin fungicide we found values for  $t_{max}$  and  $C_p(t_{max})$  of 43 days and  $3.6 \times 10^{-3} \text{ mg kg}^{-1}$ , respectively. Concentrations estimated by the ODE-ADE model are consistent with concentration values for imidacloprid and azoxystrobin that were observed experimentally in commercial potatoes in Alberta, Canada, by Thompson and co-workers [15].

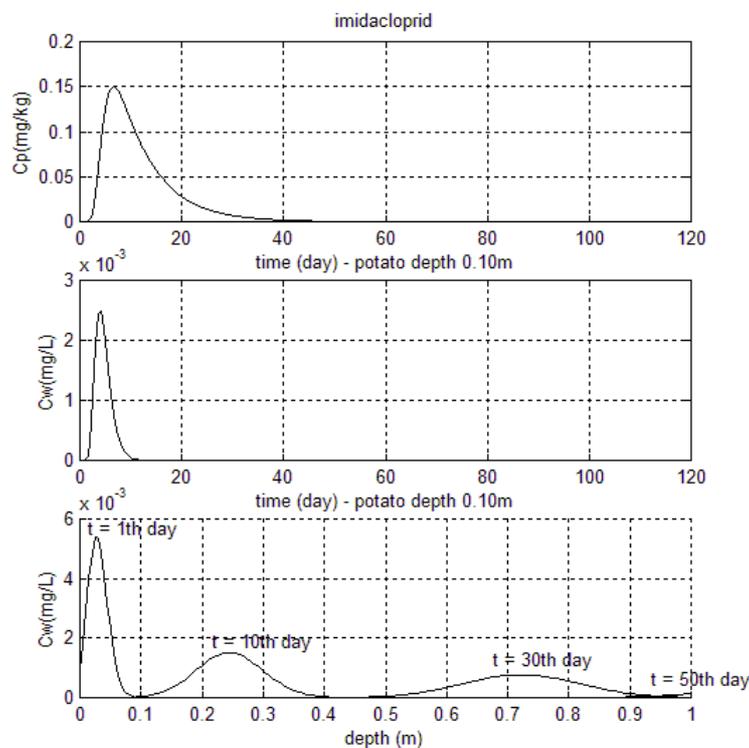


Figure 1. Concentrations along time of the imidacloprid insecticide in potato and soil, simulated by means of an ODE\_ADE model.

Figure 1 presents imidacloprid concentrations resulting from simulations of the uptake by potatoes of imidacloprid in soil solution,  $C_w(t)$ , at the average depth of potatoes in the soil – 10cm ( $\delta_r = 0.10\text{m}$ ). For depths between 0.0 and 1.0m and times  $t = 1, t = 10, t = 30, t = 50$  days, Figure 1 presents concentrations of imidacloprid resulting from dispersion and advection of the solution along the soil profile. The overall pattern of imidacloprid concentrations along the soil profile indicates that potatoes located at depths of over 0.20m would be less exposed to absorbing imidacloprid from the soil solution. The concentration pattern along the soil profile will have an influence on imidacloprid uptake by potatoes.

### 3.1 ODE-ADE model sensitivity analysis

Data in Table 1 and Table 2, together with the numerical method programmed in Matlab<sup>®</sup>, were used to analyze the effect of the variation of a potato's average depth into the soil,  $\delta_r$  (m), on the maximum concentration of imidacloprid in potatoes,  $C_p(t_{\max})$ , and on the time taken for the maximum concentration to occur in potatoes,  $t_{\max}$ . So, the model was executed for values of  $\delta_r$  between 0.05m and 0.4m. Figure 2 presents values of  $C_p(t_{\max})$  and  $t_{\max}$  obtained by the ODE-ADE model for values of  $\delta_r$  in  $[0.05; 0.40]$ . It can be observed in Figure 2 that values of  $C_p(t_{\max})$  decline uniformly according to an apparent negative exponential pattern and that  $t_{\max}$  grows linearly as the average depths of potatoes in soil,  $\delta_r$ , increase, that is, potatoes that are deeper down in the soil have smaller values of maximum concentrations and these ones are reached at later times.

It is possible to carry out another sensitive analysis with the ODE-ADE model in Matlab<sup>®</sup> program to investigate the effect of the conjoint variations of the pesticide octanol-water partition coefficient and the pesticide half-life in soil, upon the maximum concentration of pesticide in potatoes and upon the maximum time concentration. For this, let potato and soil data be those of Table 1 and Table 2. The colour filled contour plots in Figure 3 and Figure 4 were obtained by conjoint varying of  $\log k_{ow}$  in  $[-1.0; 6.0]$  ( $-1.0 \leq \log k_{ow} \leq 6.0$ ) and half-life in  $[1; 200]$  ( $1 \leq t_{1/2} \leq 200$  days) and assuming  $\delta_r = 0.12\text{m}$ . It can be observed that the conjoint variation of the pesticide octanol-water partition coefficient and pesticide half-life in soil indicates that pesticides with  $\log k_{ow} < 1.0$  and half-life longer than 20 days ( $t_{1/2} > 20$ ) present the highest maximum concentrations in potatoes (Figure 3). On the other hand,

pesticides with  $\log k_{ow} > 3.0$  and half-life longer than 20 days ( $t_{1/2} > 20$ ) reach maximum concentrations in potatoes at later times (Figure 4).

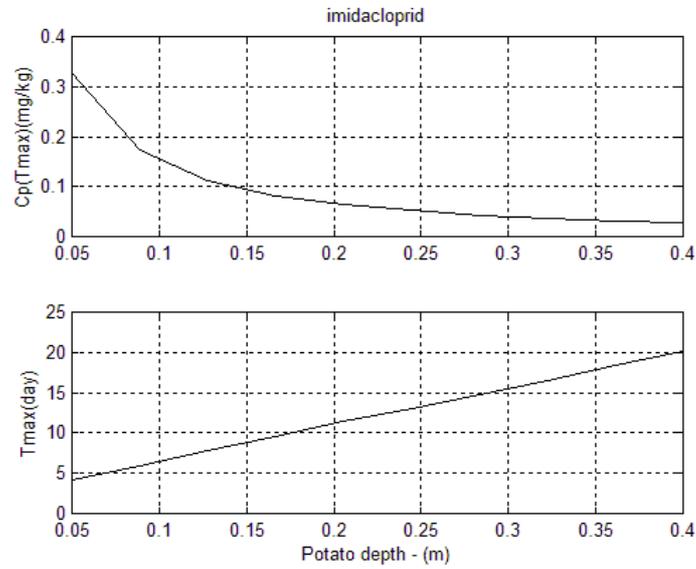


Figure 2. Sensitivity analysis of the maximum concentration  $C_p(t_{max})$  of the imidacloprid insecticide from soil solution in potato, and the time necessary to reach maximum pesticide concentration  $t_{max}$  in potato, simulated by means of an ODE\_ADE model.

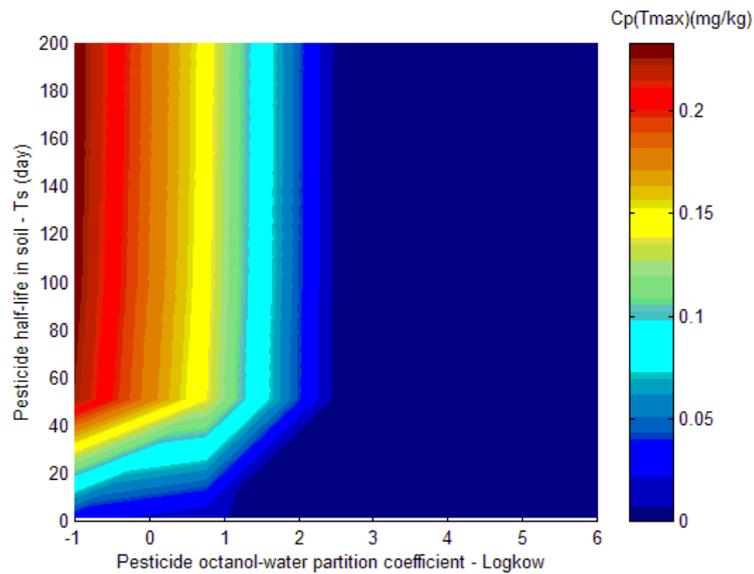


Figure 3. Color diagram representing the joint relation of pesticide octanol-water partition coefficient,  $\log k_{ow}$ , and pesticide half-life in soil,  $t_{1/2}$ , with the maximum pesticide concentration in potato,  $C_p(t_{max})$ , simulated by means of an ODE-ADE model.

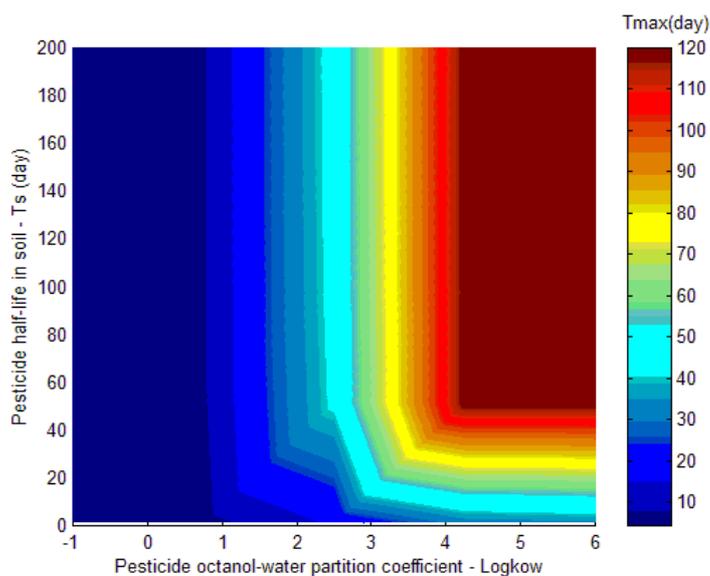


Figure 4. Color diagram representing the joint relation of pesticide octanol-water partition coefficient,  $\log k_{ow}$ , and pesticide half-life in soil,  $t_{1/2}$ , with the time necessary to reach maximum pesticide concentration in potato,  $t_{max}$ , simulated by means of an ODE-ADE model.

#### 4. Conclusions

The ODE-ADE model presented in this work allows estimation of pesticide concentrations in potato and in soil solution. Pesticides are unstable organic compounds that undergo degradation reactions in soils and can be taken up by potatoes from the soil solution through diffusion. The ODE-ADE model sensitivity analysis, regarding the pesticide octanol-water partition coefficient, the pesticide half-life in soil and the average depth of potatoes proved that soil depth affects the quantity of pesticide concentrations in potato and, consequently, affects the pesticide maximum concentration and the time it takes to reach maximum concentration. According to the model, potatoes in deeper layers may be more protected against contamination as much because they are exposed to lesser concentrations of pesticides that suffer dispersion when leached, as because such exposure occurs later on the crop cycle. Based on the model's results, was observed that values of the octanol-water partition coefficient and of the pesticide's half-life in soil have great influence upon the uptake of pesticides by potatoes. Specifically for imidacloprid, it reaches simulated maximum concentration of  $0.15 \text{ mg kg}^{-1}$  ( $C_p(t_{max}) = 0.15 \text{ mg kg}^{-1}$ ) in potatoes during 6<sup>th</sup> day ( $t_{max} = 6.7$ ) after application to the soil.

It is possible to develop agronomic strategies for both crop management and for use of pesticides in potatoes. It is even possible to define industrial goals, based on mathematical

modelling, for the efficient formulation of pesticides that bring benefits to the soil environment and avoid pesticide uptake by potatoes. This modelling tool can be used as a computationally efficient aid to field study design, model calibration (changes in what input parameters have the greatest impact on model output), and to further our understanding of the physical mechanisms involved in pesticide uptake by potatoes and pesticide transport in soil.

## 5. References

- [1] FAO. **FAOSTAT database**. Rome, Italy: Food and Agriculture Organization. Disponível em: <http://faostat.fao.org/site/339/default.aspx>.
- [2] Caldas, E. D.; Miranda, M. C. C.; Conceição, M. H.; de Souza, L. Dithiocarbamates residues in Brazilian food and the potential risk for consumers. **Food and Chemical Toxicology**. v. 42, n. 11, p. 1877–883, 2004.
- [3] López-Pérez, G. C.; Arias-Esteévez, M.; López-Periago, E.; Soto-González, B.; Cancho-Grande, B.; Simal-Gándara, J. Dynamics of pesticides in potato crops. **Journal of Agricultural and Food Chemistry**. v. 54, n. 5, p. 1797–1803, 2006.
- [4] Leistra, M.; van Den Berg, F. Volatilization of parathion and chlorothalonil from a potato crop simulated by the PEARL model. **Environmental Science & Technology**. v. 41, n. 7, p. 2243–2248, 2007
- [5] Dogheim, S. M.; El-Marsafy, A. M.; Salama, E. Y.; Gadalla, S. A.; Nabil, Y. M. Monitoring of pesticide residues in Egyptian fruits and vegetables during 1997. **Food Additives and Contaminants**. v. 19, n. 11, p. 1015–1027, 2002.
- [6] Fismes, J.; Perrin-Ganier, C.; Empereur-Bissonnet, P.; Morel, J. L. Soil-to-root transfer and translocation of polycyclic aromatic hydrocarbons by vegetables grown on industrial contaminated soils. **Journal of Environmental Quality**. v. 31, n. 5, p. 1649–1656, 2002.
- [7] Samsøe-Petersen, L.; Larsen, E. H.; Larsen, P. B.; Bruun, P. Uptake of trace elements and PAHs by fruit and vegetables from contaminated soils. **Environmental Science & Technology**. v. 36, n. 14, p. 3057–3063, 2002.
- [8] Jensen, A. F.; Petersen, A.; Granby, K. Cumulative risk assessment of the intake of organophosphorus and carbamate pesticides in the Danish diet. **Food Additives and Contaminants**. v. 20, n. 8, p. 776–785, 2003.
- [9] Poulsen, M. E.; Andersen, J. H. Results from the monitoring of pesticide residues in fruit and vegetables on the Danish market, 2000-01. **Food Additives and Contaminants Part A**. v. 20, n.8, p. 742–757, 2003.

- [10] Rissato, S. R.; Galhiane, M. S.; de Souza, A. G.; Apon, B. M. Development of a supercritical fluid extraction method for simultaneous determination of organophosphorus, organohalogen, organonitrogen and pyrethroids pesticides in fruit and vegetables and its comparison with a conventional method by GCECD and GCMS. **Journal of the Brazilian Chemical Society**. v. 16, n. 5, p. 1038–1047, 2005
- [11] Cesnik, H. B.; Gregorcic, A.; Bolta, S. V.; Kmecl, V. Monitoring of pesticide residues in apples, lettuce and potato of the Slovene origin, 2001-04. **Food Additives and Contaminants**. v. 23, n. 2, p. 164–173, 2006.
- [12] Zohair, A.; Salim, A. B.; Soyibo, A. A.; Beek, A. J. Residues of polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs) and organochlorine pesticides in organically-farmed vegetables. **Chemosphere**. v. 63, n. 4, p. 541–553, 2006.
- [13] Paraíba, L. C.; Kataguri, K. Model approach for estimating potato pesticide bioconcentration factor. **Chemosphere**. v. 73, n. 8, p. 1247–1252, 2008.
- [14] Juraske, R.; Vivas, C. S. M.; Velasquez, A. E.; Santos, G. G.; Moreno, M. B. B.; Gomez, J. D.; Binder, C. R.; Hellweg, S.; Dallos, J. A. G. Pesticide Uptake in Potatoes: Model and Field Experiments. **Environmental Science & Technology**. v. 45, n. 2, p. 651–657, 2011.
- [15] Thompson, T. S.; Dimock, R.; Bradbury, R. W.; Rieve, R.; Fehr, M. Pesticides in Fresh Potatoes Sold in Farmers' Markets in Alberta, Canada. **Bulletin of Environmental Contamination and Toxicology**. v. 87, n. 5, p. 580–585, 2011.
- [16] Trapp, S.; Cammarano, A.; Capri, E.; Reichenberg, F.; Mayer, P. Diffusion of PAH in potato and carrot slices and application for a potato model. **Environmental Science & Technology**. v. 41, n. 9, p. 3103–3108, 2007.
- [17] Pavlis, M.; Cummins, E.; McDonnell, K. Groundwater vulnerability assessment of plant protection products: a review. **Human and Ecological Risk Assessment**. v. 16, n. 3, p. 621–650, 2010.
- [18] J. Crank. **The Mathematics of diffusion**. Oxford: Clarendon Press. 1975.
- [19] EC. **EUSES, the European Union System for the Evaluation of Substances National Institute of Public Health and the Environment (RIVM), Bilthoven, the Netherlands**. Ispra, Italy: Prepared for the European Chemicals Bureau, Joint Research Centre. 1996. Disponível em:  
[http://ihcp.jrc.ec.europa.eu/our\\_activities/public-health/risk\\_assessment\\_of\\_Biocides/euses](http://ihcp.jrc.ec.europa.eu/our_activities/public-health/risk_assessment_of_Biocides/euses).
- [20] van Beelen, P. **The risk evaluation of difficult substances in USES 2.0 and EUSES: a decision tree for data gap filling of Kow, Koc, and BCF, (RIVM Report 679102050)**.

Bilthoven, the Netherlands: National Institute of Public Health and the Environment. 2000. Disponível em <http://www.rivm.nl/bibliotheek/rapporten/679102050.pdf>.

[21] Chiou, C. T.; Sheng, G. Y.; Manes, M. A partition-limited model for the plant uptake of organic contaminants from soil and water. **Environmental Science & Technology**. v. 35, n. 7, p. 1437–1444, 2001.

[22] Jury, W. A.; Horton, R. **Soil physics**. New York: John Wiley & Sons. 2004.

[23] Clark, M. M. **Transport Modeling for Environmental Engineers and Scientists**. New York: Wiley-Interscience Publication. 1996.

[24] Freijer, J. I.; Veling, E. J. M.; Hassanizadeh, S. M. Analytical solutions of the convection-dispersion equation applied to transport of pesticides in soil columns. **Environmental Modelling & Software**. v. 13, n. 2, p. 139–149, 1998.

[25] Contreras, W. A.; Ginestar, D.; Paraiba, L. C.; Bru, R. Modelling the pesticide concentration in a rice field by a level IV fugacity model coupled with a dispersion-advection equation. **Computers & Mathematics with Applications**. v. 56, n. 3, p. 657–669, 2008.

[26] Tomlin, C. D. S. **The pesticide manual**. Farnham, UK: British Crop Protection Council. 2000.

[27] Nauen, R.; Reckmann, U.; Armbrorst, S.; Stupp, H. P.; Elbert, A. Whitefly-active metabolites of imidacloprid: biological efficacy and translocation in cotton plants. **Pesticide Science**. v. 55, n. 3, p. 265–271, 1999.

[28] Suchail, S.; Guez, D.; Belzunces, L. P. Discrepancy between acute and chronic toxicity induced by imidacloprid and its metabolites in *Apis mellifera*. **Environmental Toxicology and Chemistry**. v. 20, n. 11, p. 2482–2486, 2001.

[29] Juraske, R.; Castells, F.; Vijay, A.; Muñoz, P.; Antón, A. Uptake and persistence of pesticides in plants: Measurements and model estimates for imidacloprid after foliar and soil application. **Journal of Hazardous Materials**. v. 165, n. 1-3, p. 683–689, 2009.

[30] WHO. **Recommended classification of pesticides by hazard and guidelines to the classification: 2009**. Geneva, Switzerland: World Health Organization. 2009.

[31] Fernandez-Gomez, M. J.; Romero, E.; Nogales, R. Impact of imidacloprid residues on the development of *Eisenia fetida* during vermicomposting of greenhouse plant waste. **Journal of Hazardous Materials**. v. 192, n. 3, p. 1886–1889, 2011.

[32] Scorza, R. P.; Boesten, J. Simulation of pesticide leaching in a cracking clay soil with the PEARL model. **Pest Management Science**. v. 61, n. 5, p. 432–448, 2005.

- [33] Scorza, R. P.; Smelt, J. H.; Boesten, J.; Hendriks, R. F. A.; van der Zee, S. Preferential flow of bromide, bentazon, and imidacloprid in a dutch clay soil. **Journal of Environmental Quality**. v. 33, n. 4, p. 1473–1486, 2004.
- [34] Gonzalez-Pradas, E.; Flores-Cespedes, F.; Urena-Amate, M. D.; Fernandez-Perez, M.; Garratt, J.; Wilkins, R. Leaching and persistence of imidacloprid and diuron in a citrus crop in Valencia. **Fresenius Environmental Bulletin**. v. 9, n. 9-10, p. 638–645, 2000.
- [35] Nemeth-Konda, L.; Fuleky, G.; Morovjan, G.; Csokan, P. Sorption behaviour of acetochlor, atrazine, carbendazim, diazinon, imidacloprid and isoproturon on Hungarian agricultural soil. **Chemosphere**, v. 48, n. 5, p. 545–552, 2002.
- [36] Trapp, S.; Harland, B. Field test of volatilization models. **Environmental Science and Pollution Research**. v. 2, n. 3, p. 164–169, 1995.