

### Thermal and Rheological behavior of native and modified starch *Araucaria angustifolia* (pinhão)

<sup>1</sup>Polyanna Silveira Hornung, <sup>2</sup>Andressa Gabardo Granza, <sup>2</sup>Simone Rosa da Silveira Lazzarotto, <sup>1</sup>Rosemary Hoffmann Ribani, <sup>3</sup>Marcelo Lazzarotto, <sup>2</sup>Egon Schnitzler(\*)

<sup>1</sup>UFPR – Federal University of Paraná, Rua Coronel Francisco H. dos Santos, s/n. ZIP 81531-980 Curitiba, PR, Brazil.

<sup>2</sup>UEPG - Ponta Grossa State University, Av. Carlos Cavalcanti, 4748. ZIP 84030-900 Ponta Grossa, PR, Brazil. - (\*) [egons@uepg.br](mailto:egons@uepg.br)

<sup>3</sup>EMBRAPA FORESTS, Brazilian Agricultural Research Corporation, Estrada da Ribeira, Km 111 - P.O. Box 319. ZIP 83411-000 Colombo, PR, Brazil.

#### Abstract

*Araucaria angustifolia* is known in Brazil as Paraná Pine is endangered due to reckless extraction. The preservation and maintenance of the culture could be stimulated by use derivative products like the starch from its seeds (*pinhão*). In this work were studied samples of unmodified and modified *pinhão* starch. The hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) were used in the concentrations 0.1, 0.2 and 0.3 mol.L<sup>-1</sup> (a, b, c) and simultaneously exposed in ultraviolet rays ( $\lambda = 256$  nm) for one hour. One sample (N) was maintained as received for comparisons of the analysis results. The samples were analyzed by the thermoanalytical techniques: thermogravimetry and differential thermal analysis (TG-DTA) and differential scanning calorimetry (DSC); and by the rheological analysis: Rapid Viscoamylographic Analysis (RVA). The TGA showed an increase on the stability values form the (N) sample to the modified ones. The DSC showed a strong decrease on the gelatinization enthalpy with increasing of the H<sub>2</sub>O<sub>2</sub> concentrations solutions valued for the modified samples. The RVA results presented that the Setback reduced with the treatment that used hydrogen peroxide in a higher concentration. The obtained results showed that the modification method applied altered the thermal behavior of the *pinhão* starch.

#### Keywords

Modified Starch; Hydrogen Peroxide, Ultraviolet rays, *Araucaria angustifolia*, TG/DTA, DSC

#### Introduction

*Araucaria angustifolia* is a conifer, grows naturally in mixed forests in Brazil (south), Argentina, Paraguay and Chile, these trees can reach from 35 to 60 m in height and 0.8 to 2 m in diameter at breast height (dbh) [1]. Known locally as the Paraná pine, due to its imprudent extraction for commercial purposes or through deforestation, *Araucaria* is endangered. Research on the sustainable use of *Araucaria* derivatives could stimulate the preservation of this species. The seed of this pine tree, known as *pinhão*, is produced early in winter and constitutes a traditional high-calorie food that is usually consumed, after being cooked or roasted [2].

According to Bello-Pérez et al. [3] and Bicudo, et al. [4], the wet basis of raw seeds contains approximately: starch (36 %), proteins (3 %), lipids (1 %), soluble sugars (2.4 %), as well as fibres, minerals and phenolic compounds (< 0.2 %). Both the amount of starch and its type are critical for the texture of a given product [4].

Starch granules are made up of glucose polymers, amylose and amylopectin, and they are found inside vegetable cells. The glucose polymers that make up the starch come in two molecular forms, linear and branched. The first is referred to as amylose and the latter as amylopectin. Amylose is a linear polymer of  $\alpha$  1,4-linked glucose, whereas amylopectin is a highly branched polysaccharide consisting of  $\alpha$  1,4 -linked glucose with  $\alpha$  1,6- linkages at the branch points [5].

The use of starch in the food industry is to impart functional properties and to modify food texture and consistency. Efforts have been made to find native starches with the necessary properties (syneresis, transparency, freeze/thaw stability). In this way, the knowledge about the characteristics of different starches is important to select the most suitable for a specific application [3]. The modifications seek to improve the native starch so it becomes more efficient, a much studied technique is the oxidative modification [6].

The thermogravimetry analysis (TGA) can be helpful to show the behavior of starch granules when heating leads to depolymerization. The differential scanning calorimetry (DSC) is particularly appropriate to investigate the phase transitions of starch/water systems, because it allows the study of the starch gelatinization over a wide range of starch/water ratio, and the estimation of transition enthalpies [4].

The aim of this work was study the modification of *pinhão* starch by hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) in concentrations 0.1, 0.2 and 0.3 mol.L<sup>-1</sup> with simultaneously ultraviolet rays ( $\lambda = 256$  nm) exposition for one hour. In this way were investigate the changes using the following thermoanalytical techniques: thermogravimetry (TG), differential thermal analysis (DTA) and

differential scanning calorimetry (DSC), and the rheological parameters were analyzed by Rapid Viscoamylographic Analysis (RVA).

## Material and Methods

### Origin and modifications of the starch

The native *pinhão* starch (200 g) was bought in the commercial area of Colombo, PR, Brazil. The starch was divided into four parts of 50 g (dry basis). One of these was maintained as received and was designated the (N) untreated sample; the other three samples were treated following the literature methodology [7, 8] with modifications. The starches were treated with standard hydrogen peroxide ( $H_2O_2$ ) solutions for one hour of simultaneously exposure in ultraviolet rays ( $\lambda = 256$  nm). For samples (a), (b) and (c), 0.1, 0.2 and 0.3 mol.L<sup>-1</sup> of  $H_2O_2$  was used, respectively. The modified samples were then washed and filtered until the complete elimination of the reagent. The samples were dried in an oven with forced air circulation for 24 hours at 35 °C and then analysis was performed.

### Thermogravimetric study (TG/DTA)

The TG/DTA curves were obtained using a thermal analysis system (Shimadzu, DTG-60H model); the samples were heated from 30 °C to 600 °C using open alumina crucibles with approximately 5.0 mg of the sample under a nitrogen flow of 150 mL.min<sup>-1</sup> at a heating rate of 10 °C min<sup>-1</sup>. The instrument was preliminarily calibrated with standard weight and with standard calcium oxalate monohydrate. All mass loss percentages were determined using TA-60 WS data analysis software [8, 9].

### Differential scanning calorimetry (DSC)

The DSC curves were obtained using a thermal analysis system (TA-Instruments, DSC-Q200 model, USA). The DSC curves were recorded under an air flow of 50 mL.min<sup>-1</sup>, heating rate of 10 °C min<sup>-1</sup> and the samples weighed about 2.5 mg. The 4:1 (water:starch w/w) mixture was prepared and maintained for 60 minutes in order to equilibrate the moisture content. The aluminum crucibles were sealed and then the curves were performed. The instrument was previously calibrated with 99.99 % purity Indium, melting point of  $T_p = 156.6$  °C,  $\Delta H = 28.56$  J.g<sup>-1</sup> [8, 9].

### Rapid viscoamylographic analysis (RVA)

The pasting properties of the samples were obtained by using a viscometer (Newport Sci., RVA-4 model, Australia). A suspension of 3.0 g of starch in 25.0 g of distilled water underwent a controlled heating and cooling cycle under constant shear where it was held at 50 °C for two min, heated from 50 to 95 °C at 6 °C min<sup>-1</sup>, held at 95 °C for 5 min, cooled to 50 °C at 6 °C min<sup>-1</sup> and held at 50 °C for 2 min [9, 10].

### Statistical analysis

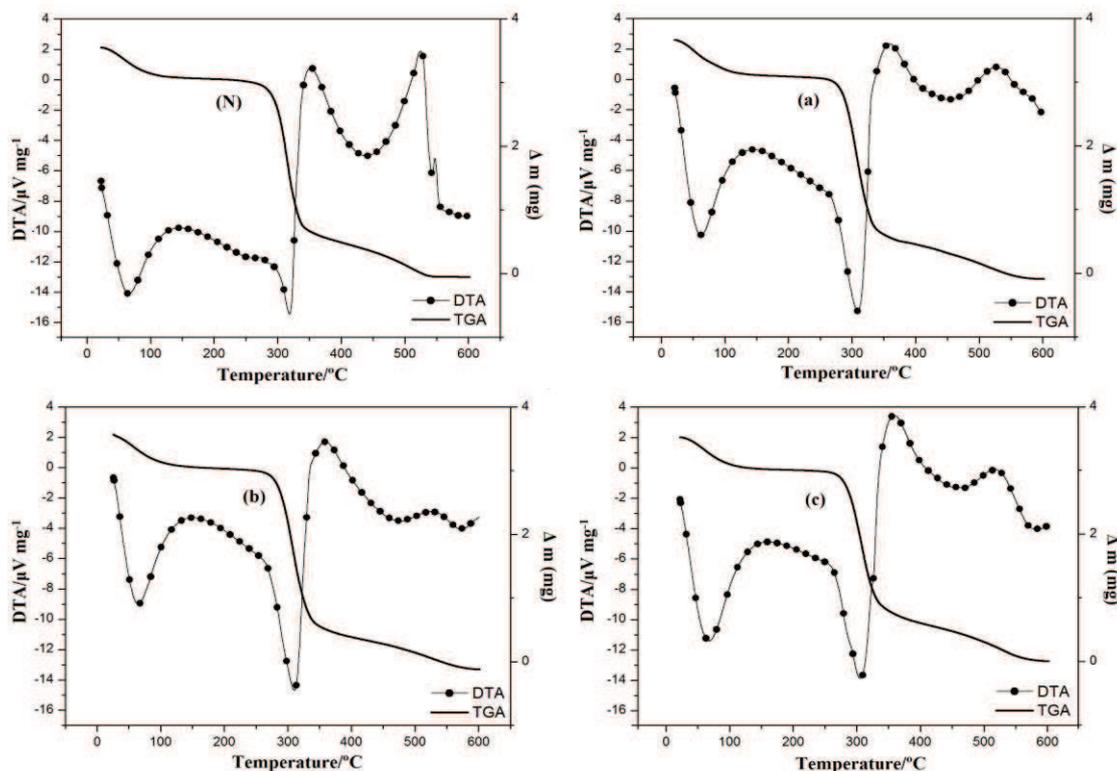
All the results were analysed for variance (ANOVA) with the Tukey test to compare sample means at 95% confidence level ( $p < 0.05$ ) using STATISTICA 7.0 software (StatSoft, Inc., Tulsa, OK, USA).

## Results and Discussion

### Simultaneous thermogravimetry and differential thermal analysis (TG–DTA)

The thermogravimetric curves (TG/DTA), Fig. 1, were performed and it was possible to verify that the profile of each curve were very similar showing three main events of mass loss. The first event, well marked with an endothermic peak in the DTA curve, is attributed to the evaporation of the water and volatile compounds, followed by stability. Once dehydrated, the second and third main regions in the TG curves were due to the degradation of the organic matter (amylose) and the formation of final residues (ash). Similar behavior were found in earlier works [1, 4]. The calculated results (TG/DTA) are shown in Table 1.

On Table 1, the first step correspond to the evaporation, which can be related to the moisture content of the samples. Thermal analysis permit the determination of moisture content in samples on a temperature range of 30 to 150 °C [11]. This fact was verified by Costa et al [1], that studied the characterization of native starches of seeds of *Araucaria angustifolia* from four germplasm collections, which were found similar values by both official method and thermogravimetric ones. The thermogravimetric analysis presents advantages in relation of the official method by requiring a much less amount of sample and it is faster could show the results in less than a half hour [1]. It can also be observed in Table 1, that the stability values increased significantly from the sample (N) to the modified ones, in a range of 209 - 243 °C, demonstrating that the modification by  $H_2O_2$ +UV caused alterations on the thermal properties of the *pinhão* starch.



**Fig.1** - TG/DTA curves: (N) native *pinhão* starch; (a; b; c) native *pinhão* starch treated with UV-rays ( $\lambda = 256$  nm) at one hour exposure and  $H_2O_2$  standard solutions 0.1; 0.2 and 0.5 mol L<sup>-1</sup>, respectively.

**Table 1** TG and DTA results of: (N) native *pinhão* starch; (a; b; c) native *pinhão* starch treated with UV-rays ( $\lambda = 256$  nm) at one hour exposure and  $H_2O_2$  standard solutions 0.1; 0.2 and 0.5 mol L<sup>-1</sup>, respectively.

Samples	TG Results		DTA Results	
	Step	$\Delta m/\%$	$\Delta T/^\circ C$	$T_p/^\circ C$
(N)	1 <sup>st</sup>	13.02	30 - 133	59.26
	stability	-	133 - 209	-
	2 <sup>nd</sup>	72.99	209 - 424	315.60
	3 <sup>rd</sup>	13.25	424 - 576	510.64
(a)	1 <sup>st</sup>	11.90	30 - 140	62.11
	stability	-	140 - 235	-
	2 <sup>nd</sup>	74.06	235 - 419	316.77
	3 <sup>rd</sup>	12.85	419 - 573	514.44
(b)	1 <sup>st</sup>	11.86	30 - 145	62.70
	stability	-	145 - 243	-
	2 <sup>nd</sup>	72.01	243 - 428	316.58
	3 <sup>rd</sup>	14.93	428 - 569	507.74
(c)	1 <sup>st</sup>	14.24	30 - 149	56.41
	stability	-	149 - 238	-
	2 <sup>nd</sup>	72.49	238 - 414	315.19
	3 <sup>rd</sup>	12.05	414 - 567	494.23

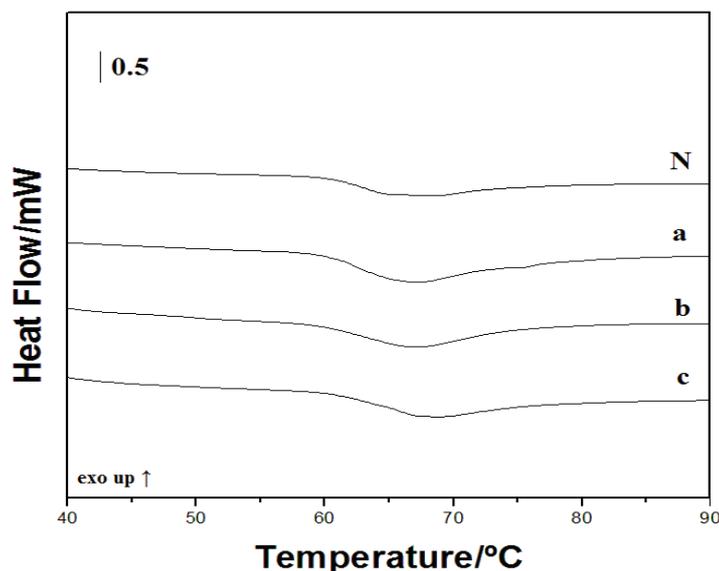
$\Delta m$  mass loss (%),  $\Delta T$  temperature range,  $T_p$  peak temperature.

### Differential scanning calorimetry (DSC)

DSC was used to determine de gelatinization properties of native and oxidized *pinhão* starch samples. The results are showed in Fig. 2 and Table 2.

As can be seen in Table 2, the oxidized samples (a, b and c) had a higher onset temperature ( $T_o$ ) than sample (N), however samples (a) and (b) did not differ significantly from sample (N) with respect to the peak temperature ( $T_p$ ), and only a higher  $H_2O_2$  concentration (sample c) increased the same. Final temperature ( $T_c$ ) decreased significantly when low concentrations of  $H_2O_2$  were used (samples a and b) and sample (c) (highest concentration of  $H_2O_2$ ) not differ significantly from sample (N). Sangseethong et al. [12] showed an increase in all transition temperatures ( $T_o$ ,  $T_p$  and  $T_c$ ) after oxidize cassava starch with

H<sub>2</sub>O<sub>2</sub> (3 %). Liu et al. [13] showed similar results after oxidize corn starch with H<sub>2</sub>O<sub>2</sub> (12 %). In this study, very low concentrations of H<sub>2</sub>O<sub>2</sub> were used, then some results are different from those found in the literature. Oxidation promoted a reduction in gelatinization enthalpy ( $\Delta H_{\text{gel}}$ ) and this reduction was more pronounced in higher concentrations of H<sub>2</sub>O<sub>2</sub> (samples a and b). Pinto et al. [2] and Klein et al. [14] found that gelatinization enthalpy of *pinhão* starch reduced after heat-moisture treatment. The reduction in  $\Delta H_{\text{gel}}$  can be attributed to degradation of the starch chains caused by modification, consequently, less energy was needed to gelatinize starch [12].



**Fig. 2** DSC gelatinization curves: (N) native *pinhão* starch; (a; b; c) native *pinhão* starch treated with UV-rays ( $\lambda = 256$  nm) at one hour exposure and H<sub>2</sub>O<sub>2</sub> standard solutions 0.1; 0.2 and 0.5 mol L<sup>-1</sup>, respectively.

**Table 2** DSC gelatinization of: (N) native *pinhão* starch; (a; b; c) native *pinhão* starch treated with UV-rays ( $\lambda = 256$  nm) at one hour exposure and H<sub>2</sub>O<sub>2</sub> standard solutions 0.1; 0.2 and 0.5 mol L<sup>-1</sup>, respectively.

Samples	DSC gelatinization			
	<i>To</i> /°C	<i>Tp</i> /°C	<i>Tc</i> /°C	$\Delta H_{\text{gel}}$ /J.g <sup>-1</sup>
(N)	59.08±0.04 <sup>a</sup>	66.02±0.05 <sup>a</sup>	75.54±0.03 <sup>c</sup>	14.64±0.02 <sup>d</sup>
(a)	60.41±0.23 <sup>b</sup>	66.49±0.09 <sup>a</sup>	70.96±0.18 <sup>a</sup>	6.75±0.04 <sup>c</sup>
(b)	60.78±0.22 <sup>b</sup>	66.74±0.06 <sup>a</sup>	74.50±0.22 <sup>b</sup>	5.85±0.01 <sup>b</sup>
(c)	62.08±0.05 <sup>c</sup>	67.80±0.07 <sup>c</sup>	75.14±0.11 <sup>c</sup>	5.24±0.06 <sup>b</sup>

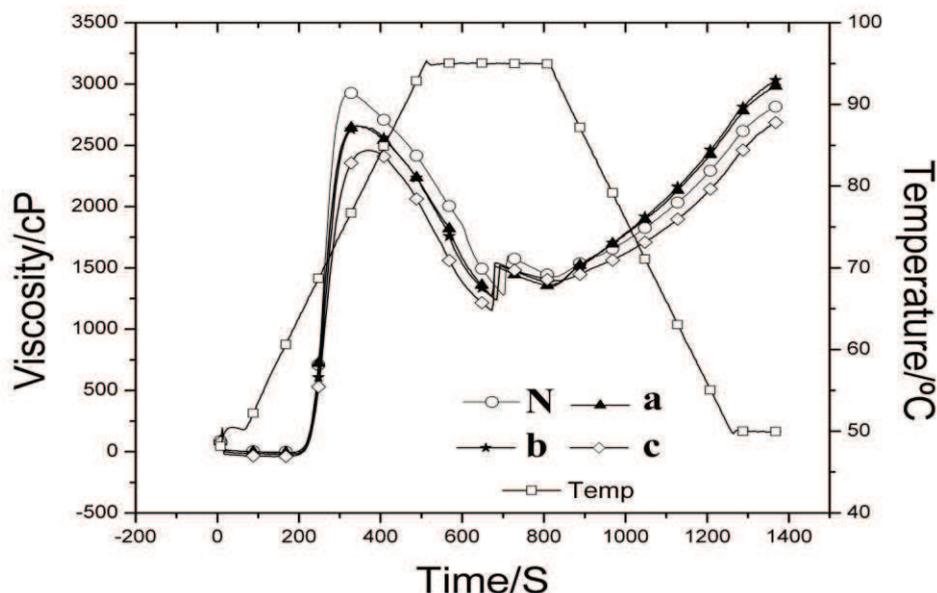
(\*) *To* “onset” initial temperature, *Tp* peak temperature, *Tc* “endset” final temperature,  $\Delta H_{\text{gel}}$  gelatinization enthalpy, (\*\*) different letters in the same column represents significative difference according to Duncan test ( $p < 0.05$ ).

### Rapid viscoamylographic analysis (RVA)

The RVA results are shown in Fig. 3 and Table 3. Oxidation caused a reduction in trough (hot paste viscosity) and viscosity peak, regardless of the concentration of peroxide used. Similar results were obtained by Pietrzyk et al. [15] to corn starch oxidized with 2 % and 4 % NaOCl and Gumul et al. [16] to potato starch oxidized with H<sub>2</sub>O<sub>2</sub> 2 %. The reduction in these viscosities can be attributed to degradation of the starch chains caused by oxidation, with consequent reduction of the molecular weight [17]. On the other hand, final viscosity of samples (a), (b) and (c) were higher than sample (N). During the heating step, amylose chains were solubilized forming a network of more compact structure in cooling step and increasing final viscosity [18].

The results of the pasting temperature, oxidation provided a reduction, except for the sample (b) that was not significantly different when compared with the sample (N). Similar results were obtained by Tethool, Jading and Santoso [19] after oxidation of sago starch using hydrogen peroxide (3 %) and catalyzed by UV irradiation.

Breakdown is an average resistance to degradation of the starch granule [20]. The oxidized samples (a), (b) e (c) had a reduction in breakdown indicating the higher tendency of starch to resist shear force. Setback reduced with the treatment that used hydrogen peroxide in a higher concentration (c), however in lower hydrogen peroxide concentrations occurred a increase of setback. Chon et al. [21] showed similar results. Higher setback value could be due higher amount of solubilized starch granules, which have a greater tendency to recouping during paste cooling [22].



**Fig. 3** RVA curves: (N) native *pinhão* starch; (a; b; c) native *pinhão* starch treated with UV-rays ( $\lambda = 256$  nm) at one hour exposure and  $H_2O_2$  standard solutions 0.1; 0.2 and 0.5 mol  $L^{-1}$ , respectively.

**Table 3** RVA results of: (N) native *pinhão* starch; (a; b; c) native *pinhão* starch treated with UV-rays ( $\lambda = 256$  nm) at one hour exposure and  $H_2O_2$  standard solutions 0.1; 0.2 and 0.5 mol  $L^{-1}$ , respectively.

Samples	Pasting temperature/ $^{\circ}C$	Trough/cP	Viscosity peak/cP	Setback/cP	Break/cP	Final viscosity/cP
(N)	65.55 $\pm$ 0.02 <sup>a</sup>	1426 $\pm$ 2.64 <sup>a</sup>	2925 $\pm$ 1.73 <sup>a</sup>	1398 $\pm$ 2.00 <sup>c</sup>	1499 $\pm$ 1.00 <sup>a</sup>	1426 $\pm$ 2.00 <sup>d</sup>
(a)	64.66 $\pm$ 0.01 <sup>c</sup>	1349 $\pm$ 2.64 <sup>d</sup>	2659 $\pm$ 2.64 <sup>b</sup>	1645 $\pm$ 3.60 <sup>b</sup>	1310 $\pm$ 2.00 <sup>c</sup>	2994 $\pm$ 1.73 <sup>b</sup>
(b)	65.55 $\pm$ 0.03 <sup>a</sup>	1362 $\pm$ 2.64 <sup>c</sup>	2659 $\pm$ 3.33 <sup>b</sup>	1680 $\pm$ 3.00 <sup>a</sup>	1297 $\pm$ 2.64 <sup>b</sup>	3042 $\pm$ 2.00 <sup>a</sup>
(c)	65.20 $\pm$ 0.02 <sup>b</sup>	1392 $\pm$ 1.73 <sup>b</sup>	2459 $\pm$ 1.00 <sup>c</sup>	1308 $\pm$ 2.00 <sup>d</sup>	1067 $\pm$ 3.00 <sup>d</sup>	2700 $\pm$ 2.64 <sup>c</sup>

(<sup>a</sup>) cP “centipoises”, sec “seconds”, (\*\*)(\*\*) different letters in the same column represents significative difference according to Duncan test ( $p < 0.05$ ).

## Conclusion

The TG-DTA analysis showed that the thermal properties of the *pinhão* starch was altered, highlighting the increased on the thermal stability event.

The DSC curves showed a similarity in their profiles related to the (N) sample, except for sample (c), and the calculated results showed a great decreased of the gelatinization enthalpy with the increasing of the  $H_2O_2$  concentrations solutions.

The RVA results presented that the oxidation caused a reduction in trough (hot paste viscosity) and viscosity peak of the modified samples related to unmodified one.

By the results obtained with the thermal analysis, it possible to conclude that the modification by  $H_2O_2$  and UV- rays exposure, altered the thermal behavior of the native *pinhão* starch.

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## References

- [1] Costa FJOG, Leivas CL, Waszczyński N, Godoi RCB, Helm CV, Colman TAD, Schnitzler E. Characterization of native starches of seeds of *Araucaria angustifolia* from four germplasm collection. *Thermochim. Acta.* 2013; 565:172-77.
- [2] Pinto NL, Vanier B, Klein ER, Zavareze MC, Elias LC, Gutkoski, Helbig E, Dias ARG. Physicochemical, crystallinity, pasting and termal properties of heat–moisture-treated pinhão starch. *Starch/Stärke.* 2012; 64:855–63.

- [3] Bello-Pérez LA, García-Suárez FJ, Méndez-Montealvo G, Do Nascimento JRO, Lajolo FM, Cordenunsi BR. Isolation and Characterization of Starch from Seeds of *Araucaria brasiliensis*: A Novel Starch for Application in Food Industry. *Starch/Stärke*. 2006; 58: 283–91.
- [4] Bicudo SCW, Demiate IM, Bannach G, Lacerda LG, Carvalho-Filho MAS, Ionashiro M, Schnitzler E. Thermoanalytical study and characterization of native starches of Paraná pine seeds (*Araucaria angustifolia*, Bert O. Ktze) and European chestnut seeds (*Castanea sativa* Mill). *Eclét. Quim*. 2009; 34:7-12.
- [5] Ribeiro LS, Cordoba LP, Colman TAD, Oliveira CS, Andrade MMP, Schnitzler E. Influence of some sugars on the thermal, rheological and morphological properties of “pinhão” starch. *J Therm Anal Calorim*. 2014; 117:935-42.
- [6] Thys RCS, Aires AG, Marczak LDF, Noreña CPZ. The effect of acid hydrolysis on the technological functional properties of pinhão (*Araucaria brasiliensis*) starch. *Food Sci Technol (Campinas)*. 2013; 33: 89-94.
- [7] Dias ARG, Zavareze R, Helbig E, Moura FA, Vargas CG, Ciacco CF. Oxidation of fermented cassava starch using hydrogen peroxide. *Carbohydr Polym*. 2011; 86:185-91.
- [8] Beninca C, Colman TAD, Lacerda LG, Carvalho-Filho MAS, Demiate IM, Schnitzler E. Thermal, rheological, and structural behaviors of natural and modified cassava starch granules, with hypochlorite solutions. *J Therm Anal Calorim*. 2013; 111:2217-22.
- [9] Colman TAD, Demiate IM, Schnitzler E. The effect of microwave radiation on some thermal, rheological and structural properties of cassava starch. *J Therm Anal Calorim*. 2014; 115:2245-52.
- [10] Andrade MMP, Oliveira CS, Colman TAD, Costa FJOG, Schnitzler E. Effects of heat-moisture treatment on organic cassava starch. *J. Therm. Anal. Calorim*. 2014; 115:2115-22.
- [11] Soliman AAA, El-Shinnawy A, Mobarak F. Thermal behavior of starch and oxidized starch. *Thermochim. Acta*. 1997; 296:149-53.
- [12] Sangseethong K, Termvejsayanon N, Sriroth K. Characterization of physicochemical properties of hypochlorite- and peroxide-oxidized cassava starches. *Carbohydr Polym*. 2010; 82:446-53.
- [13] Liu J, Wang B, Lin L, Zhang J, Liu W, Xie J, Ding Y. Functional, physicochemical properties and structure of cross-linked oxidized maize starch. *Food Hydrocolloid*. 2014; 36:45-52.
- [14] Klein B, Pinto VZ, Vanier NL, Zavareze ER, Colussi R, Evangelho JA, Gutkoski LC, Dias ARG. Effect of single and dual heat-moisture treatments on properties of rice, cassava, and pinhao starches. *Carbohydr Polym*. 2013; 98:1578-84.
- [15] Pietrzyk S, Juszczak L, Fortuna T, Ciemniowska A. Effect of the oxidation level of corn starch on its acetylation and physicochemical and rheological properties. *J Food Eng*. 2014; 120:50–6.
- [16] Gumul D, Krystyan M, Buksa K, Ziobro R, Zięba T. The influence of oxidation, extrusion and oxidation/extrusion on physico-chemical properties of potato starch. *Starch/Stärke*. 2014; 60:190–98.
- [17] Kuakpetoon D, Wang YJ. Characterization of different starches oxidized by hypochlorite. *Starch/Stärke*. 2001; 53:211–18.
- [18] Sanchez-Rivera MM., Garcia-Suarez FJL, Valle VM., Gutierrez-Meraz F, Bello-Perez, LA. Partial Characterization of banana starches oxidized by different levels of sodium hypochlorite. *Carbohydr Polym*. 2005; 62:50-6.
- [19] Tethool EF, Jading A, Santoso B. Characterization of Physicochemical and Baking Expansion Properties of Oxidized Sago Starch Using Hydrogen Peroxide and Sodium Hypochlorite Catalyzed By UV Irradiation. *Food Sci Qualit Manag*. 2012; 10:1-11.
- [20] Sandhu KS, Singh N. Some properties of corn starches II: physicochemical, gelatinization, retrogradation, pasting and gel textural properties. *Food Chem*. 2007; 101:1499-1507.
- [21] Chong W, Uthumporn U, Karim A, Cheng L. The influence of ultrasound on the degree of oxidation of hypochlorite-oxidized corn starch. *LWT – Food Sci. Technol*. 2012; 50:439:43.
- [22] Chan HT, Bhat R, Karim AA. Effects of sodium dodecyl sulphate and sonication treatment on physicochemical properties of starch. *Food Chem*. 2010; 120:703-09.